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#### (57) Abstract

The sequences of nucleic acids encoding proteins required for *E. coli* proliferation are disclosed. The nucleic acids can be used to express proteins or portions thereof, to obtain antibodies capable of specifically binding to the expressed proteins, and to use those expressed proteins as a screen to isolate candidate molecules for rational drug discovery programs. The nucleic acids can also be used to screen for homologous genes that are required for proliferation in microorganisms other than *E. coli*. The nucleic acids can also be used to design expression vectors and secretion vectors. The nucleic acids of the present invention can also be used in various assay systems to screen for proliferation required genes in other organisms as well as to screen for antimicrobial agents.

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#### ESCHERICHIA COLI

#### BACKGROUND OF THE INVENTION

Since the discovery of penicillin, the use of antibiotics to treat the ravages of bacterial infections has saved millions of lives. With the advent of these "miracle drugs," for a time it was popularly believed that humanity might, once and for all, be saved from the scourge of bacterial infections. In fact, during the 1980s and early 1990s, many large pharmaceutical companies cut back or eliminated antibiotics research and development. They believed that infectious disease caused by bacteria finally had been conquered and that markets for new drugs were limited. Unfortunately, this belief was overly optimistic.

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The tide is beginning to turn in favor of the bacteria as reports of drug resistant bacteria become more frequent. The United States Centers for Disease Control announced that one of the most powerful known antibiotics, vancomycin, was unable to treat an infection of the common Staphylococcus aureus (staph). This organism is commonly found in our environment and is responsible for many nosocomial infections. The import of this announcement becomes clear when one considers that vancomycin was used for years to treat infections caused by stubborn strains of bacteria, like staph. In short, the bacteria are becoming resistant to our most powerful antibiotics. If this trend continues, it is conceivable that we will return to a time when what are presently considered minor bacterial infections are fatal diseases.

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There are a number of causes for the predicament in which practitioners of medical arts find themselves. Overprescription and improper prescription habits by some physicians have caused an indiscriminate increase in the availability of antibiotics to the public. The patient is also partly responsible, for even in instances where an antibiotic is the appropriate treatment, patients will often improperly use the drug, the result being yet another population of bacteria that is resistant, in whole or in part, to traditional antibiotics.

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The bacterial scourges that have haunted humanity remain, in spite of the development of modern scientific practices to deal with the diseases that they cause. Drug resistant bacteria are now advancing on the health of humanity. A new generation of antibiotics to once again deal with the pending health threat that bacteria present is required.

#### Discovery of New Antibiotics

As more and more bacterial strains become resistant to the panel of available antibiotics, new compounds are required. In the past, practitioners of pharmacology would have to rely upon traditional methods of drug discovery to generate novel, safe and efficacious compounds for the treatment of disease. Traditional drug discovery methods involve blindly testing potential drug candidate-molecules, often selected at random, in the hope that one might prove to be an effective treatment for some disease. The process is painstaking and laborious, with no guarantee of success. Today, the average cost to discover and develop a new drug is nearly US \$500 million, and the average time is 15 years from laboratory to patient. Improving this process, even incrementally, would represent a huge advance in the generation of novel antimicrobial agents.

Newly- emerging practices in drug discovery utilize a number of biochemical techniques to provide for directed approaches to creating new drugs, rather than discovering them at random. For example, gene sequences and proteins encoded thereby that are required for the proliferation of an organism make for excellent targets since exposure of bacteria to compounds active against these targets would result in the inactivation of the organism. Once a target is identified, biochemical analysis of that target can be used to discover or to design molecules that interact with and alter the functions of the target. Using physical and computational techniques, to analyze structural and biochemical targets in order to derive compounds that interact with a target is called rational drug design and offers great future potential. Thus, emerging drug discovery practices use molecular modeling techniques, combinatorial chemistry approaches, and other means to produce and screen and/or design large numbers of candidate compounds.

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Nevertheless, while this approach to drug discovery is clearly the way of the future, problems remain. For example, the initial step of identifying molecular targets for investigation can be an extremely time consuming task. It may also be difficult to design molecules that interact with the target by using computer modeling techniques. Furthermore, in cases where the function of the target is not known or is poorly understood, it may be difficult to design assays to detect molecules that interact with and alter the functions of the target. To improve the rate of novel drug discovery and development, methods of identifying important molecular targets in pathogenic microorganisms and methods for identifying molecules that interact with and alter the functions of such molecular targets are urgently required.

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Escherichia coli represents an excellent model system to understand bacterial biochemistry and physiology. The estimated 4288 genes scattered along the 4.6 x 10<sup>6</sup> base pairs of the Escherichia coli (E. coli) chromosome offer tremendous promise for the understanding of bacterial biochemical processes. In turn, this knowledge will assist in the development of new tools for the diagnosis and treatment of bacteria-caused human disease. The entire E. coli genome has been sequenced, and this body of information holds a tremendous potential for application to the discovery and development of new antibiotic compounds. Yet, in spite of this accomplishment, the general functions or roles of many of these genes are still unknown. For example, the total number of proliferation-required genes contained within the E. coli genome is unknown, but has been variously estimated at around 200 to 700 (Armstrong, K.A. and Fan, D.P. Essential Genes in the metB-malB Region of Escherichia coli K12, 1975, J. Bacteriol. 126: 48-55).

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Novel, safe and effective antimicrobial compounds are needed in view of the rapid rise of antibiotic resistant microorganisms. However, prior to this invention, the characterization of even a single bacterial gene was a painstaking process, requiring years of effort. Accordingly, there is an urgent need for more novel methods to identify and characterize bacterial genomic sequences that encode gene products required for proliferation and for methods to identify molecules that interact with and alter the functions of such genes and gene products.

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#### SUMMARY OF THE INVENTION

One embodiment of the present invention is a purified or isolated nucleic acid sequence consisting essentially of one of SEQ ID NOs: 1-81, 405-485, wherein said nucleic acid inhibits microorganism proliferation. The nucleic acid sequence may be complementary to at least a portion of a coding sequence of a gene whose expression is required for

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microorganism proliferation. The nucleic acid sequence may comprise a fragment of one of SEQ ID NOs. 1-81, 405-485, said fragment selected from the group consisting of fragments comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEQ ID NOs: 1-81, 405-485. The nucleic acid sequence may be complementary to a coding sequence of a gene whose expression is required for microorganism proliferation.

Another embodiment of the present invention is a vector comprising a promoter operably linked to a nucleic acid comprising a sequence selected from the group consisting of SEQ ID NOs. 1-81, 405-485. The promoter may be active in an organism selected from the group consisting of Escherichia coli, Staphylococcus aureus, Pseudomonas aeruginosa, Enterobacter cloacae, Helicobacter pylori, Neisseria gonorrhoeae, Enterococcus faecalis, Streptococcus pneumoniae, Haemophilus influenzae, Salmonella typhimurium, Saccharomyces cerevisiae, Candida albicans, Cryptococcus neoformans, Aspergillus fumigatus, Klebsiella pneumoniae, Salmonella typhi, Salmonella paratyphi, Salmonella cholerasuis, Staphylococcus epidermidis, Mycobacterium tuberculosis, Mycobacterium leprae, Treponema pallidum, Bacillus anthracis, Yersinia pestis, Clostridium botulinum, campylobacter jejuni, Chlamydia trachomatus, Chlamydia pneumoniae or any species falling within the genera of any of the above species.

Another embodiment of the present invention is a host cell containing the vectors described above.

Another embodiment of the present invention is a purified or isolated nucleic acid consisting essentially of the coding sequence of one of SEO ID NOs: 82-88, 90-242. One aspect of this embodiment is a fragment of the nucleic acid comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEO ID NOs: 82-88, 90-242.

Another embodiment of the present invention is a vector comprising a promoter operably linked to the nucleic acids of the preceding embodiment.

Another aspect of the present invention is a purified or isolated nucleic acid comprising a nucleic acid sequence complementary to at least a portion of an intragenic sequence, intergenic sequence, sequences spanning at least a portion of two or more genes, 5' noncoding region, or 3' noncoding region within an operon encoding a polypeptide comprising a sequence selected from the group consisting of SEO ID NOs: 243-357, 359-398.

Another embodiment of the present invention is a purified or isolated nucleic acid comprising a nucleic acid having at least 70% homology to a sequence selected from the group consisting of SEQ ID NOs 1-81, 405-485, 82-88, 90-242 or the sequences complementary thereto as determined using BLASTN version 2.0 with the default parameters. The nucleic acid may be from an organism selected from the group consisting of Staphylococcus aureus, Pseudomonas aeruginosa, Enterobacter cloacae, Helicobacter pylori, Neisseria gonorrhoeae, Enterococcus faecalis, Streptococcus pneumoniae, Haemophilus influenzae, Salmonella typhimurium, Saccharomyces cerevisiae, Candida albicans, Cryptococcus neoformans, Aspergillus fumigatus, Klebsiella pneumoniae, Salmonella typhi, Salmonella paratyphi, Salmonella cholerasuis, Staphylococcus epidermidis, Mycobacterium tuberculosis, Mycobacterium leprae, Treponema pallidum, Bacillus anthracis, Yersinia pestis, Clostridium botulinum, campylobacter jejuni, and Chlamydia trachomatus, Chlamydia pneumoniae or any species falling within the genera of any of the above species.



Another embodiment of the present invention is a purified or isolated nucleic acid consisting essentially of a nucleic acid encoding a polypeptide having a sequence selected from the group consisting of SEQ ID NOs.: 243-357, 359-398.

Another embodiment of the present invention is a vector comprising a promoter operably linked to a nucleic acid encoding a polypeptide having a sequence selected from the group consisting of SEQ ID NOs.: 243-357, 359-398.

Another embodiment of the present invention is a host cell containing the vector of the preceding embodiment.

Another embodiment of the present invention is purified or isolated polypeptide comprising the sequence of one of SEO ID NOs: 243-357, 359-398.

Another embodiment of the present invention is purified or isolated polypeptide comprising a fragment of one of the polypeptides of SEO ID NOs. 243-357, 359-398, said fragment selected from the group consisting of fragments comprising at least 5, at least 10, at least 20, at least 30, at least 40, at least 50, at least 60 or more than 60 consecutive amino acids of one of the polypeptides of SEO ID NOs.: 243-357, 359-398.

Another embodiment of the present invention is an antibody capable of specifically binding the polypeptide of the preceding embodiment.

Another embodiment of the present invention is method of producing a polypeptide, comprising introducing a vector comprising a promoter operably linked to a nucleic acid encoding a polypeptide having a sequence selected from the group consisting of SEQ ID NOs. 243-357, 359-398into a cell. The method may further comprise the step of isolating said protein.

Another embodiment of the present invention is a method of inhibiting proliferation comprising inhibiting the activity or reducing the amount of a polypeptide having a sequence selected from the group consisting of SEO ID NOs. 243-357, 359-398 or inhibiting the activity or reducing the amount of a nucleic acid encoding said polypeptide.

Another embodiment of the present invention is method for identifying compounds which influence the activity of a polypeptide required for proliferation comprising:

contacting a polypeptide comprising a sequence selected from the group consisting of 243-357, 359-398with a candidate compound; and

determining whether said compound influences the activity of said polypeptide.

The activity may be an enzymatic activity. The activity may be a carbon compound catabolism activity. The activity may be a biosynthetic activity. The activity may be a transporter activity. The activity may be a transcriptional activity. The activity may be a DNA replication activity. The activity may be a cell division activity.

Another embodiment of the present invention is a compound identified using the above method.

Another embodiment of the present invention is method for assaying compounds for the ability to reduce the activity or level of a polypeptide required for proliferation, comprising:

providing a target, wherein said target comprises the coding sequence of a sequence selected from the group consisting of SEO ID NOs. 82-88, 90-242:

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contacting savarget with a candidate compound; and measuring an activity of said target.

The target may be a messenger RNA molecule transcribed from a coding region of one of SEQ ID. NOs.: 82-88, 90-242 and said activity is translation of said messenger RNA. The target may be a coding region of one of SEQ ID. NOs. 82-88, 90-242 and said activity is transcription of said messenger RNA.

Another embodiment of the present invention is a compound identified using the method above.

Another embodiment of the present invention is a method for identifying compounds which reduce the activity or level of a gene product required for cell proliferation comprising the steps of:

expressing an antisense nucleic acid against a nucleic acid encoding said gene product in a cell to reduce the activity or amount of said gene product in said cell, thereby producing a sensitized cell;

contacting said sensitized cell with a compound; and

determining whether said compound inhibits the growth of said sensitized cell to a greater extent than said compound inhibits the growth of a nonsensitized cell.

The cell may be selected from the group consisting of bacterial cells, fungal cells, plant cells, and animal cells. The cell may be an *E. coli* cell. The cell may be from an organism selected from the group consisting of *Staphylococcus aureus, Pseudomonas aeruginosa, Enterobacter cloacae, Helicobacter pylori, Neisseria gonorrhoeae, Enterococcus faecalis, Streptococcus pneumoniae, Haemophilus influenzae, Salmonella typhimurium, Saccharomyces cerevisiae, Candida albicans, Cryptococcus neoformans, Aspergillus fumigatus, Klebsiella pneumoniae, Salmonella typhi, Salmonella paratyphi, Salmonella cholerasuis, Staphylococcus epidermidis, Mycobacterium tuberculosis, Mycobacterium leprae, Treponema pallidum, Bacillus anthracis, Yersinia pestis, Clostridium botulinum, campylobacter jejuni, and Chlamydia trachomatus, Chlamydia pneumoniae or any species falling within the genera of any of the above species. The antisense nucleic acid may be transcribed from an inducible promoter. The method may, further comprise the step of contacting said cell with a concentration of inducer which induces said antisense nucleic acid to a sublethal level. The sub-lethal concentration of said inducer may be such that growth inhibition is 8% or more. The inducer may be isopropyl-1-thio-β-D-galactoside. The growth inhibition may be measured by monitoring optical density of a culture growth solution. The gene product may be a polypeptide. The gene product may be an RNA. The gene product may comprise a polypeptide having a sequence selected from the group consisting of SEO ID NOs.: 243-357, 359-398.* 

Another embodiment of the present invention is a compound identified using the method above.

Another embodiment of the present invention is a method for inhibiting cellular proliferation comprising introducing a compound with activity against a gene corresponding to one of SEQ ID NOs.: 82-88, 90-242 or with activity against the product of said gene into a population of cells expressing a gene. The compound may be an antisense oligonucleotide comprising a sequence selected from the group consisting of SEQ ID NOs.: 1-81, 405-485, or a proliferation-inhibiting portion thereof. The proliferation inhibiting portion of one of SEQ ID NOs. 1-81, 405-485

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may be a fragment comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEQ ID NOs: 1-81, 405-485. The compound may be a triple helix oligonucleotide.

Another embodiment of the present invention is a preparation comprising an effective concentration of an antisense oligonucleotide comprising a sequence selected from the group consisting of SEQ ID NOs.: 1-81, 405-485, or a proliferation-inhibiting portion thereof in a pharmaceutically acceptable carrier. The proliferation-inhibiting portion of one of SEQ ID NOs. 1-81, 405-485 may comprise at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEQ ID NOs: 1-81, 405-485.

Another embodiment of the present invention is a method for inhibiting the expression of a gene in an operon required for proliferation comprising contacting a cell in a cell population with an antisense nucleic acid, said cell expressing a gene corresponding to one of SEQ ID NOs.: 82-88, 90-242, wherein said antisense nucleic acid comprises at least a proliferation-inhibiting portion of said operon in an antisense orientation that is effective in inhibiting expression of said gene. The antisense nucleic acid may be complementary to a sequence of a gene comprising one or more of SEQ ID NOs.: 82-88, 90-242. The antisense nucleic acid may be a sequence of one of SEQ ID NOs.: 1-81, 405-485, or a portion thereof. The cell may be contacted with said antisense nucleic acid by introducing a plasmid which expresses said antisense nucleic acid into said cell population. The cell may be contacted with said antisense nucleic acid by introducing a phage which expresses said antisense nucleic acid into said cell population. The cell may be contacted with said antisense nucleic acid by introducing a sequence encoding said antisense nucleic acid into the chromosome of said cell into said cell population. The cell may be contacted with said antisense nucleic acid by introducing a retron which expresses said antisense nucleic acid into said cell population. The cell may be contacted with said antisense nucleic acid by introducing a ribozyme into said cell-population, wherein a binding portion of said ribozyme is complementary to said antisense oligonucleotide. The cell may be contacted with said antisense nucleic acid by introducing a liposome comprising said antisense oligonucleotide into said cell. The cell may be contacted with said antisense nucleic acid by electroporation. The antisense nucleic acid may be a fragment comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEO ID NOs: 82-88, 90-242. The antisense nucleic acid may be an oligonucleotide.

Another embodiment of the present invention is a method for identifying bacterial strains comprising the steps of:

providing a sample containing a bacterial species; and

identifying a bacterial species using a species specific probe having a sequence selected from the group consisting of SEQ ID NOs. 1-81, 405-485, 82-88, 90-242.

Another embodiment of the present invention is a method for identifying a gene in a microorganism required for proliferation comprising:

- (a) identifying an inhibitory nucleic acid which inhibits the activity of a gene or gene product required for proliferation in a first microorganism;
- (b) contacting a second microorganism with said inhibitory nucleic acid;

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- (c) determining whether said inhibitory nucleic acid from said first microorganism inhibits proliferation of said second microorganism; and
- (d) identifying the gene in said second microorganism which is inhibited by said inhibitory nucleic acid.

Another embodiment of the present invention is a method for assaying a compound for the ability to inhibit proliferation of a microorganism comprising:

- (a) identifying a gene or gene product required for proliferation in a first microorganism;
- (b) identifying a homolog of said gene or gene product in a second microorganism;
- (c) identifying an inhibitory nucleic acid sequence which inhibits the activity of said homolog in said second microorgansim;
- (d) contacting said second microorganism with a proliferation-inhibiting amount of said inhibitory nucleic acid, thus sensitizing said second microorganism;
  - (e) contacting the sensitized microorganism of step (d) with a compound; and
  - (f) determining whether said compound inhibits proliferation of said sensitized microorganism to a greater extent than said compound inhibits proliferation of a nonsensitized microorganism.
  - The step of identifying a gene involved in proliferation in a first microorganism may comprise:

introducing a nucleic acid comprising a random genomic fragment from said first microorganism operably linked to a promoter wherein said random genomic fragment is in the antisense orientation; and

comparing the proliferation of said first microorganism transcribing a first level of said random genomic fragment to the proliferation of said first microorganism transcribing a lower level of said random genomic fragment, wherein a difference in proliferation indicates that said random genomic fragment comprises a gene involved in proliferation.

The step of identifying a homolog of said gene in a second microorganism may comprise identifying a homologous nucleic acid or a nucleic acid encoding a homologous polypeptide in a database using an algorithm selected from the group consisting of BLASTN version 2.0 with the default parameters and FASTA version 3.0t78 algorithm with the default parameters. The step of identifying a homolog of said gene in a second microorganism may comprise identifying a homologous nucleic acid or a nucleic acid encoding a homologous polypeptide by identifying nucleic acids which hybridize to said first gene. The step of identifying a homolog of said gene in a second microorganism may comprise expressing a nucleic acid which inhibits the proliferation of said first microorganism in said second microorganism. The inhibitory nucleic acid may be an antisense nucleic acid. The inhibitory nucleic acid may comprise an antisense nucleic acid to a portion of said homolog. The inhibitory nucleic acid may comprise an antisense nucleic acid to a portion of the operon encoding said homolog. The step of contacting the second microorganism with a proliferation inhibiting amount of said nucleic acid sequence may comprise directly contacting said second microorganism with said nucleic acid sequence may comprise expressing an antisense nucleic acid to said homolog in said second microorganism.

Another embodiment of the present invention is a compound identified using the method above.

Another embodiment of the present invention is a method of assaying a compound for the ability to inhibit proliferation comprising:

- (a) identifying an inhibitory nucleic acid sequence which inhibits the activity of a gene or gene product required for proliferation in a first microorgansim;
- (b) contacting a second microorganism with a proliferation-inhibiting amount of said inhibitory nucleic acid, thus sensitizing said second microorganism;
- (c) contacting the proliferation-inhibited microorganism of step (b) with a compound; and
- (d) determining whether said compound inhibits proliferation of said sensitized second microorganism to a greater extent than said compound inhibits proliferation of a nonsensitized second microorganism.

The inhibitory nucleic acid may be an antisense nucleic acid which inhibits the proliferation of said first microorganism. The inhibitory nucleic acid may comprise a portion of an antisense nucleic acid which inhibits the proliferation of said first microorganism. The inhibitory nucleic acid may comprise an antisense molecule against the entire coding region of the gene involved in proliferation of the first microorganism. The inhibitory nucleic acid may comprise an antisense nucleic acid to a portion of the operon encoding the gene involved in proliferation of the first microorganism.

Another embodiment of the present invention is a compound identified using the method above.

Another embodiment of the present invention is a method for assaying compounds for activity against a biological pathway required for proliferation comprising:

sensitizing a cell by expressing an antisense nucleic acid against a nucleic acid encoding a gene product required for proliferation in a cell to reduce the activity or amount of said gene product;

contacting the sensitized cell with a compound; and

determining whether said compound inhibits the growth of said sensitized cell to a greater extent than said compound inhibits the growth of an nonsensitized cell.

The cell may be selected from the group consisting of bacterial cells, fungal cells, plant cells, and animal cells. The cell may be an *E. coli* cell. The cell may be an organism selected from the group consisting of *Staphylococcus aureus, Pseudomonas aeruginosa, Enterobacter cloacae, Helicobacter pylori, Neisseria gonorrhoeae, Enterococcus faecalis, Streptococcus pneumoniae, Haemophilus influenzae, Salmonella typhimurium, Saccharomyces cerevisiae, Candida albicans, Cryptococcus neoformans, Aspergillus fumigatus, Klebsiella pneumoniae, Salmonella typhi, Salmonella paratyphi, Salmonella cholerasuis, Staphylococcus epidermidis, Mycobacterium tuberculosis, Mycobacterium leprae, Treponema pallidum, Bacillus anthracis, Yersinia pestis, Clostridium botulinum, campylobacter jejuni, and Chlamydia trachomatus, Chlamydia pneumoniae* or any species falling within the genera of any of the above species. The antisense nucleic acid may be transcribed from an inducible promoter. The method may further comprise contacting the cell with an agent which induces expression of said antisense nucleic acid from said inducible promoter, wherein said antisense nucleic acid is expressed at a sublethal level. The sublethal level of said antisense nucleic acid

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may inhibit proliferation of 8% or more. The agent may be isopropyl-1-thio- $\beta$ -D-galactoside (IPTG). The inhibition of proliferation may be measured by monitoring the optical density of a liquid culture. The gene product may comprise a polypeptide having a sequence selected from the group consisting of SEQ ID NOs: 243-357, 359-398.

Another embodiment of the present invention is a compound identified using the method above.

Another embodiment of the present invention is a method for assaying a compound for the ability to inhibit cellular proliferation comprising:

contacting a cell with an agent which reduces the activity or level of a gene product required for proliferation of said cell;

contacting said cell with said compound; and

determining whether said compound reduces proliferation to a greater extent than said compound reduces proliferation of cells which have not been contacted with said agent.

The agent which reduces the activity or level of a gene product required for proliferation of said cell may comprise an antisense nucleic acid to a gene or operon required for proliferation. The agent which reduces the activity or level of a gene product required for proliferation of said cell may comprise an antibiotic. The cell may contain a temperature sensitive mutation which reduces the activity or level of said gene product required for proliferation of said cell. The antisense nucleic acid may be directed against the same functional domain of said gene product required for proliferation of said cell to which said antisense nucleic acid is directed. The antisense nucleic acid may be directed against a different functional domain of said gene product required for proliferation of said cell than the functional domain to which said antisense nucleic acid is directed.

Another embodiment of the present invention is a compound identified using the method above.

Another embodiment of the present invention is a method for identifying the pathway in which a proliferation-required nucleic acid or its gene product lies comprising:

expressing a sublethal level of an antisense nucleic acid directed against said proliferation-required nucleic acid in a cell;

contacting said cell with an antibiotic, wherein the a biological pathway on which said antibiotic acts is known; and

determining whether said cell has a substantially greater sensitivity to said antibiotic than a cell which does not express said sublethal level of said antisense nucleic acid.

Another embodiment of the present invention is a method for determining the pathway on which a test compound acts comprising:

- (a) expressing a sublethal level of an antisense nucleic acid directed against a proliferation-required nucleic acid in a cell, wherein the biological pathway in which said proliferation-required nucleic acid lies is known,
  - (b) contacting said cell with said test compound; and
- (c) determining whether said cell has a substantially greater sensitivity to said test compound than a cell which does not express said sublethal level of said antisense nucleic acid.

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The method may further comprise:

(d) expressing a sublethal level of a second antisense nucleic acid directed against a second proliferation-required nucleic acid in said cell, wherein said second proliferation-required nucleic acid is in a different biological pathway than said proliferation-required nucleic acid in step (a); and

(e) determining whether said cell has a substantially greater sensitivity to said test compound than a cell which does not express said sublethal level of said second antisense nucleic acid.

Another embodiment of the present invention is a purified or isolated nucleic acid consisting essentially of one of SEQ ID NOs: 358, 399-402.

Another embodiment of the present invention is a purified or isolated nucleic acid comprising a sequence selected from the group consisting of 1-81, 405-485, 82-88, 90-242, 358, 399-402.

Another embodiment of the present invention is a compound which interacts with the gene or gene product of a nucleic acid comprising a sequence of one of SEQ ID NOs: 82-88, 90-242 to inhibit proliferation.

Another embodiment of the present invention compound which interacts with a polypeptide comprising one of SECIID NOs. 243-357, 359-398 to inhibit proliferation.

Another embodiment of the present invention is a compound which interacts with a nucleic acid comprising one of SEQ ID NOs: 358, 399-402 to inhibit proliferation.

#### BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is an IPTG dose response curve in *E. coli* transformed with an IPTG-inducible plasmid containing either an antisense clone to the *E. coli* ribosomal protein rpIW (AS-rpIW) which is required for protein synthesis and essential cell proliferation, or an antisense clone to the elaD (AS-elaD) gene which is not known to be involved in protein synthesis and which is also essential for proliferation.

Figure 2A is a tetracycline dose response curve in E. coli transformed with an IPTG-inducible plasmid containing antisense to rpIW(AS-rpIW) in the presence of 0, 20 or 50  $\mu$ M IPTG.

Figure 2B is a tetracycline dose response curve in  $\it E.~coli$  transformed with an IPTG-inducible plasmid containing antisense to elaD (AS-elaD) in the presence of 0, 20 or 50  $\mu$ M IPTG.

Figure 3 is a graph showing the fold increase in tetracycline sensitivity of *E. coli* transfected with antisense clones to essential ribosomal proteins L23 (AS-rpIW) and L7/L12 and L10 (AS-rpILrpIJ). Antisense clones to genes known not to be involved in protein synthesis (atpB/E(AS-atpB/E)), visC (AS-visC, elaD (AS-elaD), yohH (AS-yohH) are much less sensitive to tetracycline.

#### **Definitions**

By "biological pathway" is meant any discrete cell function or process that is carried out by a gene product or a subset of gene products. Biological pathways include enzymatic, biochemical and metabolic pathways as well as pathways involved in the production of cellular structures such cell walls. Biological pathways that are usually required for proliferation of microorganisms include, but are not limited to, cell division, DNA synthesis & replication,

RNA synthesis (transcription), protein synthesis (translation), protein processing, protein transport, fatty acid biosynthesis, cell wall synthesis, cell membrane synthesis & maintenance, etc.

By "inhibit activity against a gene or gene product" is meant having the ability to interfere with the function of a gene or gene product in such a way as to decrease expression of the gene or to reduce the level or activity of a product of the gene. Agents which have activity against a gene include agents that inhibit transcription of the gene and agents that inhibit translation of the mRNA transcribed from the gene. In microorganisms, agents which have activity against a gene can act to decrease expression of the operon in which the gene resides or alter the processing of operon RNA such as to reduce the level or activity of the gene product. The gene product can be a non-translated RNA such as ribosomal RNA, a translated RNA (mRNA) or the protein product resulting from translation of the gene mRNA: Of particular utility to the present invention are anti-sense RNAs that have activities against the operons or genes to which they specifically hybridze.

By "activity against a gene product" is meant having the ability to inhibit the function or to reduce the level or activity of the gene product in a cell.

By "activity against a protein" is meant having the ability to inhibit the function or to reduce the level or activity of the protein in a cell.

By "activity against nucleic acid" is meant having the ability to inhibit the function or to reduce the level or activity of the nucleic acid in a cell.

As used herein, "sublethal" means a concentration of an agent below the concentration required to inhibit all cell growth.

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#### **DETAILED DESCRIPTION OF THE INVENTION**

The present invention describes a group of *E. coli* genes and gene families required for growth and/or proliferation. A proliferation-required gene or gene family is one where, in the absence of a gene transcript and/or gene product, growth or viability of the microorganism is reduced or eliminated. Thus, as used herein the terminology "proliferation-required" or "required for proliferation" encompasses sequences where the absence of a gene transcript and/or gene product completely eliminates cell growth as well as sequences where the absence of a gene transcript and/or gene product merely reduces cell growth. These proliferation-required genes can be used as potential targets for the generation of new antimicrobial agents. To achieve that goal, the present invention also encompasses novel assays for analyzing proliferation-required genes and for identifying compounds which interact with the gene products of the proliferation-required genes. In addition, the present invention contemplates the expression of genes and the purification of the proteins encoded by the nucleic acid sequences identified as required proliferation genes and reported herein. The purified proteins can be used to generate reagents and screen small molecule libraries or other candidate compound libraries for compounds that can be further developed to yield novel antimicrobial compounds. The present invention also describes methods for identification of homologous genes in organisms other than *E. coli*.

The present invention utilizes a novel method to identify proliferation-required *E. coli* sequences. Generally, a library of nucleic acid sequences from a given source are subcloned or otherwise inserted into an inducible expression

vector, thus forming an expression library. Although the insert nucleic acids may be derived from the chromosome of the organism into which the expression vector is to be introduced, because the insert is not in its natural chromosomal location, the insert nucleic acid is an exogenous nucleic acid for the purposes of the discussion herein. The term expression is defined as the production of an RNA molecule from a gene, gene fragment, genomic fragment, or operon. Expression can also be used to refer to the process of peptide or polypeptide synthesis. An expression vector is defined as a vehicle by which a ribonucleic acid (RNA) sequence is transcribed from a nucleic acid sequence carried within the expression vehicle. The expression vector can also contain features that permit translation of a protein product from the transcribed RNA message expressed from the exogenous nucleic acid sequence carried by the expression vector. Accordingly, an expression vector can produce an RNA molecule as its sole product or the expression vector can produce a RNA molecule that is ultimately translated into a protein product.

Once generated, the expression library containing the exogenous nucleic acid sequences is introduced into an *E. coli* population to search for genes that are required for bacterial proliferation. Because the library molecules are foreign to the population of *E. coli*, the expression vectors and the nucleic acid segments contained therein are considered exogenous nucleic acid.

Expression of the exogenous nucleic acid fragments in the test population of *E. coli* containing the expression vector library is then activated. Activation of the expression vectors consists of subjecting the cells containing the vectors to conditions that result in the expression of the exogenous nucleic acid sequences carried by the expression vector library. The test population of *E. coli* cells is then assayed to determine the effect of expressing the exogenous nucleic acid fragments on the test population of cells. Those expression vectors that, upon activation and expression, negatively impact the growth of the *E. coli* screen population were identified, isolated, and purified for further study.

A variety of assays are contemplated to identify nucleic acid sequences that negatively impact growth upon expression. In one embodiment, growth in *E. coli* cultures expressing exogenous nucleic acid sequences and growth in cultures not expressing these sequences is compared. Growth measurements are assayed by examining the extent of growth by measuring optical densities. Alternatively, enzymatic assays can be used to measure bacterial growth rates to identify exogenous nucleic acid sequences of interest. Colony size, colony morphology, and cell morphology are additional factors used to evaluate growth of the host cells. Those cultures that failed to grow or grow with reduced efficiency under expression conditions are identified as containing an expression vector encoding a nucleic acid fragment that negatively affects a proliferation-required gene.

Once exogenous nucleic acid sequences of interest are identified, they are analyzed. The first step of the analysis is to acquire the nucleic acid sequence of the nucleic acid fragment of interest. To achieve this end, the insert in those expression vectors identified as containing a sequence of interest is sequenced, using standard techniques well known in the art. The next step of the process is to determine the source of the nucleic acid sequence.

Determination of sequence source is achieved by comparing the obtained sequence data with known sequences in various genetic databases. The sequences identified are used to probe these gene databases. The result of this

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procedure is a list of sequences nucleic acid sequences corresponding to a list that includeds novel bacterial genes required for proliferation as well as genes previously identified as required for proliferation.

The number of DNA and protein sequences available in database systems has been growing exponentially for years. For example, at the end of 1998, the complete sequences of *Caenorhabditis elegans*, *Saccharomyces cerevisiae* and nineteen bacterial genomes, including *E. coli* were available. This sequence information is stored in a number of databanks, such as GenBank (the National Center for Biotechnology Information (NCBI), and is publicly available for searching.

A variety of computer programs are available to assist in the analysis of the sequences stored within these databases. FastA, (W. R. Pearson (1990) "Rapid and Sensitive Sequence Comparison with FASTP and FASTA" Methods in Enzymology 183:63-98), Sequence Retrieval System (SRS), (Etzold & Argos, SRS an indexing and retrieval tool for flat file data libraries. Comput. Appl. Biosci. 9:49-57, 1993) are two examples of computer programs that can be used to analyze sequences of interest. In one embodiment of the present invention, the BLAST family of computer programs, which includes BLASTN version 2.0 with the default parameters, or BLASTX version 2.0 with the default parameters, is used to analyze nucleic acid sequences.

BLAST, an acronym for "Basic Local Alignment Search Tool," is a family of programs for database similarity searching. The BLAST family of programs includes: BLASTN, a nucleotide sequence database searching program, BLASTX, a protein database searching program where the input is a nucleic acid sequence; and BLASTP, a protein database searching program. BLAST programs embody a fast algorithm for sequence matching, rigorous statistical methods for judging the significance of matches, and various options for tailoring the program for special situations. Assistance in using the program can be obtained by e-mail at <a href="mailto:blast@ncbi.nlm.nih.gov">blast@ncbi.nlm.nih.gov</a>.

Bacterial genes are often transcribed in polycistronic groups. These groups comprise operons, which are a collection of genes and intergenic sequences. The genes of an operon are co-transcribed and are often related functionally. Given the nature of the screening protocol, it is possible that the identified exogenous nucleic acid sequence corresponds to a gene or portion thereof with or without adjacent noncoding sequences, an intragenic sequence (i.e. a sequence within a gene), an intergenic sequence (i.e. a sequence between genes), a sequence spanning at least a portion of two or more genes, a 5' noncoding region or a 3' noncoding region located upstream or downstream from the actual sequence that is required for bacterial proliferation. Accordingly, determining which of the genes that are encoded within the operons are individually required for proliferation is often desirable.

In one embodiment of the present invention, an operon is dissected to determine which gene or genes are required for proliferation. For example, the RegulonDB DataBase described by Huerta et al. (*Nucl. Acids Res.* 26:55-59, 1998), which may also be found on the website http://www.cifn.unam.mx/Computational\_Biology/regulondb/, may be used. to identify the boundaries of operons encoded within microbial genomes. A number of techniques that are well known in the art can be used to dissect the operon. In one aspect of this embodiment, gene disruption by homologous recombination is used to individually inactivate the genes of an operon that is thought to contain a gene required for proliferation.

Several gene disruption techniques have been described for the replacement of a functional gene with a mutated, non-functional (null) allele. These techniques generally involve the use of homologous recombination. The

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method described by Link et al. (J. Bacteriol 1997 179:6228; incorporated herein by reference in it's entirety) serves as an excellent example of these methods as applicable to disruption of genes in *E. coli*. This technique uses crossover PCR to create a null allele with an in-frame deletion of the coding region of a target gene. The null allele is constructed in such a way that sequences adjacent to the wild type gene (ca. 500 bp) are retained. These homologous sequences surrounding the deletion null allele provide targets for homologous recombination so that the wild type gene on the *E. coli* chromosome can be replaced by the constructed null allele.

The crossover PCR amplification product is subcloned into the vector pKO3, the features of which include a chloramphenicol resistance gene, the counter-selectable marker sacB, and a temperature sensitive autonomous replication function. Following transformation of an *E. coli* cell population with such a vector, selection for cells that have undergone homologous recombination of the vector into the chromosome is achieved by growth on chloramphenicol at the non-permissive temperature of 43°C. Under these conditions, autonomous replication of the plasmid cannot occur and cell are resistant to chloramphinicol only if the chloramphenicol resistance gene has been integrated into the chromosome. Usually a single crossover event is responsible for this integration event such that the *E. coli* chromosome now contains a tandem duplication of the target gene consisting of one wild type allele and one deletion null allele separated by vector sequence.

This new *E. coli* strain containing the tandem duplication can be maintained at permissive temperatures in the presence of drug selection (chloramphenicol). Subsequently, cells of this new strain are cultured at the permissive temperature 30°C without drug selection. Under these conditions, the chromosome of some of the cells within the population will have undergone an internal homologous recombination event resulting in removal of the plasmid sequences. Subsequent culturing of the strain in growth medium lacking chloramphenicol but containing sucrose is used to select for such recombinative resolutions. In the presence of the counter-selectable marker *sacB*, sucrose is rendered into a toxic metabolite. Thus, cells that survive this counter-selection have lost both the plasmid sequences from the chromosome and the autonomously replicating plasmid that results as a byproduct of recombinative resolution.

There are two possible outcomes of the above recombinative resolution via homologous recombination. Either the wild type copy of the targeted gene is retained on the chromosome or the mutated null allele is retained on the chromosome. In the case of an essential gene, a single copy of the null allele would be lethal and such cells should not be obtained by the above procedure when applied to essential genes. In the case of a non-essential gene, roughly equal numbers of cells containing null alleles and cells containing wild type alleles should be obtained. Thus, the method serves as a test for essentiality of the targeted gene: when applied to essential genes, only cells with a wild type allele on the chromosome will be obtained.

Other techniques have also been described for the creation of disruption mutations in *E. coli*. For example, Link et al. also describe inserting an in-frame sequence tag concommitantly with an in-frame deletion in order to simplify analysis of recombinants obtained. Further, Link et al. describe disruption of genes with a drug resistance marker such as a kanamycin resistance gene. Arigoni et al., (Arigoni, F. et al. A Genome-based Approach for the

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Identification of Essa. A Bacterial Genes, Nature Biotechnology 16: 851-856, the disclosure of which is incorporated herein by reference in its entirety) describe the use of gene disruption combined with engineering a second copy of a test gene such that the expression of the gene is regulated by and inducible promoter such as the arabinose promoter to test the essentiality of the gene. Many of these techniques result in the insertion of large fragments of DNA into the gene of interest, such as a drug selection marker. An advantage of the technique described by Link et al. is that it does not rely on an insertion into the gene to cause a functional defect, but rather results in the precise removal of the coding region. This insures the lack of polar effects on the expression of genes downstream from the target gene.

Recombinant DNA techniques can be used to express the entire coding sequences of the gene identified as required for proliferation, or portions thereof. The over-expressed proteins can be used as reagents for further study. The identified exogenous sequences are isolated, purified, and cloned into a suitable expression vector using methods well known in the art. If desired, the nucleic acids can contain the sequences encoding a signal peptide to facilitate secretion of the expressed protein.

Expression of fragments of the bacterial genes identified as required for proliferation is also contemplated by the present invention. The fragments of the identified genes can encode a polypeptide comprising at least 5, at least 10, at least 15, at least 20, at least 25, at least 30, at least 35, at least 40, at least 45, at least 50, at least 55, at least 60, at least 65, at least 75, or more than 75 consecutive amino acids of a gene complementary to one of the identified sequences of the present invention. The nucleic acids inserted into the expression vectors can also contain sequences upstream and downstream of the coding sequence.

When expressing the coding sequence of an entire gene identified as required for bacterial proliferation or a fragment thereof, the nucleic acid sequence to be expressed is operably linked to a promoter in an expression vector using conventional cloning technology. The expression vector can be any of the bacterial, insect, yeast, or mammalian expression systems known in the art. Commercially available vectors and expression systems are available from a variety of suppliers including Genetics Institute (Cambridge, MA), Stratagene (La Jolla, California), Promega (Madison, Wisconsin), and Invitrogen (San Diego, California). If desired, to enhance expression and facilitate proper protein folding, the codon usage and codon bias of the sequence can be optimized for the particular expression organism in which the expression vector is introduced, as explained by Hatfield, et al., U.S. Patent No. 5,082,767, incorporated herein by this reference. Fusion protein expression systems are also contemplated by the present invention.

Following expression of the protein encoded by the identified exogenous nucleic acid sequence, the protein is purified. Protein purification techniques are well known in the art. Proteins encoded and expressed from identified exogenous nucleic acid sequences can be partially purified using precipitation techniques, such as precipitation with polyethylene glycol. Chromatographic methods usable with the present invention can include ion-exchange chromatography, gel filtration, use of hydroxyapaptite columns, immobilized reactive dyes, chromatofocusing, and use of high-performance liquid chromatography. Electrophoretic methods such one-dimensional gel electrophoresis, high-resolution two-dimensional polyacrylamide electrophoresis, isoelectric focusing, and others are contemplated as purification methods.

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Also, affinity chromatographic methods, comprising antibody columns, ligand presenting columns and other affinity chromatographic matrices are contemplated as purification methods in the present invention.

The purified proteins produced from the gene coding sequences identified as required for proliferation can be used in a variety of protocols to generate useful antimicrobial reagents. In one embodiment of the present invention, antibodies are generated against the proteins expressed from the identified exogenous nucleic acid sequences. Both monoclonal and polyclonal antibodies can be generated against the expressed proteins. Methods for generating monoclonal and polyclonal antibodies are well known in the art. Also, antibody fragment preparations prepared from the produced antibodies discussed above are contemplated.

Another application for the purified proteins of the present invention is to screen small molecule libraries for candidate compounds active against the various target proteins of the present invention. Advances in the field of combinatorial chemistry provide methods, well known in the art, to produce large numbers of candidate compounds that can have a binding, or otherwise inhibitory effect on a target protein. Accordingly, the screening of small molecule libraries for compounds with binding affinity or inhibitory activity for a target protein produced from an identified gene sequence is contemplated by the present invention.

The present invention further contemplates utility against a variety of other pathogenic organisms in addition to *E. coli*. For example, the invention has utility in identifying genes required for proliferation in prokaryotes and eukaryotes. For example, the invention has utility with protists, such as *Plasmodium* spp.; plants; animals, such as *Entamoeba* spp. and *Contracaecum* spp; and fungi including *Candida* spp., (e.g., *Candida albicans*), *Saccharomyces cerevisiae*, *Cryptococcus neoformans*, and *Aspergillus fumigatus*. In one embodiment of the present invention, monera, specifically bacteria are probed in search of novel gene sequences required for proliferation. This embodiment is particularly important given the rise of drug resistant bacteria.

The numbers of bacterial species that are becoming resistant to existing antibiotics are growing. A partial list of these organisms includes: Staphylococcus spp., such as S. aureus; Enterococcus spp., such as E. faecalis; Pseudomonas spp., such as P. aeruginosa, Clostridium spp., such as C. botulinum, Haemophilus spp., such as H. influenzae, Enterobacter spp., such as E. cloacae, Vibrio spp., such as V. cholera; Moraxala spp., such as M. catarrhalis; Streptococcus spp., such as S. pneumoniae, Neisseria spp., such as N. gonorrhoeae; Mycoplasma spp., such as Mycoplasma pneumoniae; Salmonella typhimurium; Helicobacter pylori; Escherichia coli; and Mycobacterium tuberculosis. The sequences identified as required for proliferation in the present invention can be used to probe these and other organisms to identify homologous required proliferation genes contained therein.

In one embodiment of the present invention, the nucleic acid sequences disclosed herein are used to screen genomic libraries generated from bacterial species of interest other than *E. coli*. For example, the genomic library may be from *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Enterobacter cloacae*, *Helicobacter pylori*, *Neisseria gonorrhoeae*, *Enterococcus faecalis*, *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Salmonella typhimurium*, *Saccharomyces cerevisiae*, *Candida albicans*, *Cryptococcus neoformans*, *Aspergillus fumigatus*, *Klebsiella pneumoniae*, *Salmonella typhi*, *Salmonella paratyphi*, *Salmonella cholerasuis*, *Staphylococcus epidermidis*, *Mycobacterium* 

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tuberculosis, Myco. Ium leprae, Treponema pallidum, Bacillus anthracis, Persinia pestis, Clostridium botulinum, Campylobacter jejuni, Chlamydia trachomatus, Chlamydia pneumoniae or any species falling within the genera of any of the above species. Standard molecular biology techniques are used to generate genomic libraries from various microorganisms. In one aspect, the libraries are generated and bound to nitrocellulose paper. The identified exogenous nucleic acid sequences of the present invention can then be used as probes to screen the libraries for homologous sequences. The homologous sequences identified can then be used as targets for the identification of new, antimicrobial compounds with activity against more than one organism.

For example, the preceding methods may be used to isolate nucleic acids having a sequence with at least 97%, at least 95%, at least 90%, at least 85%, at least 80%, or at least 70% identity to a nucleic acid sequence selected from the group consisting of one of the sequences of SEQ ID NOS. 1-81, 405-485, 82-88, 90-242, fragments comprising at least 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, 150, 200, 300, 400, or 500 consecutive bases thereof, and the sequences complementary thereto. Identity may be measured using BLASTN version 2.0 with the default parameters. (Altschul, S.F. et al. Gapped BLAST and PSI-BLAST: A New Generation of Protein Database Search Programs, Nucleic Acid Res. 25: 3389-3402 (1997), the disclosure of which is incorporated herein by reference in its entirety). For example, the homologous polynucleotides may have a coding sequence which is a naturally occurring allelic variant of one of the coding sequences described herein. Such allelic variants may have a substitution, deletion or addition of one or more nucleotides when compared to the nucleic acids of SEQ ID NOs: 1-81, 405-485, 82-88, 90-242 or the sequences complementary thereto.

Additionally, the above procedures may be used to isolate nucleic acids which encode polypeptides having at least 99%, 95%, at least 90%, at least 85%, at least 80%, at least 70%, at least 60%, at least 50%, or at least 40% identity or similarity to a polypeptide having the sequence of one of SEO ID NOs: 243-357, 359-398or fragments comprising at least 5, 10, 15, 20, 25, 30, 35, 40, 50, 75, 100, or 150 consecutive amino acids thereof as determined using the FASTA version 3.0t78 algorithm with the default parameters. Alternatively, protein identity or similarity may be identified using BLASTP with the default parameters, BLASTX with the default parameters, or TBLASTN with the default parameters. (Alschul, S.F. et al. Gapped BLAST and PSI-BLAST: A New Generation of Protein Database Search Programs, Nucleic Acid Res. 25: 3389-3402 (1997), the disclosure of which is incorporated herein by reference in its entirety).

Alternatively, homologous nucleic acids or polypeptides may be identified by searching a database to identify sequences having a desired level of homology to a nucleic acid or polypeptide involved in proliferation or an antisense nucleic acid to a nucleic acid involved in microbial proliferation. A variety of such databases are available to those skilled in the art, including GenBank and GenSeq. In some embodiments, the databases are screened to identify nucleic acids or polypeptides having at least 97%, at least 95%, at least 90%, at least 85%, at least 80%, at least 70%, at least 60%, or at least 50%, at least 40% identity or similarity to a nucleic acid or polypeptide involved in proliferation or an antisense nucleic acid involved in proliferation. For example, the database may be screened to identify nucleic acids homologous to one of SEQ ID Nos. 1-81, 405-485, 82-88, 90-242 or polypeptides homologous

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to SEQ ID NOs. 243-537, 359-398. In some embodiments, the database may be screened to identify homologous nucleic acids or polypeptides from organisms other than *E. coli*, including organisms such as *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Enterobacter cloacae*, *Helicobacter pylori*, *Neisseria gonorrhoeae*, *Enterococcus faecalis*, *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Salmonella typhimurium*, *Saccharomyces cerevisiae*, *Candida albicans*, *Cryptococcus neoformans*, *Aspergillus fumigatus*, *Klebsiella pneumoniae*, *Salmonella typhi*, *Salmonella paratyphi*, *Salmonella cholerasuis*, *Staphylococcus epidermidis*, *Mycobacterium tuberculosis*, *Mycobacterium leprae*, *Treponema pallidum*, *Bacillus anthracis*, *Yersinia pestis*, *Clostridium botulinum*, *Campylobacter jejuni*, *Chlamydia trachomatus*, *Chlamydia pneumoniae* or any species falling within the genera of any of the above species.

In another embodiment, gene expression arrays and microarrays can be employed. Gene expression arrays are high density arrays of DNA samples deposited at specific locations on a glass chip, nylon membrane, or the like. Such arrays can be used by researchers to quantify relative gene expression under different conditions. Gene expression arrays are used by researchers to help identify optimal drug targets, profile new compounds, and determine disease pathways. An example of this technology is found in U.S. Patent No. 5807522, which is hereby incorporated by reference.

It is possible to study the expression of all genes in the genome of a particular microbial organism using a single array. For example, the arrays from Genosys consist of 12 x 24 cm nylon filters containing PCR products corresponding to 4290 ORFs from *E. coli*. 10 ngs of each are spotted every 1.5 mm on the filter. Single stranded labeled cDNAs are prepared for hybridization to the array (no second strand synthesis or amplification step is done) and placed in contact with the filter. Thus the labeled cDNAs are of "antisense" orientation. Quantitative analysis is done by phosphorimager.

Hybridization of cDNA made from a sample of total cell mRNA to such an array followed by detection of binding by one or more of various techniques known to those in the art results in a signal at each location on the array to which cDNA hybridized. The intensity of the hybridization signal obtained at each location in the array thus reflects the amount of mRNA for that specific gene that was present in the sample. Comparing the results obtained for mRNA isolated from cells grown under different conditions thus allows for a comparison of the relative amount of expression of each individual gene during growth under the different conditions.

Gene expression arrays may be used to analyze the total mRNA expression pattern at various time points after induction of an antisense nucleic acid against a proliferation-required gene. Analysis of the expression pattern indicated by hybridization to the array provides information on whether or not the target gene of the antisense nucleic acid is being affected by antisense induction, how quickly the antisense is affecting the target gene, and for later timepoints, what other genes are affected by antisense expression. For example, if the antisense is directed against a gene for ribosomal protein L7/L12 in the 50S subunit, its targeted mRNA may disappear first and then other mRNAs may be observed to increase, decrease or stay the same. Similarly, if the antisense is directed against a different 50S subunit ribosomal protein mRNA (e.g. L25), that mRNA may disappear first followed by changes in mRNA expression that are similar to those seen with the L7/L12 antisense expression. Thus, the mRNA expression pattern observed

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with an antinsense where acid against a proliferation required gene may identify other proliferation-required nucleic acids in the same pathway as the target of the antisense nucleic acid. In addition, the mRNA expression patterns observed with candidate drug compounds may be compared to those observed with antisense nucleic acids against a proliferation-required nucleic acid. If the mRNA expression pattern observed with the candidate drug compound is similar to that observed with the antisense nucleic acid, the drug compound may be a promising therapeutic candidate. Thus, the assay would be useful in assisting in the selection of candidate drug compounds for use in screening methods such as those described below.

In cases where the source of nucleic acid deposited on the array and the source of the nucleic acid being hybridized to the array are from two different organisms, gene expression arrays can identify homologous genes in the two organisms.

The present invention also contemplates additional methods for screening other microorganisms for proliferation-required genes. In this embodiment, the conserved portions of sequences identified as proliferation-required can be used to generate degenerate primers for use in the polymerase chain reaction (PCR). The PCR technique is well known in the art. The successful production of a PCR product using degenerate probes generated from the sequences identified herein would indicate the presence of a homologous gene sequence in the species being screened. This homologous gene is then isolated, expressed, and used as a target for candidate antibiotic compounds. In another aspect of this embodiment, the homologous gene is expressed in an autologous organism or in a heterologous organism in such a way as to alter the level or activity of a homologous gene required for proliferation in the autologous or heterologus organism. In still another aspect of this embodiment, the homologous gene or portion is expressed in an antisense orientation in such a way as to alter the level or activity of a nucleic acid required for proliferation of an autologous or heterologous organism.

The homologous sequences to proliferation-required genes identified using the techniques described herein may be used to identify proliferation-required genes of organisms other than *E. coli*, to inhibit the proliferation of organisms other than *E. coli* by inhibiting the activity or reducing the amount of the identified homologous nucleic acid or polypeptide in the organism other than *E. coli*, or to identify compounds which inhibit the growth of organisms other than *E. coli* as described below.

In another embodiment of the present invention, *E. coli* sequences identified as required for proliferation are transferred to expression vectors capable of function within non *E coli* species. As would be appreciated by one of ordinary skill in the art, expression vectors must contain certain elements that are species specific. These elements can include promoter sequences, operator sequences, repressor genes, origins of replication, ribosomal binding sequences, termination sequences, and others. To use the identified exogenous sequences of the present invention, one of ordinary skill in the art would know to use standard molecular biology techniques to isolate vectors containing the sequences of interest from cultured bacterial cells, isolate and purify those sequences, and subclone those sequences into an expression vector adapted for use in the species of bacteria to be screened.

Expression vectors for a variety of other species are known in the art. For example, Cao et al. report the expression of steroid receptor fragments in *Staphylococcus aureus*. J. Steroid Biochem Mol Biol. 44(1):1-11

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(1993). Also, Pla et al. have reported an expression vector that is functional in a number of relevant hosts including: Salmonella typhimurium, Pseudomonas putida, and Pseudomonas aeruginosa. J. Bacteriol. 172(8):4448-55 (1990). These examples demonstrate the existence of molecular biology techniques capable of constructing expression vectors for the species of bacteria of interest to the present invention.

Following the subcloning of the identified nucleic acid sequences into an expression vector functional in the microorganism of interest, the identified nucleic acid sequences are conditionally transcribed to assay for bacterial growth inhibition. Those expression vectors found to contain sequences that, when transcribed, inhibit bacterial growth are compared to the known genomic sequence of the pathogenic microorganism being screened or, if the homologous sequence from the organism being screened is not known, it may be identified and isolated by hybridization to the proliferation-required *E. coli* sequence interest or by amplification using primers based on the proliferation-required *E. coli* sequence of interest as described above.

The antisense sequences from the second organism which are identified as described above may then be operably linked to a promoter, such as an inducible promoter, and introduced into the second organism. The techniques described herein for identifying *E. coli* genes required for proliferation may thus be employed to determine whether the identified sequences from a second organism inhibit the proliferation of the second organism.

Antisense nucleic acids required for the proliferation of organisms other than *E. coli* or the genes corresponding thereto, may also be hybridized to a microarray containing the *E. coli* ORFs to gauge the homology between the *E. coli* sequences and the proliferation-required nucleic acids from other organisms. For example, the proliferation-required nucleic acid may be from *Staphylococcus aureus*, *Pseudomonas aeruginosa*, *Enterobacter cloacae*, *Helicobacter pylori*, *Neisseria gonorrhoeae*, *Enterococcus faecalis*, *Streptococcus pneumoniae*, *Haemophilus influenzae*, *Salmonella typhimurium*, *Saccharomyces cerevisiae*, *Candida albicans*, *Cryptococcus neoformans*, *Aspergillus fumigatus*, *Klebsiella pneumoniae*, *Salmonella typhi*, *Salmonella paratyphi*, *Salmonella cholerasuis*, *Staphylococcus epidermidis*, *Mycobacterium tuberculosis*, *Mycobacterium leprae*, *Treponema pallidum*, *bacillus anthracis*, *Yersinia pestis*, *Clostridium botulinum*, *Campylobacter jejuni or Chlamydia trachomatus*, *Chlamydia pneumoniae* or any species falling within the genera of any of the above species. The proliferation-required nucleic acids from an organism other than *E. coli* may be hybridized to the array under a variety of conditions which permit hybridization to occur when the probe has different levels of homology to the sequence on the microarray. This would provide an indication of homology across the organisms as well as clues to other possible essential genes in these organisms.

In still another embodiment, the exogenous nucleic acid sequences of the present invention that are identified as required for bacterial growth or proliferation can be used as antisense therapeutics for killing bacteria. The antisense sequences can be directed against the proliferation-required genes whose sequence corresponds to the exogenous nucleic acid probes identified here (i.e. the antisense nucleic acid may hybridize to the gene or a portion thereof). Alternatively, antisense therapeutics can be directed against operons in which proliferation-required genes reside (i.e. the antisense nucleic acid may hybridize to any gene in the operon in which the proliferation-required genes reside). Further, antisense

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therapeutics can be prected against a proliferation-required gene or portion thereof with or without adjacent noncoding sequences, an intragenic sequence (i.e. a sequence within a gene), an intergenic sequence (i.e. a sequence between genes), a sequence spanning at least a portion of two or more genes, a 5' noncoding region or a 3' noncoding region located upstream or downstream from the actual sequence that is required for bacterial proliferation or an operon containing a proliferation-required gene.

In addition to therapeutic applications, the present invention encompasses the use of nucleic acid sequences complementary to sequences required for proliferation as diagnostic tools. For example, nucleic acid probes complementary to proliferation-required sequences that are specific for particular species of microorganisms can be used as probes to identify particular microorganism species in clinical specimens. This utility provides a rapid and dependable method by which to identify the causative agent or agents of a bacterial infection. This utility would provide clinicians the ability to prescribe species specific antimicrobial compounds to treat such infections. In an extension of this utility, antibodies generated against proteins translated from mRNA transcribed from proliferation-required sequences can also be used to screen for specific microorganisms that produce such proteins in a species-specific manner.

The following examples teach the genes of the present invention and a subset of uses for the *E. coli* genes identified as required for proliferation. These examples are illustrative only and are not intended to limit the scope of the present invention.

#### **EXAMPLES**

The following examples are directed to the identification and exploitation of *E. coli* genes required for proliferation. Methods of gene identification are discussed as well as a variety of methods to utilize the identified sequences.

#### Genes Identified as Required for Proliferation of E. coli

Exogenous nucleic acid sequences were cloned into an inducible expression vector and assayed for growth inhibition activity. Example 1 describes the examination of a library of exogenous nucleic acid sequences cloned into IPTG-inducible expression vectors. Upon activation or induction, the expression vectors produced an RNA molecule corresponding to the subcloned exogenous nucleic acid sequences. The RNA product was in an antisense orientation with respect to the *E. coli* genes from which it was originally derived. This antisense RNA then interacted with sense mRNA produced from various *E. coli* genes and interfered with or inhibited the translation of the sense messenger RNA (mRNA) thus preventing protein production from these sense mRNA molecules. In cases where the sense mRNA encoded a protein required for the proliferation, bacterial cells containing an activated expression vector failed to grow or grew at a substantially reduced rate.

#### **EXAMPLE 1**

## Inhibition of Bacterial Proliferation after IPTG induction

To study the effects of transcriptional induction in liquid medium, growth curves were carried out by back diluting cultures 1:200 into fresh media with or without 1 mM IPTG and measuring the  $OD_{450}$  every 30 minutes (min). To

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study the effects of transcriptional induction on solid medium,  $10^2$ ,  $10^3$ ,  $10^4$ ,  $10^5$ ,  $10^6$ ,  $10^7$  and  $10^8$  fold dilutions of overnight cultures were prepared. Aliquots of from 0.5 to 3  $\mu$ l of these dilutions were spotted on selective agar plates with or without 1 mM IPTG. After overnight incubation, the plates were compared to assess the sensitivity of the clones to IPTG.

Of the numerous clones tested, some clones were identified as a containing sequence that inhibited *E. coli* growth after IPTG induction. Accordingly, the gene to which the inserted nucleic acid sequence corresponds, or a gene within the operon containing the inserted nucleic acid, may be required for proliferation in *E. coli*.

## Characterization of Isolated Clones Negatively Affecting E. coli Proliferation

Following the identification of those expression vectors that, upon expression, negatively impacted *E. coli* growth or proliferation, the inserts or nucleic acid fragments contained in those expression vectors were isolated for subsequent characterization. Expression vectors of interest were subjected to nucleic acid sequence determination.

#### **EXAMPLE 2**

# Nucleic Acid Sequence Determination of Identified Clones Expressing Nucleic Acid Fragments with Detrimental Effects of *E. coli* Proliferation

The nucleotide sequences for the exogenous identified sequences were determined using plasmid DNA isolated using QIAPREP (Qiagen, Valencia, CA) and methods supplied by the manufacturer. The primers used for sequencing the inserts were 5' - TGTTTATCAGACCGCTT - 3' (SEQ ID NO: 403) and 5' - ACAATTTCACACAGCCTC - 3' (SEQ ID NO: 404). These sequences flank the polylinker in pLEX5BA. Sequence identification numbers (SEQ ID NOs) for the identified inserts are listed in Table I and discussed below.

#### **EXAMPLE 3**

## Comparison Of Isolated Sequences to Known Sequences

The nucleic acid sequences of the subcloned fragments obtained from the expression vectors discussed above were compared to known *E. coli* sequences in GenBank using BLAST version 1.4 or version 2.0.6 using the following default parameters: Filtering off, cost to open a gap = 5, cost to extend a gap = 2, penalty for a mismatch in the blast portion of run = .3, reward for a match in the blast portion of run = 1, expectation value (e) = 10.0, word size = 11, number of one-line descriptions = 100, number of alignments to show (B) = 100. BLAST is described in Altschul, J Mol Biol. 215:403-10 (1990), the disclosure of which is incorporated herein by reference in its entirety. Expression vectors were found to contain nucleic acid sequences in both the sense and antisense orientations. The presence of known genes, open reading frames, and ribosome binding sites was determined by comparison to public databases holding genetic information and various computer programs such as the Genetics Computer Group programs FRAMES and CODONPREFERENCE. Clones were designated as "antisense" if the cloned fragment was oriented to the promoter such that the RNA transcript produced was complementary to the expressed mRNA from a chromosomal locus. Clones were designated as "sense" if they coded for an RNA fragment that was identical to a portion of a wild type mRNA from a chromosomal locus.

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The sequents described in Examples 1-2 that inhibited bacterial proliferation and contained gene fragments in an antisense orientation are listed in Table I. This table lists each identified sequence by: a sequence identification number; a Molecule Number; a gene to which the identified sequence corresponds, listed according to the National Center for Biotechnology Information (NCBI), Blattner (Science 277:1453-1474(1997); also contains the *E. coli* K-12 genome sequence), or Rudd (Micro. and Mol. Rev. 62:985-1019 (1998)), (both papers are hereby incorporated by reference) nomenclatures. The CONTIG numbers for each identified sequence is shown, as well as the location of the first and last base pairs located on the *E. coli* chromosome. A Molecule Number with a "\*\*" indicates a clone corresponding to an intergenic sequence.

The sequences of the nucleic acid inserts of SEQ ID NOs: 1-81 from U.S. Provisional Patent Application No. 60/117,405 which inhibited proliferation were further analyzed. The reanalyzed sequences corresponding to SEQ ID NOs. 1-81 of U.S. Provisional Patent Application No. 60/117,405 have SEQ ID NOs. 405-485 in the present application.

SEQ ID NOs: 82-242 in U.S. Provisional Patent Application No. 60/117,405 are identical to SEQ ID NOs: 82-242 of the present application with the following exceptions. SEQ ID NO: 148 in the present application is the complementary strand of SEQ ID NO: 148 in U.S. Provisional Patent Application No. 60/117,405. Accordingly, the protein of SEQ ID NO: 308 which is encoded by SEQ ID NO: 148 has also been revised. SEQ ID NO: 163 in the present application is the complementary strand of SEQ ID NO: 163 in U.S. Provisional Patent Application No. 60/117,405. Accordingly, the protein of SEQ ID NO: 323 which is encoded by SEQ ID NO: 163 has also been revised.

The target gene of SEQ ID NOs. 18 and 19 of U.S. Provisional Patent Application No. 60/117,405 (SEQ ID NOs. 18, 19, 422, 423 of the present application) has been revised from dicF to ftsZ to reflect the fact that these SEQ ID NOs. include natural antisense molecules which inhibit ftsZ expression.

The gene products of the nucleic acids of SEQ ID NOs. 198 and 239-242 in U.S. Provisional Patent Application No. 60/117,405 and in the present application (SEQ ID NOs. 358 and 399-402 of the present application) have been revised to reflect the fact that these nucleic acids encode nontranslated tRNAs and rRNAs. Tables I and II have been revised accordingly. The SEQ ID NOs. in Table II were also revised to reflect the fact that SEQ ID NOs: 89 and 402 were identical in U.S. Provisional Patent Application No. 60/117,405.

TABLE I

Identified Clones with Corresponding Genes and Operons

SEQ ID NO.	Molecule No.	Gene (NCBI)	Gene (Blattner)	Gene (Rudd)	CONTIG
1, 405	EcXA001	yhhQ.	b3471	yhhΩ	AE000423
2, 406	EcXA002	lepB	lepB	lepB	AE000343
3, 407	EcXA003	f586	b0955	ycbZ	AE000197
4, 408	EcXA004	rpsG, rpsL	b3341	rpsG, rpsL	AE000410
5, 409	EcXA005a	rpIL, rpIJ	<i>b3986</i>	rplL, rplJ	AE000472
6, 410	EcXA005b	rpIL	rp/L	rpIL	AE000472
7, 411	EcXA005c	rpIL, rpIJ	rplL, rplJ	rplL, rplJ	AE000472
8, 412	EcXA005d	rpIL, rpIJ	rplL, rplJ	rplL, rplJ	AE000472
9, 413	EcXA005e	rpIL	rpIL	rpIL	AE000472

11, 415	Oro	T		<del></del>		
11.415	NO.		(NCBI)		:	CONTIG
11, 415				rpl	L rplL	AE000472
13, 416				rpli		
13, 417   ECXAOOR   YiP   b3666   YiP   AEDOO44     14, 418   ECXAOOR   YhAU   YhAU   YhAU   AEDOO39     15, 419   ECXAOOR   YhAU   YhAU   YhAU   AEDOO39     16, 420   ECXAOOR   YhAU   YhAU   YhAU   AEDOO39     17, 421   ECXAOOR   YhAU   YhAU   YhAU   AEDOO39     18, 422   ECXAOOR   YhAU   YhAU   YhAU   AEDOO39     18, 422   ECXAOOR   YhAU   YhAU   YhAU   AEDOO39     19, 423   ECXAOOR   YhAU   YhAU   YhAU   AEDOO39     19, 423   ECXAOOR   YhAU   YhAU   YhAU   YhAU   YhAU   AEDOO25     20, 424   ECXAOOL   YHAU   YHAU   YHAU   YHAU   AEDOO24     21, 425   ECXAOOL   YHAU   YHAU   YHAU   YHAU   AEDOO34     22, 426   ECXAOOL   YHAU   YHAU   YHAU   AEDOO34     23, 427   ECXAOOL   YHAU   YHAU   YHAU   AEDOO34     24, 428   ECXAOOL   YHAU   YHAU   YHAU   AEDOO34     25, 429   ECXAOOL   YHAU   YHAU   YHAU   YHAU   AEDOO34     26, 430   ECXAOOL   YHAU   YHAU   YHAU   YHAU   YHAU   AEDOO34     26, 430   ECXAOOL   YHAU   Y			<del> </del>	b2297		
14, 418	<del></del>	<del></del>	yicP	b366t	_	
15, 419		EcXA008a	yhaU	b3127		
16, 420		EcXA008b	yhaU	yhal.		
17, 421			yhaU	yhaL		
18, 422		EcXA009	ydeY	yde		<del></del>
19, 423	18, 422	1	dicF	b1575		
20, 424						
20, 424		EcXA010b	dicF	dicF	dicF	AE000253
21, 425         EcXA012a         fusA         b3340         fusA         AE00041           22, 426         EcXA012b         fusA         fusA         fusA         AE00041           23, 427         EcXA012c         fusA         fusA         fusA         AE00041           24, 428         EcXA013a         o86         b2562         yfhL         AE00034           25, 429         EcXA013c         o86         b2562         yfhL         AE00034           26, 430         EcXA014         visC         b2906         visC         AE00034           27, 431         EcXA014         visC         b2906         visC         AE00037           28, 432         EcXA015         yfdl         yfdl         yfdl         yfdl         AE00037           29, 433         EcXA016         yea0         yea0         yea0         yea0         AE00037           30, 434         EcXA017b         yggE         b2922         yggE         AE00037           31, 435         EcXA018a         o464         b2074         yegM         AE00029           32, 436         EcXA018b         o464         b2074         yegM         AE00029           34, 438         EcXA018b		EcXA011	fdnG	61474		
22, 426         EcXA012b         fusA         fusA         fusA         AE000411           23, 427         EcXA012c         fusA         fusA         fusA         AE000411           24, 428         EcXA013a         o86         b2562         yfhL         AE00034:           25, 429         EcXA013b         o86         b2562         yfhL         AE00034:           26, 430         EcXA013c         o86         b2562         yfhL         AE00034:           27, 431         EcXA014         visC         b2906         visC         AE00037.           28, 432         EcXA015         yfdl         yfdl         yfdl         AE00027.           29, 433         EcXA016         yea0         yea0         yea0         yea0           30, 434         EcXA017a         yggE         b2922         yggE         AE000375           31, 435         EcXA018a         o464         b2074         yegM         AE000297           33, 437         EcXA018b         o464         b2074         yegM         AE000293           34, 438         EcXA019a         yehA         yehA         yehA         yehA         AE000293           35, 439         EcXA021b         o172, yeh		EcXA012a	fusA	b3340		
23, 427         EcXA012c         fusA         fusA         fusA         AE000411           24, 428         EcXA013a         086         b2562         yfhL         AE000342           25, 429         EcXA013b         086         b2562         yfhL         AE000342           26, 430         EcXA013c         086         b2562         yfhL         AE000342           27, 431         EcXA014         visC         b2906         visC         AE000372           28, 432         EcXA015         yfdl         xfdl         xfa000372         xfa06         x62022         yggF         AE000372 <td></td> <td>EcXA012b</td> <td>fusA</td> <td>fusA</td> <td></td> <td></td>		EcXA012b	fusA	fusA		
24, 428         EcXA013b         086         b2562         yfhl         AE00034           25, 429         EcXA013b         086         b2562         yfhl         AE00034           26, 430         EcXA013c         086         b2562         yfhl         AE00037           27, 431         EcXA014         visC         b2906         visC         AE00037           28, 432         EcXA015         yfdl         yfdl         yfdl         yfdl         AE00037           29, 433         EcXA016         yeaQ         yeaQ         yeaQ         yeaQ         AE00037           30, 434         EcXA017a         yggE         b2922         yggE         AE00037           31, 435         EcXA017b         yggE         yggE         yggE         yggE         AE00037           32, 436         EcXA018a         o464         b2074         yegM         AE00029           34, 438         EcXA018b         o464         b2074         yegM         AE00029           35, 439         EcXA019b         o172, yehA         o172, yehA         o172, yehA         AE00029           36, 440         EcXA021a         f112         b0218         yafU         AE00029           39, 4		EcXA012c	fusA		<del></del>	
25. 429         EcXA013b         o86         b2562         yfhL         AE00034:           26, 430         EcXA013c         o86         b2562         yfhL         AE00034:           27, 431         EcXA014         visC         b2906         visC         AE00037.           28, 432         EcXA015         yfdl         yfdl         yfdl         yfdl         AE00037.           29, 433         EcXA016         yea0         yea0         yea0         yea0         AE00027.           30, 434         EcXA017a         yggE         b2922         yggE         AE00037.           31, 435         EcXA017b         yggE         yggE         yggE         yggM         AE00037.           32, 436         EcXA018a         o464         b2074         yegM         AE00029.           34, 438         EcXA018b         o464         b2074         yegM         AE00029.           35, 439         EcXA019b         o172, yehA         o172, yehA         o172, yehA         AE00029.           36, 440         EcXA021a         f112         b0218         yafU         AE000130.           39, 443         EcXA021b         f112         b0218         yafU         AE000130.		EcXA013a	086	b2562		
26, 430         EcXA013c         a86         b2562         yfhl         AE00034           27, 431         EcXA014         visC         b2906         visC         AE000374           28, 432         EcXA015         yfdl         yfdl         yfdl         yfdl         AE00032           29, 433         EcXA016         yea0         yea0         yea0         yea0         AE000274           30, 434         EcXA017a         yggE         b2922         yggE         AE000375           31, 435         EcXA017b         yggE         yggE         yggM         AE000297           32, 436         EcXA018a         o464         b2074         yegM         AE000297           34, 438         EcXA018b         o464         b2074         yegM         AE000293           34, 438         EcXA019a         yehA         yehA         yehA         yehA         AE000293           35, 439         EcXA019b         o172, yehA         o172, yehA         o172, yehA         AE000299           36, 440         EcXA021a         f112         b0218         yafU         AE000130           39, 443         EcXA021b         f112         b0218         yafU         AE000130		EcXA013b	086	b2562		
Carrell		EcXA013c	o86	b2562		
28, 432         EcXA015         yfdl         yfdl         yfdl         yfdl         AE000323           29, 433         EcXA016         yea0         yea0         yea0         AE000274           30, 434         EcXA017a         yggE         b2922         yggE         AE000375           31, 435         EcXA017b         yggE         yggE         yggE         yggM         AE000297           32, 436         EcXA018a         0464         b2074         yegM         AE000293           34, 438         EcXA019a         yehA         yehA         yehA         yehA         yehA         AE000293           35, 439         EcXA019b         0172, yehA         0172, yehA         0172, yehA         AE000293           36, 440         EcXA021a         f112         b0218         yafU         AE000130           38, 442         EcXA021a         f112         b0218         yafU         AE000130           39, 443         EcXA022b         0740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         yde	27, 431	EcXA014	visC	b2906		
29,433   EcXA016	28, 432	EcXA015	yfd/	yfdl		
Yoa6	29, 433	EcXA016	yea ()			
30, 434			yoaG			112000274
31, 435         EcXA017b         yggE         yggE         yggE         AE00037E           32, 436         EcXA018a         0464         b2074         yegM         AE000297           33, 437         EcXA018b         0464         b2074         yegM         AE000297           34, 438         EcXA019a         yehA         yehA         yehA         yehA         AE000300           35, 439         EcXA019b         0172, yehA         0172, yehA         0172, yehA         AE000299           36, 440         EcXA020         0384, 182         b1794, b1795         yeaP, yeaO         AE000299           37, 441         EcXA021a         1112         b0218         yafU         AE000130           38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         0740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000289 </td <td></td> <td>EcXA017a</td> <td>yggE</td> <td></td> <td></td> <td>AF000375</td>		EcXA017a	yggE			AF000375
32, 436         EcXA018a         0464         b2074         yegM         AE000297           33, 437         EcXA018b         0464         b2074         yegM         AE000297           34, 438         EcXA019a         yehA         yehA         yehA         yehA         AE000299           35, 439         EcXA019b         o172, yehA         o172, yehA         o172, yehA         AE000299           36, 440         EcXA020         o384, f82         b1794, b1795         yeaP, yeaO         AE000274           37, 441         EcXA021a         f112         b0218         yafU         AE000130           38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         o740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygiM, ygiN         b3082         ygiM, ygiN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289<		EcXA017b				<del>,</del>
33, 437         EcXA018b         0464         b2074         yegM         AE000297           34, 438         EcXA019a         yehA         yehA         yehA         yehA         AE000299           35, 439         EcXA019b         0172, yehA         0172, yehA         0172, yehA         AE000299           36, 440         EcXA020         0384, f82         b1794, b1795         yeaP, yeaO         AE000274           37, 441         EcXA021a         f112         b0218         yafU         AE000130           38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         0740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE00039           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA027a         yohH         yohH         yohH         yohH						
34, 438         EcXA019a         yehA         yehA         yehA         AE000300           35, 439         EcXA019b         o172, yehA         o172, yehA         o172, yehA         AE000299           36, 440         EcXA020         o384, f82         b1794, b1795         yeaP, yeaQ         AE000274           37, 441         EcXA021a         f112         b0218         yafU         AE000130           38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         o740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted <td< td=""><td>33, 437</td><td>EcXA018b</td><td>0464</td><td></td><td></td><td></td></td<>	33, 437	EcXA018b	0464			
35. 439	34, 438	EcXA019a	yehA			
35, 439         EcXA019b         o172, yehA         o172, yehA         o172, yehA         AE000299           36, 440         EcXA020         o384, f82         b1794, b1795         yeaP, yeaO         AE000274           37, 441         EcXA021a         f112         b0218         yafU         AE000130           38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         o740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000389           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000303           45, 449         EcXA027a         yohH         yohH         yohH         yohH         yohH           46, 450         EcXA027b         yohH         yohH         yohH         y					/	
36, 440         EcXA020         o384, f82         b1794, b1795         yeaP, yeaU         AE000274           37, 441         EcXA021a         f112         b0218         yafU         AE000130           38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         o740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000390           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000303           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         yohH         AE000303           48, 452         EcXA028         f296         b2305	35, 439	EcXA019b	o172, yehA	o172, yehA	0172. vehA	
37, 441         EcXA021a         f112         b0218         yafU         AE000130           38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         o740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         AE000303           48, 452         EcXA028         f296         b2305         yfc/	36, 440	EcXA020	o384, f82			
38, 442         EcXA021b         f112         b0218         yafU         AE000130           39, 443         EcXA022         o740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         AE000303           48, 452         EcXA027d         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305	37, 441	EcXA021a	f112			
39, 443         EcXA022         0740         b1629         ydgN         AE000258           40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         yohH           48, 452         EcXA027d         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305         yfc/         AE000319		EcXA021b	f112			
40, 444         EcXA023a         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         AE000303           48, 452         EcXA027d         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305         yfc/         AE000319	39, 443	EcXA022	0740			
41, 445         EcXA023b         f176, f382         b1504, b1505         ydeS, ydeT         AE000247           42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH           48, 452         EcXA027d         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305         yfc/         AE000319		EcXA023a	f176, f382	b1504, b1505		
42, 446         EcXA024         ygjM, ygjN         b3082         ygjM, ygjN         AE000390           43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         yohH           48, 452         EcXA027d         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305         yfcl         AE000319	41, 445	EcXA023b	1176, 1382			
43, 447         EcXA025         02383         b1878         yeeJ         AE000289           44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         yohI           48, 452         EcXA027d         yohH         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305         yfcl         AE000319	42, 446	EcXA024	ygjM, ygjN	b3082		
44, 448         EcXA026         o61         Unpre-dicted         Unpre-dicted         AE000138           45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         yohH           48, 452         EcXA027d         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         1296         b2305         yfc/         AE000319		EcXA025	02383			
45, 449         EcXA027a         yohH         yohH         yohH         yohH         AE000303           46, 450         EcXA027b         yohH         yohH         yohH         yohH         AE000303           47, 451         EcXA027c         yohH         yohH         yohH         yohH         yohH           48, 452         EcXA027d         yohH         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305         yfc/         AE000319	44, 448	EcXAD26	061			
46, 450   EcXA027b   yohH   yohH   yohH   AE000303	45, 449	EcXA027a	yohH			
47, 451   EcXA027c   yohH   yohH   yohH   AE000303	46, 450	EcXA027b	yohH			
	47, 451	EcXA027c	yohH			
48, 452         EcXA027d         yohH         yohH         yohH         AE000303           49, 453         EcXA028         f296         b2305         yfcl         AE000319           50, 454         EcxA028         iii         iii         AE000319			yoh/			
49, 453	48, 452	EcXA027d	yohH			AF000303
50 454 FeVA020		EcXA028				
		EcXA029	yjjK	b4391	уіјК	AE000579
51, 455 EcXA030 <i>yi5A b3557 yi5A</i> AE000433	51, 455	EcXA030				
52, 456 EcXA031 <i>rplE B3308 rplE</i> AE000408	52, 456	EcXA031				
53, 457 EcXA032a ybgD ybgD ybgD AF000175		EcXA032a				
54, 458 EcXA032b**	54, 458	EcXA032b**				

SEQ ID	Molecule No.	Gene	Gene	Gene	CONTIG
NO.		(NCBI)	(Blattner)	(Rudd)	,
		gltA	gltA	gltA	
55, 459	EcXA033a	f477 (as)	b3052	· waaE	AE000387
	·	·			AE000386
56, 460	EcXA033b	f477	b3052	waaE	AE000387
57, 461	EcXA034a	cspA	b3556	cspA	AE000433
58, 462	EcXA034b	сѕрА	b3556	cspA	AE000433
59, 463	EcXA035	yhjU	yhjU	yhjU	AE000431
60, 464	EcXA036	yąjF	b3101	yqjF	AE000392
		o99	b3100,	yąjK	
61, 465	EcXA037	ydeH	b1535	ydeH	AE000251
62, 466	EcXA038	sieB	b1353	sieB	AE000233
63, 467	EcXA039	ybbD		ybbD	AE000156
64, 468	EcXA040	InsB 6	b3445	insB 6	AE000420
65, 469	EcXA041	f234	b1138	·· ymfE	AE000214
66, 470	EcXA042a	rpIY	rplY	rplY	AE000308
67, 471	EcXA042b	rplY	rplY	rplY	AE000308
68, 472	EcXA043	ybgB	ybgB	ybgB	AE000176
		cydA	cydA	cydA	
69, 473	EcXA044	purB	b1131	purB	AE000213
70, 474	EcXA045**	csrA	csrA	csrA	AE000353
	·	serV	serV	serV	
71, 475	EcXA046**	fimE, fimA	b4313	fimE, fimA	AE000502
72, 476	EcXA047**	f96, cspB	f96, cspB	cspB, ydfS	AE000252
73, 477	EcXA048	yefE	yefE	yefE	AE000294
74, 478	EcXA049	yaiC	b0385	yaiC	AE000145
75, 479	EcXA050	0467, 0222	yaiU,yaiV	yaiU, yaiV	AE000144
76, 480	EcXA051a	rpl8, rplW	rplB, rplW	rplB, rplW	AE000408
77, 481	EcXA051b	rpIW	rpIW	rpIW	AE000408
78, 482	EcXA052	infC	infC	infC	AE000267
	)				AE000266
79, 483	EcXA053	gor	gor	gor	AE000426
80, 484	EcXA054	rpIF	rpIF	rpIF	AE000408
81, 485	EcXA055	rrlG	rrlG	rrIG	AE000345

**EXAMPLE 4** 

## Identification of Genes and their Corresponding Operons Affected by Antisense Inhibition

The sequencing of the entire E. coli genome is described in Blattner et al., Science 277:1453-1474(1997) the entirety of which is hereby incorporated by reference and the sequence of the genome is listed in GenBank Accession No.U00096, the disclosure of which is incorporated herein by reference in its entirety. The operons to which the proliferation inhibiting nucleic acids correspond were identified using RegulonDB and information in the literature. The coordinates of the boundaries of these operons on the E. coli genome are listed in Table III. Table II lists the molecule numbers of the inserts containing the growth inhibiting nucleic acid fragments, the genes in the operons corresponding to the inserts, the SEQ ID NOs of the genes containing the inserts, the SEQ ID NOs of the proteins encoded by the genes, the start and stop points of the genes on the E. coli genome, the orientation of the genes on the genome, whether the operons

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are predicted or documented, and the predicted functions of the genes. The identified operons, their putative functions, and whether or not the genes are presently thought to be required for proliferation are discussed below.

Functions for the identified genes were determined by using either Blattner functional class designations or by comparing identified sequence with known sequences in various databases. A variety of biological functions were noted for the genes to which the clones of the present invention correspond. The functions for the genes of interest appear in Table II.

The proteins that are listed in Table II are involved in a wide range of biological functions.

TABLE II

All Operon Data with Whole Chromosome Coordinates

Predicted functional class of encoded proteins	Hypothetical outer	Resistance to phage C1; periplasmic protein perhaps anchored to inner membrane	Secretion	Protease	Translation (Elongation	Translation (elongation	factor efg) Translation	Translation (rRNA)	Translation	Translation	Translation	Carbon compound	Probable adenine deaminase
Blattner functional class of encoded proteins	Hypothetical ORF, unclassified, unknown	Hypothetical ORF, unclassified, unknown	Transport and binding proteins	Unknown	Translation, post- translational modification	Translation, post	Translation, post	Translational modification translation, post translational modification	Translation, post- translational modification	Translation, post- translational modification	Translation, post- translational modification		al ORF, d, unknown
Predicted (P) Or Documented (D) Operon	(P)		(P)	(P)	(0)			-		(D)		(P)	(P)
Right Coordinate	3607513	3608143	2703329	1017522	3468966	3471151	3471718	2729178	3471815	4178071	4178503	2414911	3843357
Left Coordinate	3606848	3607532	2702355	1015762	3467782	3469037	3471179	2727636	3471815	4177574	4178138	2412767	3841591
Genes On Operon	DyyA	dcrB	ІерВ	усь Z	tufA	fusA	gsdı	nse	rsdı	rplJ	ıpll	pta	уісР
Mole. No.	EcXA001		EcXA002	EcXA003	EcXA004			EcXA055		EcXA005a-g		EcXA006	EcXA007
Gene Prod. Seq ID No.	243	244	245	246	247	248	249	402	250	7.5.1	252	253	254
Gene Seq 1D No.	82	83	84	85	98	87	88	89	06	F (	26	93	94

Blattner functional class   Predicted functional class	of encoded proteins								Probable integral membrane	protein Phthalate permease		Putative ABC transporter					-				Regulator of cell division	Anaprohic respiration	(formate dehydro-genase)				No homologues, no motifs	Ilhiminana svatbasis	ordening synthesis
Blattner functional class	of encoded proteins			Hypothetical ORF,	unclassified, unknown	Putative enzymes	Hypothetical ORF,	unclassified, unknown	Carbon compound	catabolism	Putative transport proteins	Putative transport proteins	Hypothetical ORF,	unclassified, unknown		Adaptation, protection) Energy metabolism		Energy metabolism	Energy metabolism			Hypothetical ORF	own						
Predicted (P)	. Or	Documented (D)	Operon	(P)							(P)										( <u>H</u>	(0)					(P)	(B)	
Right	Coordinate			3269492		3270407	3271198		3272548		1601049	1602071	1603063		1604097		1604999		1605313		106456	1548472		1549369	1550015		2697943	3050337	
Left Coordinate				3268266		3269508	3270428		3271214	. # 1 V	1599514	1601043	1602071	·	1603075		1604124		1605023		105305	1545425		1548485	1549362		2697683	3049135	
Genes On	Operon			Dey		yhaE	yhaF		yhaU		Хәрл	ydeY	ZapA		yneA		yneB		Jaux		1187	9upj_		Hupj	tqu	Same operon as EcXA004	NHIL	visC	
Mole. No.				EcXA008a.c	-		,		·		EcXA009								J		ECXAU10a-b	EcXA011				EcXA 012a·c	EcXA013a-c	EcXA014	
Gene Prod.	Sed ID No.			255	1	992	257		. 258		259	260	261		262		263		797		765	266		267	268		269	270	
GeneSeq	2 :			66 		.gg	97		86		66	100	101		102		201		104	10,	ca Ca	106		107	108	-	109	110	

1			<del>,-</del> -			<del>-</del> -					_	_																			
Predicted functional class										-			Putative membrane protein		Homologue to transolv.	cosylase associated protein		No homologues				Homologues in multiple	bacteria, no motifs	Transport (multiple	transferable resistance)						Weak homology to pilin precursor from H Inf
Blattner functional class	of encoded proteins		Biosynthesis of cofactors.	prosthetic groups and	carriers	Translation, post-	translational modification	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Cell structure	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown		Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Structural proteins	_	Putative transport proteins		Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Putative transport proteins	<del>                                     </del>
Predicted (P)	ŏ	Documented (D)	operon							(d)					(P)		.	<u> </u>				(d)		<u> </u>			3		3		(P)
Right	Coordinate		3051538			3052860		3053470		2466237		2467154	2468482		1877279	-		1877609		1877972		3066100		2153285		2156407		.2159485		2160901	2186434
Left Coordinate			3050360		100	3051535	:: 000	3052886		24658/5		2466234	2467151		1877031		1011101	18/142/	0,0110,	18//613	0001000	2003300	111001	7151881		2153285		2156408	0010	7123480	2185400
Genes On	Operon		Hiqn		0.00	Jdad	9	Ygib	9	nan	1117	YIGH	yfal	,	nead		0,000	Yeau	Que.	head	June	Yyyc	Mach	Negru		Negiv		ngal	door	dist	уепд
Möle. No.	-	The sage of the sa							CAYANIE	LLAMOID		-		T. VA010	ECAAUID						FrYA017ah	a.B. (1000)	Er YAO182 h	LLANU 104:U						7.VA040.1	CCAAU 198-D
Gene Prod.	oed in Mo.		271		27.9	7/7	27.3	3	27.4	1,	275	212	9/7	77.6	//7		278	27	979	3	280		281		787	707	200	607	284	200	607
GeneSeq	2 S		==		117	:	113	2	114		115	110	<u> </u>	117	-		118		119		120		121	<u>.</u>	122	17.	123		124	125	631

Predicted functional class	of encoded proteins		-								•	Homologues in H. Inf. and S.	Pombe., no motifs,	transmem-brane region																fimf-like	
Blattner functional class   P	of encoded proteins			Hypothetical ORF,	unclassified, unknown	Putative chaperones	Cell structure					Hypothetical ORF,	unclassified, unknown   Po	<u>.</u>	Hypothetical ORF,	unclassified, unknown	Hypotnetical UKF, unclassified. unknown	Hypothetical ORF,	unclassified, unknown	Transcription, RNA	processing and degradation	Hypothetical ORF,	unclassified, unknown		unclassified, unknown						
Predicted (P)	0 <b>r</b>	Documented (D)	Operon									(P)			(P)												<u></u>	(d)			<b>-</b>
Right	Coordinate			2188930		2189665	2190242					239084		į	1704372		1704950		1707165		1708224	1700040	0 6000 / 1	1709547		1710182		1586320		1586863	
Left Coordinate				2186450		7188946	2189700			¥		238746			1703791		1704372		1704943	000	1/0/166	0000001	0770071	1708852		1709547		1585817		1586333	
Genes On	Uperon			kehB		henc	уенО	Same operon	as EcXA016	(one of the	(wo)	yafU			16p.l		NgbN		NgbA		Nago	doby	1.091	Vdga		nth		удеВ		Sapl	
Mole. No.								EcXA020				EcXA021a·b		·	EcXA022													EcXA023a-b	-		7
Gene Prod.	on or hac		000	987	787	/07	288				·	289			290		167	.00	767	202	C67	294		295	CUC	987		297	000	967	
GeneSeq	2 · 2		000	971	177	13.	871					129			130	101	2	192	761	133	<u> </u>	134		132	126	95		137	120	000	

	· · · · · · · · · · · · · · · · · · ·		_		4									_			 											
Pre	of encoded proteins	find-like	Weak homology to long	chain fatty acid coa ligase in	Archaeglobus	Homologues in various	bacteria	Strong similarity to	numerous attaching amd	elfacing proteins and	invasins	nifm like			Xylose binding protein·like		Similar to S. Typhi histidine	fransport gene	Similar to ABC transporter		IS150 orf A							
Blattner functional class		Structural proteins	Hypothetical ORF,	unclassified, unknown		Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown				Putative transport proteins	4401 . 1.	Hypothetical URF,	Putative regulatory protein	Hypothetical ORF,	Unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	Phage, transposon, or	plasmid	Translation, post-	translational modification	Putative transport proteins	Translation, post-	translational modification
Predicted (P)	Documented (D) Operon		(a)					(P)				unpredicted	( <b>b</b> )			(P)	(P)		€		<u>(</u> )			(0)				
Right		1588025	3231785			3232036		2050036	<del></del>			331184	2226539	Japanen	6000777	2228405	2421559		4628091		3/18830	3719678		3440371		3441734	3442176	
Left Coordinate		1586877	3231369	·		3231782	:: *	2042885				331001	2225343	222660	6060777	2227458	2420669		4626424		37 18309	3718827	·	3440255	-	3440403	3441742	
Genes On Operon		rdeT	WgjM			NgjN		reeJ				rajA	Эчол	Мфил	, in a	yohl	hcf!	1	y/l/	į	HCI/	yi5B		Cmdı		prlA	Dldı	
Mole. No.			EcXA024					EcXA025			20047	ECXAU26	EcXA027a d				EcXA028	7. ٧٨٥٥٥	ECAAUZY	7.74000	ECANUSU			EcXA031				
Gene Prod. Seq ID No.		299	300			30.		305			000	303	304	305		306	307	000	200	000	60°	310		311		312	333	-
GeneSeq ID	No.	139	140			141		14.5			112	143	144	145		146	147	140	0	140	<u> </u>	150		151	5	761	153	

SS				T		T	-	7		T		Γ		Т		Т		Т		T		Τ-			т-				170		1			
Predicted functional class										Translation						Translation						Hypothetical fimbrial protein		Glutamine biosynthesis	ADP heptose synthase/	autotrophic growth protein					RNA chaperonin			•
Blattner functional class of encoded proteins				Translation, post-	translational modification	Translation, post-	translational modification	Translation, post-	translational modification	Translation, post	translational modification	Translation, post-	translational modification	Cell processes (incl.	Adaptation, protection)	Energy metabolism	Putative enzymes		Translation, post-	translational modification	Hypothetical ORF,	unclassified, unknown		ction)	Translation, post-	translational modification								
Predicted (P) Or	Documented	(Q)	Operon											•								(d)			(P)						(P)		(d)	1
Right Coordinate				3442359		3442866		3443234		3443777		3444182		3444521		3445075		3445404		3445786		752018		753691	3194394		3197282		3198606		3717890	100	3695658	
Left Coordinate				3442180		3442363		3442881		3443244		3443790		3444216		3444536		3445090		3445415		751452		/52408	3192961		3194442		3197305	1	3717678	1004000	3694087	
Genes On Operon				<i>Omds</i>		ıpsE		rpIR		lds		rpsH		Nsdı .		rplE		Xldı		Md		DbqX		gitA	waa£	l.	gint		Ygit	•	cspA	1.10	clul	-
Mole. No.																						EcXAU32a·b		. 00047. 1.	ECAAU33a-b					F-V4004-1	ECAAU34a.D	Ervange	CCAAU33	
Gene Prod. Seq ID No.		-		314	2,50	315		316	,	- 31/		318	9	316		370		321	000	377	000	353	866	324	979	300	320	700	776	97.6	370	320	670	
GeneSeq 1D			1	154		133	0.1	061	[	/cl		28		661		ngı	,,,,	9	25.	791	55	<u> </u>	164	10.	COI	100	001	167	<u> </u>	169	9	169	3	

_					,												_	_					U		'									
	Predicted functional class of encoded proteins						Regions similar to dehydro-	genases, nucleases etc.	·	•							Homologues in many	bacteria, blocks; secretion/	ATP synthase/ftsz		Similar to carboxy-kinase,	oxidase, symporters	Super-infection exclusion	factor B-like					-		Rhs-like element		ATP synthase, desaturase	
Diatement from the	of encoded proteins			•	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Similar to mukb from H. Inf.	flh-1,1 onr	Hypothetical URF,	unclassified, unknown					ransposon, or	plasmid	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown		unclassified, unknown	Hypothetical ORF,	unclassified, unknown
Dradiated (D)	or Or	Documented	<u>(a)</u>	Operon				٠.	(P)							10/				į	<u>=</u>	1	<u>E</u>				( <u>a</u>	·						
Right	Coordinate				3695846		3697522		3246977		3247320		3247727		3248016	22/8504	9240394			100000	16218/4	00,000	141/183		1417368		526765		527173		527883		528124	
left Coordinate					3695658		3695843		3246594		3247015		3247323		3247717	32/112	7110470			1000001	1620984	4440110	7/00/14		1417192		522485		526805		527173		527864	
Genes On	Operon .				YhjT		Uįhy		Sign	·	ng/b/		yajĒ		YajK	Finn	<i>'h</i>			1120	нап	0.7	SIEB		rajB (b1354)		asur		Jagk		Haly		Dady	
Mole. No.							-		EcXA036											E.YA027	ELAMU3/	CaVA020	ELAAU30				ECXA039							
Gene Prod.	Seq ID No.				330		331		332	*	333		334		335	336	2	_		127	) Co	320	occ	000	338		340				342		343	
GeneSeq	₽;	S.		01	R.		<u> </u>		172		173		b/1	,	1/2	176	)			177	<u> </u>	178	2	SF.	 8/-		<u></u>		 80		791		183	

Prodicted functional alana									N	IVO ASSIGNEO FOIE	No assigned role		Translation				Unknown	Cytochrome D oxidase		Purine biosynthesis		Carbon storage regulator	(mRNA decay factor)	Translation (tRNA)			Fimbrae	Regulator of inversion
Blattner functional class	of encoded proteins		Hypothetical ORF, unclassified, unknown	Phage, transposon, or plasmid	Phage, transposon, or plasmid	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	Unclassified, unknown	inypuneucal unr, unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Translation, post-	translational modification	Translation, post	translational modification	Carbon compound catabolism	Energy metabolism	Energy metabolism	nesis and	metabolism	Regulatory function		Translation, post-	translational modification	Cell structure	Cell structure	Cell structure
Predicted (P)	ō	Documented (D) Operon		(P)				•	idi	1.1			, (P)		<u>G</u>			(0)		(0)		<u>.</u>		Unpredicted		(D)		
Right	Coordinate		528354	351389	3581811	3581085		3580672	1106755	66.0011	1197460		2280821		767183		769834	772249	773404	1191209		2817168		2816667	·	4539127	4540201	4541231
Left Coordinate			528163	351114	351308	3580669		3579494	1196090		1196756		2280537		765207		767201	770678	772265	1189839		2816983		2816575		4538525	4539605	4540683
Genes On	Operon		ylbl	insB_6	insA	yrhA		Zyy	Ujun		ymſE		rp/Y		hrsA		увдВ	судА	судВ	purB		csrA		\\ ser\		fimB	fimE	fimA
Mole. No.				EcXA040					FrXANA1				EcXA042a·b		EcXA043		-			EcXA044		EcXA045				EcXA046		
Gene Prod.	Seq. 1D No.		344	345	346	347		348	349	2	350		351		352		353	354	355	356		357		328		359	360	361
GeneSeq	<u> </u>	No.	184	185	186	187	00,	88	189	3	130		191		192		193	194	195	196		197		861		199	200	201

Predicted functional class	of encoded proteins															Lysis protein								-					UDP galacto-pyranase	mutase	Unknown		i utative auto-transporter
Blattner functional class	of encoded proteins			Cell-structure	Cell structure	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown	Hypothetical ORF,	unclassified, unknown		wn	Cell processes (incl.	Adaptation, protection)	Phage, transposon, or	plasmid	Putative enzymes	Hypothetical ORF,	unclassified, unknown	Putative enzymes	Hypothetical ORF,	unclassified, unknown	Cell structure	·	lown	Hypothetical ORE, U						
Predicted (P)		Documented (D)	Uperon							(P)								( <u>a</u> )		æ				·		-					<u> </u>	(P)	
Right	Coordinate		10077	4241832	4542597	4545301	4545841	4546357	4547279	1638684		1638081		1638389		1638684		1639578		2100933		2101411	2102531		. 2103106	2104079		2105248	2106351		404042	393642	
Left Coordinate			4541100	4041100	4541872	4542665	4545311	4545854	4546377	1637054	-	1637548		1638078	5	1638394		1639363		2099917		2100938	2101413		2102516	2103087		2104082	2105248	100001	776704	392239	
-	Operon		limit		rimC	Qwij	fimF	fimG	fimH	ydfP	1	DJpA		ydfR		SJpA		ядсэ		/ Zci/		YerJ	) het		yetH	) here		311	yefE	c:	Jaic	VaiU	
Mole. No.										EcXA047									7.74040	ECAAU48										E-VADAD	- CEVA043	EcXA050	
Gene Prod.	Seq ID NO.		36.7	363	200	354	365	366	367	368	000	808	076	3/0	120	ر ا ا	27.2	7/6	27.9	2/5	27.4	975	3/3	37.6	370	3//	278	0/0	379	380		381	
GeneSeq	2 2		202	2013	207	502	cn7	206	207	208	000	£0.7	210	710	211	117	212	717	213	ر اع	214	215	617	216	217	/17	218	210	219	220		221	

Predicted functional class	of encoded proteins		Hypothetical outer	membrane protein													Translation	•	Translation					-							Translation	
Blattner functional class	of encoded proteins		Hypothetical ORF,	unclassified, unknown	Translation, post-	translational modification	Translation, post-	translational modification	Translation, post	translational modification	Translation, post-	translational modification	Translation, post	translational modification	Translation, post-	translational modification	Translation, post	translational modification	Translation, post	translational modification												
Predicted (P)	Ur Documented	(D) Operon			<u>(</u> )						;															-	<u> </u>		<del>-</del>			
Right	Coordinate		394353		3446205		3446396		3446806		3447520		3447870		3448163		3449001		3449321		3449923		3450563		3450907		179773		1798023		1798662	
Left Coordinate			393685		3445951		3446205		3446396		3446819		3447538	,	3447885		3448180		3449019		3449318		3449934		3450596		1797417		1797826		1798120	
Genes On			yaiV		Dsds		Эшдэ		dlds	,	.jsdi		Ndr		sdı		пр/В		M/ds		Olds		Jldu		rps/		Ildı		/wds		Jui	
Mole. No.	-				EcXAU51a-b									3													ECXAU52					
Gene Prod.	3		382		283	700	384	700	385	200	200	100	/85	000	388		 585		380		 188		385		383 	100	485	700	CR2	200	390	-
GeneSeq	No.		222		677	224	b77	300	677	200	077	200	/77	900	977	200	R77	000	ne7		F7		767	200	733		<b>567</b>	206	667	300	720	1

Predicted functional class of encoded proteins		Glutathione oxido reductase			Translation (rRNA)	Translation (rRNA)	Translation (tRNA)	Translation (rRNA)
Blattner functional class of encoded proteins	Translation, post- translational modification	Biosynthesis of cofactors,	carriers		Translation, post	Translation, post- translation, post-	Translation, post- translational modification	Translation, post- translational modification
Predicted (P) Or Documented (D)		(P)			(0)			
Right Coordinate	1800594	3645281	•		2727204	2724208	2727464	2729178
Left Coordinate	1798666	3643929		\rightarrow **	2724301	2724089	2727389	2727636
Genes On Operon	thrS	gor		Same operon as EcXA031	gju	Эри	MIIB	тѕв
Mole. No.		EcXA053		EcXA054	EcXA055			
Gene Prod. Seq ID No.	397	398			399	400	401	402
GeneSeq ID No.	237	238			239	240	. 241	242

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Several of the expression vectors contain fragments that correspond to genes of unknown function or if the function is known, it is not known whether the gene is essential. For example, EcXADD1, 003, 007, 008, 013, 015, 016, 017, 018, 019, 020, 021, 022, 023, 024, 025, 026, 027, 028, 029, 030, 032, 033, 034, 035, 036, 037, 038, 039, 040, 041, 047, 048, 049 and 050 are all exogenous nucleic acid sequences that correspond to *E. coli* proteins that have no known function or where the function has not been shown to be essential or nonessential.

The present invention reports a number of novel *E. coli* genes and operons that are required for proliferation. From the list clone sequences identified here, each was identified to be a portion of a gene in an operon required for the proliferation of *E. coli*. Cloned sequences corresponding to genes already known to be required for proliferation in *E. coli* include EcXA002, 004, 005, 010, 012, 014, 031, 02, 043, 045, 051, 052, 054, and 055. The remaining identified sequences correspond to *E. coli* genes previously undesignated as required for proliferation in the art.

An interesting observation of the present invention is that there are also several sequence fragments that correspond to *E. coli* genes that are not thought to be required for *E. coli* proliferation. Nevertheless, under the conditions described above, the antisense expression of these gene fragments causes a reduction in cell growth. This result implies that the genes corresponding to the identified sequences are actually required for proliferation. Molecule Nos. corresponding to these genes are EcXAOO6, 044, 046, and 053.

Following identification of the sequences of interest, these sequences were localized into operons. Since bacterial genes are expressed in a polycistronic manner, the antisense inhibition of a single gene in an operon might effect the expression of all the other genes on the operon or the genes down stream from the single gene identified. In order to determine which of the gene products in an operon are required for proliferation, each of the genes contained within an operon may be analyzed for their effect on viability as described below.

TABLE III

# Operon Boundaries

Mole. No.	Left	Right
	Coordinate	Coordinate
EcXA001	3606848	3608143
EcXA002	2702355	
EcXA003	1015762	1017522
EcXA004	3467782	3472189
EcXA005	4177574	4178503
EcXAD06	2412767	2414911
EcXA007	3841591	3843357
EcXA008	3268266	3272548
EcXA009	1599514	1605313
EcXA010	1647406	1647458
EcXA011	1545425	1550015
EcXA012	3467782	3472189
EcXAD13	2697683	2697943
EcXA014	3049135	3053470
EcXA015	2465875	2468482
EcXA016	1877031	1877972
EcXA017	3065360	3066100
EcXA018	2151891	2160901
EcXA019	2185400	2190242
EcXA020	1877031	1877972
EcXA021	238746	239084
EcXA022	1703791	1710182
EcXA023	1585817	1588025
EcXA024	3231369	3232096
EcXA025	2042885	2050036
EcXA026	331001	. 331184
EcXA027c	2225343	2228405
EcXA028	2420669	2421559
EcXA029	4626424	4628091
EcXA030	3718309	3719678
EcXA031	3440255	3445786
EcXA032b	751452	753691
EcXA033	3192961	3198606
EcXA034	3717678	3717890
EcXA035	3694087	3697522
EcXA036	3246594	3248594
EcXA037	1620984	1621874
EcXA038	1416572	1417368
EcXA039	522485	528354
EcXA040	3580669	3580672
EcXA041	1196090	1197460
EcXA042	2280537	2280821

Mole. No.	Left	Right
	Coordinate	Coordinate
EcXA043	765207	773404
EcXA044	1189839	1191209
EcXA045	2816575	2817168
EcXA046	4538525	4547279
EcXA047	1637054	1639578
EcXA048	2099917	2106351
EcXA049	402927	404042
EcXA050	392239	394353
EcXA051	3445951	3450907
EcXA052	1797417	1800594
EcXA053	3643929	3645281
EcXA054	3440255	3445786
EcXA055	2724301	2729178

**EXAMPLE 5** 

# Identification of Individual Genes within an Operon Required for Proliferation

The following example illustrates a method for determining which gene in an operon is required for proliferation. The clone insert corresponding to Molecule No. EcXA004 possesses nucleic acid sequence homology to the *E. coli* genes rspG and rspL. This molecule corresponds to an operon containing two additional genes fusA and tufA. The rpsL gene is the first gene in the operon. To determine which gene or genes in this operon are required for proliferation, each gene is selectively inactivated using homologous recombination. Gene rpsL is the first gene to be inactivated.

Deletion inactivation of a chromosomal copy of a gene in *E. coli* can be accomplished by integrative gene replacement. The principle of this method (Hamilton, C. M., et al 1989. *J. Bacteriol.* 171: 4617-4622) is to construct a mutant allele of the targeted gene, introduce that allele into the chromosome using a conditional suicide vector, and then force the removal of the native wild type allele and vector sequences. This will replace the native gene with a desired mutation(s) but leave promoters, operators, etc. intact. Essentiality of a gene is determined either by deduction from genetic analysis or by conditional expression of a wild type copy of the targeted gene (trans complementation).

The first step is to generate a mutant <code>rpsl</code> allele using PCR amplification. Two sets of PCR primers are chosen to produce a copy of <code>rpsl</code> with a large central deletion to inactivate the gene. In order to eliminate polar effects, it is desirable to construct a mutant allele comprising an in-frame deletion of most or all of the coding region of the <code>rpsl</code> gene. Each set of PCR primers is chosen such that a region flanking the gene to be amplified is sufficiently long to allow recombination (typically at least 500 nucleotides on each side of the deletion). The targeted deletion or mutation will be contained within this fragment. To facilitate cloning of the PCR product, the PCR primers may also contain restriction endonuclease sites found in the cloning region of a conditional knockout vector such as pK03 (Link, et al 1997 J. Bacteriol. 179 (20): 6228-6237). Suitable sites include Notl, Sall, BamHl and Smal. The <code>rpsl</code> gene fragments are produced using standard PCR conditions including, but not limited to, those outlined in the manufacturers directions for the

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Hot Start Taq PCR kit (Qiagen, Inc., Valencia, CA). The PCR reactions will produce two fragments that can be fused together. Alternatively, crossover PCR can be used to generate a desired deletion in one step (Ho, S. N., et al 1989. *Gene* 77: 51-59, Horton, R. M., et al 1989. *Gene* 77: 61-68). The mutant allele thus produced is called a "null" allele because it cannot produce a functional gene product.

The mutant allele obtained from PCR amplification is cloned into the multiple cloning site of pKO3. Directional cloning of the *rpsL* null allele is not necessary. The pKO3 vector has a temperature-sensitive origin of replication derived from pSC101. Therefore, clones are propagated at the permissive temperature of 30°C. The vector also contains two selectable marker genes: one that confers resistance to chloramphenical and another, the *Bacillus subtilis sacB* gene, that allows for counter-selection on sucrose containing growth medium. Clones that contain vector DNA with the null allele inserted are confirmed by restriction endonuclease analysis and DNA sequence analysis of isolated plasmid DNA. The plasmid containing the *rpsL* null allele insert is known as a knockout plasmid.

Once the knockout plasmid has been constructed and its sequence verified, it is transformed into a Rec\* *E. coli* host cell. Transformation can be by any standard method such as electroporation. In some fraction of the transformed cells, plasmids will integrate into the *E. coli* chromosome by homologous recombination between the *rpsL* null allele in the plasmid and the *rpsL* gene in the chromosome. Transformant colonies in which such an event has occurred are readily selected by growth at the non-permissive temperature of 43°C and in the presence of choramphenicol. At this temperature, the plasmid will not replicate as an episome and will be lost from cells as they grow and divide. These cells are no longer resistant to chloramphenicol and will not grow when it is present. However, cells in which the knockout plasmid has integrated into the *E. coli* chromosome remain resistant to chloramphenicol and propagate.

Cells containing integrated knock-out plasmids are usually the result of a single crossover event that creates a tandem repeat of the mutant and native wild type alleles of *rpsl* separated by the vector sequences. A consequence of this is that *rpsl* will still be expressed in these cells. In order to determine if the gene is essential for growth, the wild type copy must be removed. This is accomplished by selecting for plasmid excision, a process in which homologous recombination between the two alleles results in looping out of the plasmid sequences. Cells that have undergone such an excision event and have lost plasmid sequences including *sacB* gene are selected for by addition of sucrose to the medium. The sacB gene product converts sucrose to a toxic molecule. Thus counter selection with sucrose ensures that plasmid sequences are no longer present in the cell. Loss of plasmid sequences is further confirmed by testing for sensitivity to chloramphenicol (loss of the chloramphenicol resistance gene). The latter test is important because occasionally a mutation in the *sacB* gene can occur resulting in a loss of *sacB* function with no effect on plasmid replication (Link, et. al., 1997 *J. Bacteriol.* 179 (20): 6228-6237). These artifact clones retain plasmid sequences and are therefore still resistant to chloramphenicol.

In the process of plasmid excision, one of the two rpsL alleles is lost from the chromosome along with the plasmid DNA. In general, it is equally likely that the null allele or the wild type allele will be lost. Therefore, if the rpsL

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gene is not essential, half of the clones obtained in this experiment will have the wild type allele on the chromosome and half will have the null allele. However, if the rpsl gene is essential, cells containing the null allele will not be obtained as a single copy of the null allele would be lethal.

To determine the essentiality of *rpsl*, a statistically significant number of the resulting clones, at least 20, are analyzed by PCR amplification of the *rpsl* gene. Since the null allele is missing a significant portion of the *rpsl* gene, its PCR product is significantly shorter than that of the wild type gene and the two are readily distinguished by gel electrophoretic analysis. The PCR products may also be subjected to sequence determination for further confirmation by methods well known to those in the art.

The above experiment is generally adequate for determining the essentiality of a gene such as rpsl. However, it may be necessary or desirable to more directly confirm the essentiality of the gene. There are several methods by which this can be accomplished. In general, these involve three steps: 1) construction of an episome containing a wild type allele, 2) isolation of clones containing a single chromosomal copy of the mutant null allele as described above but in the presence of the episomal wild type allele, and then 3) determining if the cells survive when the expression of the episomal allele is shut off. In this case, the trans copy of wild type rpsL is made by PCR cloning of the entire coding region of rpsL and inserting it in the sense orientation downstream of an inducible promoter such as the E. coli lac promoter. Transcription of this allele of rpst will be induced in the presence of IPTG which inactivates the lac repressor. Under IPTG induction rpsL protein will be expressed as long as the recombinant gene also possesses a ribosomal binding site, also known as a "Shine-Dalgarno Sequence". The trans copy of rpsL is cloned on a plasmid that is compatible with pSC101. Compatible vectors include p15A, pBR322, and the pUC plasmids, among others. Replication of the compatible plasmid will not be temperature-sensitive. The entire process of integrating the null allele of rpsL and subsequent plasmid excision is carried out in the presence of IPTG to ensure the expression of functional rpsL protein is maintained throughout. After the null rpsl allele is confirmed as integrated on the chromosome in place of the wild type rpsl allele, then IPTG is withdrawn and expression of functional rpsL protein shut off. If the rpsL gene is essential, cells will cease to proliferate under these conditions. However, if the rpsL gene is not essential, cells will continue to proliferate under these conditions. In this experiment, essentiality is determined by conditional expression of a wild type copy of the gene rather than inability to obtain the intended chromosomal disruption.

An advantage of this method over some other gene disruption techniques is that the targeted gene can be deleted or mutated without the introduction of large segments of foreign DNA. Therefore, polar effects on downstream genes are eliminated or minimized. There are methods described to introduce inducible promoters upstream of potential essential bacterial genes. However in such cases, polarity from multiple transcription start points can be a problem. One way of preventing this is to insert a gene disruption cassette that contains strong transcriptional terminators upstream of the integrated inducible promoter (Zhang, Y, and Cronan, J. E. 1996 J. Bacteriol. 178 (12): 3614-3620). The described techniques will all be familiar to one of ordinary skill in the art.

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Following the analysis of the *rpsL* gene, the other genes of the operon are investigated to determine if they are required for proliferation.

#### **EXAMPLE 6**

# Expression of the Proteins Encoded by Genes Identified as Required for E. coli Proliferation

The following is provided as one exemplary method to express the proliferation-required proteins encoded by the identified sequences described above. First, the initiation and termination codons for the gene are identified. If desired, methods for improving translation or expression of the protein are well known in the art. For example, if the nucleic acid encoding the polypeptide to be expressed lacks a methionine codon to serve as the initiation site, a strong Shine-Delgarno sequence, or a stop codon, these sequences can be added. Similarly, if the identified nucleic acid sequence lacks a transcription termination signal, this sequence can be added to the construct by, for example, splicing out such a sequence from an appropriate donor sequence. In addition, the coding sequence may be operably linked to a strong promoter or an inducible promoter if desired. The identified nucleic acid sequence or portion thereof encoding the polypeptide to be expressed is obtained by PCR from the bacterial expression vector or genome using oligonucleotide primers complementary to the identified nucleic acid sequence or portion thereof and containing restriction endonuclease sequences for *Ncol* incorporated into the 5' primer and *Bg/*II at the 5' end of the corresponding 3'-primer, taking care to ensure that the identified nucleic acid sequence is positioned in frame with the termination signal. The purified fragment obtained from the resulting PCR reaction is digested with *Ncol* and *Bg/*II, purified and ligated to an expression vector.

The ligated product is transformed into DH5 $\alpha$  or some other *E. coli* strain suitable for the over expression of potential proteins. Transformation protocols are well known in the art. For example, transformation protocols are described in: Current Protocols in Molecular Biology, Vol. 1, Unit 1.8, (Ausubel, et al., Eds.) John Wiley & Sons, Inc. (1997). Positive transformants are selected after growing the transformed cells on plates containing 50-100  $\mu$ g/ml Ampicillin (Sigma, St. Louis, Missouri). In one embodiment, the expressed protein is held in the cytoplasm of the host organism. In an alternate embodiment, the expressed protein is released into the culture medium. In still another alternative, the expressed protein can be sequestered in the periplasmic space and liberated therefrom using any one of a number of cell lysis techniques known in the art. For example, the osmotic shock cell lysis method described in Chapter 16 of Current Protocols in Molecular Biology, Vol. 2, (Ausubel, et al., Eds.) John Wiley & Sons, Inc. (1997). Each of these procedures can be used to express a proliferation-required protein.

Expressed proteins, whether in the culture medium or liberated from the periplasmic space or the cytoplasm, are then purified or enriched from the supernatant using conventional techniques such as ammonium sulfate precipitation, standard chromatography, immunoprecipitation, immunochromatography, size exclusion chromatography, ion exchange chromatography, and HPLC. Alternatively, the secreted protein can be in a sufficiently enriched or pure state in the supernatant or growth media of the host to permit it to be used for its intended purpose without further enrichment. The purity of the protein product

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obtained can be assessed using techniques such as Coomassie or silver staining or using antibodies against the control protein.

Coomassie and silver staining techniques are familiar to those skilled in the art.

Antibodies capable of specifically recognizing the protein of interest can be generated using synthetic peptides using methods well known in the art. See, Antibodies: A Laboratory Manual, (Harlow and Lane, Eds.) Cold Spring Harbor Laboratory (1988). For example, 15 mer peptides having a sequence encoded by the appropriate identified gene sequence of interest or portion thereof can be chemically synthesized. The synthetic peptides are injected into mice to generate antibodies to the polypeptide encoded by the identified nucleic acid sequence of interest or portion thereof. Alternatively, samples of the protein expressed from the expression vectors discussed above can be purified and subjected to amino acid sequencing analysis to confirm the identity of the recombinantly expressed protein and subsequently used to raise antibodies. An Example describing in detail the generation of monoclonal and polyclonal antibodies appears in Example 7.

The protein encoded by the identified nucleic acid sequence of interest or portion thereof can be purified using standard immunochromatography techniques. In such procedures, a solution containing the secreted protein, such as the culture medium or a cell extract, is applied to a column having antibodies against the secreted protein attached to the chromatography matrix. The secreted protein is allowed to bind the immunochromatography column. Thereafter, the column is washed to remove non-specifically bound proteins. The specifically bound secreted protein is then released from the column and recovered using standard techniques. These procedures are well known in the art.

In an alternative protein purification scheme, the identified nucleic acid sequence of interest or portion thereof can be incorporated into expression vectors designed for use in purification schemes employing chimeric polypeptides. In such strategies the coding sequence of the identified nucleic acid sequence of interest or portion thereof is inserted in-frame with the gene encoding the other half of the chimera. The other half of the chimera can be maltose binding protein (MBP) or a nickel binding polypeptide encoding sequence. A chromatography matrix having antibody to MBP or nickel attached thereto is then used to purify the chimeric protein. Protease cleavage sites can be engineered between the MBP gene or the nickel binding polypeptide and the identified expected gene of interest, or portion thereof. Thus, the two polypeptides of the chimera can be separated from one another by protease digestion.

One useful expression vector for generating maltose binding protein fusion proteins is pMAL (New England Biolabs), which encodes the *malE* gene. In the pMal protein fusion system, the cloned gene is inserted into a pMal vector downstream from the *malE* gene. This results in the expression of an MBP-fusion protein. The fusion protein is purified by affinity chromatography. These techniques as described are well known to those skilled in the art of molecular biology.

#### **EXAMPLE 7**

### Production of an Antibody to an isolated E. coli Protein

Substantially pure protein or polypeptide is isolated from the transformed cells as described in Example 6. The concentration of protein in the final preparation is adjusted, for example, by concentration on a 10,000 molecular weight cut off

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AMICON filter device (Millipore, Bedford, MA), to the level of a few micrograms/ml. Monoclonal or polyclonal antibody to the protein can then be prepared as follows:

## Monoclonal Antibody Production by Hybridoma Fusion

Monoclonal antibody to epitopes of any of the peptides identified and isolated as described can be prepared from murine hybridomas according to the classical method of Kohler, G. and Milstein, C., Nature 256:495 (1975) or any of the well-known derivative methods thereof. Briefly, a mouse is repetitively inoculated with a few micrograms of the selected protein or peptides derived therefrom over a period of a few weeks. The mouse is then sacrificed, and the antibody producing cells of the spleen isolated. The spleen cells are fused by means of polyethylene glycol with mouse myeloma cells, and the excess unfused cells destroyed by growth of the system on selective media comprising aminopterin (HAT media). The successfully fused cells are diluted and aliquots of the dilution placed in wells of a microtiter plate where growth of the culture is continued. Antibody-producing clones are identified by detection of antibody in the supernatant fluid of the wells by immunoassay procedures, such as ELISA, as described by Engvall, E., "Enzyme immunoassay ELISA and EMIT," Meth. Enzymol. 70:419 (1980), and derivative methods thereof. Selected positive clones can be expanded and their monoclonal antibody product harvested for use. Detailed procedures for monoclonal antibody production are described in Davis, L. et al. Basic Methods in Molecular Biology Elsevier, New York. Section 21-2.

## Polyclonal Antibody Production by Immunization

Polyclonal antiserum containing antibodies to heterogeneous epitopes of a single protein or a peptide can be prepared by immunizing suitable animals with the expressed protein or peptides derived therefrom described above, which can be unmodified or modified to enhance immunogenicity. Effective polyclonal antibody production is affected by many factors related both to the antigen and the host species. For example, small molecules tend to be less immunogenic than larger molecules and can require the use of carriers and adjuvant. Also, host animals vary in response to site of inoculations and dose, with both inadequate or excessive doses of antigen resulting in low titer antisera. Small doses (ng level) of antigen administered at multiple intradermal sites appears to be most reliable. An effective immunization protocol for rabbits can be found in Vaitukaitis, J. et al. J. Clin. Endocrinol. Metab. 33:988-991 (1971).

Booster injections can be given at regular intervals, and antiserum harvested when antibody titer thereof, as determined semi-quantitatively, for example, by double immunodiffusion in agar against known concentrations of the antigen, begins to fall. See, for example, Ouchterlony, O. et al., Chap. 19 in: Handbook of Experimental Immunology D. Wier (ed) Blackwell (1973). Plateau concentration of antibody is usually in the range of 0.1 to 0.2 mg/ml of serum (about 12 M). Affinity of the antisera for the antigen is determined by preparing competitive binding curves, as described, for example, by Fisher, D., Chap. 42 in: Manual of Clinical Immunology, 2d Ed. (Rose and Friedman, Eds.) Amer. Soc. For Microbiol., Washington, D.C. (1980).

Antibody preparations prepared according to either protocol are useful in quantitative immunoassays which determine concentrations of antigen-bearing substances in biological samples; they are also used semi-quantitatively or qualitatively to



identify the presence of antigen in a biological sample. The antibodies can also be used in therapeutic compositions for killing bacterial cells expressing the protein.

### **EXAMPLE 8**

## Screening Chemical Libraries

## A. Protein-Based Assays

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Having isolated and expressed bacterial proteins shown to be required for bacterial proliferation, the present invention further contemplates the use of these expressed proteins in assays to screen libraries of compounds for potential drug candidates. The generation of chemical libraries is well known in the art. For example combinatorial chemistry can be used to generate a library of compounds to be screened in the assays described herein. A combinatorial chemical library is a collection of diverse chemical compounds generated by either chemical synthesis or biological synthesis by combining a number of chemical "building blocks" reagents. For example, a linear combinatorial chemical library such as a polypeptide library is formed by combining amino acids in every possible combination to yield peptides of a given length. Millions of chemical compounds theoretically can be synthesized through such combinatorial mixings of chemical building blocks. For example, one commentator observed that the systematic, combinatorial mixing of 100 interchangeable chemical building blocks results in the theoretical synthesis of 100 million tetrameric compounds or 10 billion pentameric compounds. (Gallop et al., "Applications of Combinatorial Technologies to Drug Discovery, Background and Peptide Combinatorial Libraries," Journal of Medicinal Chemistry, Vol. 37, No. 9, 1233-1250 (1994). Other chemical libraries known to those in the art may also be used, including natural product libraries.

Once generated, combinatorial libraries can be screened for compounds that possess desirable biological properties. For example, compounds which may be useful as drugs or to develop drugs would likely have the ability to bind to the target protein identified, expressed and purified as discussed above. Further, if the identified target protein is an enzyme, candidate compounds would likely interfere with the enzymatic properties of the target protein. Any enzyme can be a target protein. For example, the enzymatic function of a target protein can be to serve as a protease, nuclease, phosphatase, dehydrogenase, transporter protein, transcriptional enzyme, and any other type of enzyme known or unknown. Thus, the present invention contemplates using the protein products described above to screen combinatorial chemical libraries.

Those in the art will appreciate that a number of techniques exist for characterizing target proteins in order to identify molecules useful for the discovery and development of therapeutics. For example, some techniques involve the generation and use of small peptides to probe and analyze target proteins both biochemically and genetically in order to identify and develop drug leads. Such techniques include the methods described in PCT publications No. W09935494, W09819162, W09954728, the disclosures of which are incorporated herein by reference in their entireties.

In another example, the target protein is a serine protease and the substrate of the enzyme is known. The present example is directed towards the analysis of libraries of compounds to identify compounds that function as inhibitors of the target enzyme. First, a library of small molecules is generated using methods of combinatorial library formation well known in

the art. U.S. Patent NOs. 5,463,564 and 5,574, 656, to Agrafiotis, et al., entitled "System and Method of Automatically Generating Chemical Compound with Desired Properties," are two such teachings. Then the library compounds are screened to identify library compounds that possess desired structural and functional properties. U.S. Patent No. 5,684,711 also discusses a method for screening libraries.

To illustrate the screening process, the combined target and chemical compounds of the library are exposed to and permitted to interact with the purified enzyme. A labeled substrate is added to the incubation. The label on the substrate is such that a detectable signal is emitted from metabolized substrate molecules. The emission of this signal permits one to measure the effect of the combinatorial library compounds on the enzymatic activity of target enzymes. The characteristics of each library compound is encoded so that compounds demonstrating activity against the enzyme can be analyzed and features common to the various compounds identified can be isolated and combined into future iterations of libraries.

Once a library of compounds is screened, subsequent libraries are generated using those chemical building blocks that possess the features shown in the first round of screen to have activity against the target enzyme. Using this method, subsequent iterations of candidate compounds will possess more and more of those structural and functional features required to inhibit the function of the target enzyme, until a group of enzyme inhibitors with high specificity for the enzyme can be found. These compounds can then be further tested for their safety and efficacy as antibiotics for use in mammals.

It will be readily appreciated that this particular screening methodology is exemplary only. Other methods are well known to those skilled in the art. For example, a wide variety of screening techniques are known for a large number of naturally-occurring targets when the biochemical function of the target protein is known.

#### B. Cell Based Assays

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Current cell-based assays used to identify or to characterize compounds for drug discovery and development frequently depend on detecting the ability of a test compound to inhibit the activity of a target molecule located within a cell or located on the surface of a cell. Most often such target molecules are proteins such as enzymes, receptors and the like. However, target molecules may also include other molecules such as DNAs, lipids, carbohydrates and RNAs including messenger RNAs, ribosomal RNAs, tRNAs and the like. A number of highly sensitive cell-based assay methods are available to those of skill in the art to detect binding and interaction of test compounds with specific target molecules. However, these methods are generally not highly effective when the test compound binds to or otherwise interacts with its target molecule with moderate or low affinity. In addition, the target molecule may not be readily accessible to a test compound in solution, such as when the target molecule is located inside the cell or within a cellular compartment such as the periplasm of a bacterial cell. Thus, current cell-based assay methods are limited in that they are not effective in identifying or characterizing compounds that interact with their targets with moderate to low affinity or compounds that interact with targets that are not readily accessible.

. Cell-based assay methods of the present invention have substantial advantages over current cell-based assays practiced in the art. These advantages derive from the use of sensitized cells in which the level or activity of a

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proliferation-required gene product (the target molecule) has been specifically reduced to the point where the presence or absence of its function becomes a rate-determining step for cellular proliferation. Bacterial, fungal, plant, or animal cells can all be used with the present method. Such sensitized cells become much more sensitive to compounds that are active against the affected target molecule. Thus, cell-based assays of the present invention are capable of detecting compounds exhibiting low or moderate potency against the target molecule of interest because such compounds are substantially more potent on sensitized cells than on non-sensitized cells. The affect may be such that a test compound may be two to several times more potent, at least 10 times more potent or even at least 100 times more potent when tested on the sensitized cells as compared to the non-sensitized cells.

Due in part to the increased appearance of antibiotic resistance in pathogenic microorganisms and to the significant side-effects associated with some currently used antibiotics, novel antibiotics acting at new targets are highly sought after in the art. Yet, another limitation in the current art related to cell-based assays is the problem of identifying hits against the same kinds of target molecules in the same limited set of biological pathways over and over again. This may occur when compounds acting at such new targets are discarded, ignored or fail to be detected because compounds acting at the "old" targets are encountered more frequently and are more potent than compounds acting at the new targets. As a result, the majority of antibiotics in use currently interact with a relatively small number of target molecules within an even more limited set of biological pathways.

The use of sensitized cells of the current invention provides a solution to the above problem in two ways. First, desired compounds acting at a target of interest, whether a new target or a previously known but poorly exploited target, can now be detected above the "noise" of compounds acting at the "old" targets due to the specific and substantial increase in potency of such desired compounds when tested on the sensitized cells of the current invention. Second, the methods used to sensitize cells to compounds acting at a target of interest may also sensitize these cells to compounds acting at other target molecules within the same biological pathway. For example, expression of an antisense molecule to a gene encoding a ribosomal protein is expected to sensitize the cell to compounds acting at that ribosomal protein and may also sensitize the cells to compounds acting at any of the ribosomal components (proteins or rRNA) or even to compounds acting at any target which is part of the protein synthesis pathway. Thus an important advantage of the present invention is the ability to reveal new targets and pathways that were previously not readily accessible to drug discovery methods.

Sensitized cells of the present invention are prepared by reducing the activity or level of a target molecule. The target molecule may be a gene product, such as an RNA or polypeptide produced from the proliferation-required nucleic acids described herein. Alternatively, the target may be a gene product such as an RNA or polypeptide which is produced form a sequence within the same operon as the proliferation-required nucleic acids described herein. In addition, the target may be an RNA or polypeptide in the same biological pathway as the proliferation-required nucleic acids described herein.

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Such biological pathways include, but are not limited to, enzymatic, biochemical and metabolic pathways as well as pathways involved in the production of cellular structures such the cell wall.

Current methods employed in the arts of medicinal and combinatorial chemistries are able to make use of structure-activity relationship information derived from testing compounds in various biological assays including direct binding assays and cell-based assays. Occasionally compounds are directly identified in such assays that are sufficiently potent to be developed as drugs. More often, initial hit compounds exhibit moderate or low potency. Once a hit compound is identified with low or moderate potency, directed libraries of compounds are synthesized and tested in order to identify more potent leads. Generally these directed libraries are combinatorial chemical libraries consisting of compounds with structures related to the hit compound but containing systematic variations including additions, subtractions and substitutions of various structural features. When tested for activity against the target molecule, structural features are identified that either alone or in combination with other features enhance or reduce activity. This information is used to design subsequent directed libraries containing compounds with enhanced activity against the target molecule. After one or several iterations of this process, compounds with substantially increased activity against the target molecule are identified and may be further developed as drugs. This process is facilitated by use of the sensitized cells of the present invention since compounds acting at the selected targets exhibit increased potency in such cell-based assays, thus; more compounds can now be characterized providing more useful information than would be obtained otherwise.

Thus, it is now possible using cell-based assays of the present invention to identify or characterize compounds that previously would not have been readily identified or characterized including compounds that act at targets that previously were not readily exploited using cell-based assays. The process of evolving potent drug leads from initial hit compounds is also substantially improved by the cell-based assays of the present invention because, for the same number of test compounds, more structure-function relationship information is likely to be revealed.

The method of sensitizing a cell entails selecting a suitable gene or operon. A suitable gene or operon is one whose expression is required for the proliferation of the cell to be sensitized. The next step is to introduce into the cells to be sensitized, an antisense RNA capable of hybridizing to the suitable gene or operon or to the RNA encoded by the suitable gene or operon. Introduction of the antisense RNA can be in the form of an expression vector in which antisense RNA is produced under the control of an inducible promoter. The amount of antisense RNA produced is limited by varying the inducer concentration to which the cell is exposed and thereby varying the activity of the promoter driving transcription of the antisense RNA. Thus, cells are sensitized by exposing them to an inducer concentration that results in a sub-lethal level of antisense RNA expression.

In one embodiment of the cell-based assays, the identified exogenous *E. coli* nucleotide sequences of the present invention are used to inhibit the production of a proliferation required protein. Expression vectors producing antisense RNA against identified genes required for proliferation are used to limit the concentration of a proliferation-required protein without severly inhibiting growth. To achieve that goal, a growth inhibition dose curve of inducer is calculated by plotting

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various doses of inducer against the corresponding growth inhibition caused by the antisense expression. From this curve, various percentages of antisense induced growth inhibition, from 1 to 100% can be determined. If the promoter contained in the expression vector contains a *lac* operator the transcription is regulated by *lac* repressor and expression from the promoer is inducible with IPTG. For example, the highest concentration of the inducer IPTG that does not reduce the growth rate (0% growth inhibition) can be predicted from the curve. Cellular proliferation can be monitored by growth medium turbidity via OD measurements. In another example, the concentration of inducer that reduces growth by 25% can be predicted from the curve. In still another example, a concentration of inducer that reduces growth by 50% can be calculated. Additional parameters such as colony forming units (cfu) can be used to measure cellular viability.

Cells to be assayed are exposed to the above-determined concentrations of inducer. The presence of the inducer at this sub-lethal concentration reduces the amount of the proliferation required gene product to the lowest amount in the cell that will support growth. Cells grown in the presence of this concentration of inducer are therefore specifically more sensitive to inhibitors of the proliferation-required protein or RNA of interest or to inhibitors of proteins or RNAs in the same biological pathway as the proliferation-required protein or RNA of interest but not to inhibitors of unrelated proteins or RNAs.

Cells pretreated with sub-inhibitory concentrations of inducer and thus containing a reduced amount of proliferation-required target gene product are then used to screen for compounds that reduce cell growth. The sub-lethal concentration of inducer may be any concentration consistent with the intended use of the assay to identify candidate compounds to which the cells are more sensitive. For example, the sub-lethal concentration of the inducer may be such that growth inhibition is at least about 5%, at least about 8%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60% at least about 75%, or more. Cells which are presensitized using the preceding method are more sensitive to inhibitors of the target protein because these cells contain less target protein to inhibit than wild-type cells.

In another embodiment of the cell based assays of the present invention, the level or activity of a proliferation required gene product is reduced using a temperature sensitive ...mutation in the proliferation-required sequence and an antisense nucleic acid against the proliferation-required sequence. Growing the cells at an intermediate temperature between the permissive and restrictive temperatures of the temperature sensitive mutant where the mutation is in a proliferation-required gene products. The antisense RNA directed against the proliferation-required sequence further reduces the activity of the proliferation required gene product. Drugs that may not have been found using either the temperature sensitive mutation or the antisense nucleic acid alone may be identified by determining whether cells in which expression of the antisense nucleic acid has been induced and which are grown at a temperature between the permissive temperature and the restrictive temperature are substantially more sensitive to a test compound than cells in which expression of the antisense nucleic acid has not been induced and which are grown at a permissive temperature. Also drugs found previously from either the antisense nucleic acid alone or the

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temperature sensitive mutation alone may have a different sensitivity profile when used in cells combining the two approaches, and that sensitivity profile may indicate a more specific action of the drug in inhibiting one or more activities of the gene product.

Temperature sensitive mutations may be located at different sites within the gene and correspond to different domains of the protein. For example, the dnaB gene of Escherichia coli encodes the replication fork DNA helicase. DnaB has several domains, including domains for oligomerization, ATP hydrolysis, DNA binding, interaction with primase, interaction with DnaC, and interaction with DnaA [(Biswas, E.E. and Biswas, S.B. 1999. Mechanism and DnaB helicase of Escherichia coli: structural domains involved in ATP hydrolysis, DNA binding, and oligomerization. Biochem. 38:10919-10928; Hiasa, H. and Marians, K.J. 1999. Initiation of bidirectional replication at the chromosomal origin is directed by the interaction between helicase and primase. J. Biol. Chem. 274:27244-27248; San Martin, C., Radermacher, M., Wolpensinger, B., Engel, A., Miles, C.S., Dixon, N.E., and Carazo, J.M. 1998. Three-dimensional reconstructions from cryoelectron microscopy images reveal an intimate complex between helicase DnaB and its loading partner DnaC. Structure 6:501-9; Sutton, M.D., Carr, K.M., Vicente, M., and Kaguni, J.M. 1998. Escherichia coli DnaA protein. The Nterminal domain and loading of DnaB helicase at the E. coli chromosomal. J. Biol. Chem. 273:34255-62.), the disclosures of which are incorporated herein by reference in their entireties). Temperature sensitive mutations in different domains of DnaB confer different phenotypes at the restrictive temperature, which include either an abrupt stop or slow stop in DNA replication with or without DNA breakdown (Wechsler, J.A. and Gross, J.D. 1971. Escherichia coli mutants temperaturesensitive for DNA synthesis. Mol. Gen. Genetics 113:273-284, the disclosure of which is incorporated herein by reference in its entirety) and termination of growth or cell death. Combining the use of temperature sensitive mutations in the dnaB gene that cause cell death at the restrictive temperature with an antisense to the dnaB gene could lead to the discovery of very specific and effective inhibitors of one or a subset of activities exhibited by DnaB.

When screening for antimicrobial agents against a gene product required for proliferation, growth inhibition of cells containing a limiting amount of that proliferation-required gene product can be assayed. Growth inhibition can be measured by directly comparing the amount of growth, measured by the optical density of the growth medium, between an experimental sample and a control sample. Alternative methods for assaying cell proliferation include measuring green fluorescent protein (GFP) reporter construct emissions, various enzymatic activity assays, and other methods well known in the art.

It will be appreciated that the above method may be performed in solid phase, liquid phase or a combination of the two. For example, cells grown on nutrient agar containing the inducer of the antisense construct may be exposed to compounds spotted onto the agar surface. A compound's effect may be judged from the diameter of the resulting killing zone, the area around the compound application point in which cells do not grow. Multiple compounds may be transferred to agar plates and simultaneously tested using automated and semi-automated equipment including but not restricted to

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multi-channel pipettes (for example the Beckman Multimek) and multi-channel spotters (for example the Genomic Solutions Flexys). In this way multiple plates and thousands to millions of compounds may be tested per day.

The compounds may also be tested entirely in liquid phase using microtiter plates as described below. Liquid phase screening may be performed in microtiter plates containing 96, 384, 1536 or more wells per microtiter plate to screen multiple plates and thousands to millions of compounds per day. Automated and semi-automated equipment may be used for addition of reagents (for example cells and compounds) and determination of cell density.

## **EXAMPLE 9**

The effectiveness of the above cell based assay was validated using constructs expressing antisense RNA to *E. coli* genes rplL, rplJ, and rplW encoding ribosomal proteins L7/L12, L10 and L23 respectively. These proteins are part of the protein synthesis apparatus of the cell and as such are required for proliferation. These constructs were used to test the effect of antisense expression on cell sensitivity to antibiotics known to bind to the ribosome and thereby inhibit protein synthesis. Constructs expressing antisense RNA to several other genes (elaD, visC, yohH, and aptE/B), the products of which are not involved in protein synthesis were used for comparison.

First expression vectors containing antisense constructs to either rpIW or to elaD were introduced into separate *E. coli* cell populations. Vector introduction is a technique well known to those of ordinary skill in the art. The expression vectors of this example contain IPTG inducible promoters that drive the expression of the antisense RNA in the presence of the inducer. However, those skilled in the art will appreciate that other inducible promoters may also be used. Suitable expression vectors are also well known in the art. The *E. coli* antisense clones encoding ribosomal proteins L7/L12, L10 and L23 were used to test the effect of antisense expression on cell sensitivity to the antibiotics known to bind to these proteins. First, expression vectors containing antisense to either the genes encoding L7/L12 and L10 or L23 were introduced into separate E. coli cell populations.

The cell populations were exposed to a range of IPTG concentrations in liquid medium to obtain the growth inhibitory dose curve for each clone (Fig. 1). First, seed cultures were grown to a particular turbidity that is measured by the optical density (OD) of the growth solution. The OD of the solution is directly related to the number of bacterial cells contained therein. Subsequently, sixteen 200 ul liquid medium cultures were grown in a 96 well microtiter plate at 37 C with a range of IPTG concentrations in duplicate two-fold serial dilutions from 1600 uM to 12.5 uM (final concentration). Additionally, control cells were grown in duplicate without IPTG. These cultures were started from equal amounts of cells derived from the same initial seed culture of a clone of interest. The cells were grown for up to 15 hours and the extent of growth was determined by measuring the optical density of the cultures at 600 nm. When the control culture reached midlog phase the percent growth of the control for each of the IPTG containing cultures was plotted against the log concentrations of IPTG to produce a growth inhibitory dose response curve for the IPTG. The concentration of IPTG that inhibits cell growth to 50% (IC<sub>50</sub>) as compared to the 0 mM IPTG control (0% growth inhibition) was then calculated from

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the curve. Under these conditions, an amount of antisense RNA was produced that reduced the expression levels of rpIW and elaD to a degree such that growth was inhibited by 50%.

Alternative methods of measuring growth are also contemplated. Examples of these methods include measurements of proteins, the expression of which is engineered into the cells being tested and can readily be measured. Examples of such proteins include green fluorescent protein (GFP) and various enzymes.

Cells were pretreated with the selected concentration of IPTG and then used to test the sensitivity of cell populations to tetracycline, erythromycin and other protein synthesis inhibitors. An example of a tetracycline dose response curve is shown in Figures 2A and 2B for the rpIW and elaD genes, respectively. Cells were grown to log phase and then diluted into media alone or media containing IPTG at concentrations which give 20% and 50% growth inhibition as determined by IPTG dose response curves. After 2.5 hours, the cells were diluted to a final OD600 of 0.002 into 96 well plates containing (1) +/- IPTG at the same concentrations used for the 2.5 hour pre-incubation; and (2) serial two-fold dilutions of tetracycline such that the final concentrations of tetracycline range from 1 µg/ml to 15.6 ng/ml and 0 µg/ml. The 96 well plates were incubated at 37°C and the OD600 was read by a plate reader every 5 minutes for up to 15 hours. For each IPTG concentration and the no IPTG control, tetracycline dose response curves were determined when the control (absence of tetracycline) reached 0.1 0D600. To compare tetracycline sensitivity with and without IPTG, tetracycline IC50s were determined from the dose response curves (Figs. 2A-B). Cells with reduced levels of L23 (rpIW) showed increased sensitivity to tetracycline (Fig. 2A) as compared to cells with reduced levels of elaD (Fig. 2B). Figure 3 shows a summary bar chart in which the ratios of tetracycline IC50s determined in the presence of IPTG which gives 50% growth inhibition versus tetracycline IC50s determined without IPTG (fold increase in tetracycline sensitivity) were plotted. Cells with reduced levels of either L7/L12 (genes rp/L, rp/J) or L23 (rp/W) showed increased sensitivity to tetracycline (Fig. 3). Cells expressing antisense to genes not known to be involved in protein synthesis (atpB/E, visC, elaD, yohH) did not show the same increased sensitivity to tetracycline, validating the specificity of this assay (Fig. 3).

In addition to the above, it has been observed in initial experiments that clones expressing antisense RNA to genes involved in protein synthesis (including genes encoding ribosomal proteins L7/L12 & L10, L7/L12 alone, L22, and L18, as well as genes encoding rRNA and Elongation Factor G) have increased sensitivity to the macrolide, erythromycin, whereas clones expressing antisense to the non-protein synthesis genes elaD, atpB/E and visC do not. Furthermore, the clone expressing antisense to rplL and rplJ does not show increased sensitivity to nalidixic acid and ofloxacin, antibiotics which do not inhibit protein synthesis.

The results with the ribosomal protein genes rplL, rplJ, and rplW as well as the initial results using various other antisense clones and antibiotics show that limiting the concentration of an antibiotic target makes cells more sensitive to the antimicrobial agents that specifically interact with that protein. The results also show that these cells are sensitized to antimicrobial agents that inhibit the overall function in which the protein target is involved but are not sensitized to antimicrobial agents that inhibit other functions.

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The cell based assay described above may also be used to identify the biological pathway in which a proliferation-required nucleic acid or its gene product lies. In such methods, cells expressing a sub-lethal level of antisense to a target proliferation-required nucleic acid and control cells in which expression of the antisense has not been induced are contacted with a panel of antibiotics known to act in various pathways. If the antibiotic acts in the pathway in which the target proliferation-required nucleic acid or its gene product lies, cells in which expression of the antisense has been induced will be more sensitive to the antibiotic than cells in which expression of the antisense has not been induced.

As a control, the results of the assay may be confirmed by contacting a panel of cells expressing antisense nucleic acids to many different proliferation-required genes including the target proliferation-required gene. If the antibiotic is acting specifically, heightened sensitivity to the antibiotic will be observed only in the cells expressing antisense to a target proliferation-required gene (or cells expressing antisense to other proliferation-required genes in the same pathway as the target proliferation-required gene) but will not be observed generally in all cells expressing antisense to proliferation-required genes.

Similarly, the above method may be used to determine the pathway on which a test antibiotic acts. A panel of cells, each of which expresses antisense to a proliferation-required nucleic acid in a known pathway, is contacted with a compound for which it is desired to determine the pathway on which it acts. The sensitivity of the panel of cells to the test compound is determined in cells in which expression of the antisense has been induced and in control cells in which expression of the antisense has not been induced. If the test antibiotic acts on the pathway on which an antisense nucleic acid acts, cells in which expression of the antisense has been induced will be more sensitive to the antibiotic than cells in which expression of the antisense has not been induced. In addition, control cells in which expression of antisense to proliferation-required genes in other pathways has been induced will not exhibit heightened sensitivity to the antibiotic. In this way, the pathway on which the test antibiotic acts may be determined.

The Example below provides one method for performing such assays.

### EXAMPLE 10

# Identification of the Pathway in which a Proliferation-Required

## Gene Lies or the Pathway on which an Antibiotic Acts

# A. Preparation of Bacterial Stocks for Assay

To provide a consistent source of cells to screen, frozen stocks of host bacteria containing the desired antisense construct are prepared using standard microbiological techniques. For example, a single clone of the organism can be isolated by streaking out a sample of the original stock onto an agar plate containing nutrients for cell growth and an antibiotic for which the antisense construct contains a gene which confers resistance. After overnight growth an isolated colony is picked from the plate with a sterile needle and transferred to an appropriate liquid growth media containing the antibiotic required for maintenance of the plasmid. The cells are incubated at 30°C to 37°C with vigorous shaking for 4 to

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6 hours to yield a culture in exponential growth. Sterile glycerol is added to 15% (volume to volume) and 100 $\mu$ L to 500  $\mu$ L aliquots are distributed into sterile cryotubes, snap frozen in liquid nitrogen, and stored at -80°C for future assays.

## B. Growth of Bacteria for Use in the Assay

A day prior to an assay, a stock vial is removed from the freezer, rapidly thawed (37°C water bath) and a loop of culture is streaked out on an agar plate containing nutrients for cell growth and an antibiotic to which the antisense construct confers resistance. After overnight growth at 37°C, ten randomly chosen, isolated colonies are transferred from the plate (sterile inoculum loop) to a sterile tube containing 5 mL of LB medium containing the antibiotic to which the antisense vector confers resistance. After vigorous mixing to form a homogeneous cell suspension, the optical density of the suspension is measured at 600 nm (0D600) and if necessary an aliquot of the suspension is diluted into a second tube of 5 mL, sterile, LB medium plus antibiotic to achieve an  $0D600 \le 0.02$  absorbance units. The culture is then incubated at 37° C for 1-2 hrs with shaking until the 0D600 reaches 0D 0.2 - 0.3. At this point the cells are ready to be used in the assay.

## C. Selection of Media to be Used in Assay

Two fold dilution series of the inducer are generated in culture media containing the appropriate antibiotic for maintenance of the antisense construct. Several media are tested side by side and three to four wells are used to evaluate the effects of the inducer at each concentration in each media. For example, M9 minimal media, L8 broth, T8D broth and Muller-Hinton media may be tested with the inducer IPTG at the following concentrations, 50 μΜ, 100 μΜ, 200 μΜ, 400 μΜ, 600 μΜ, 800 μΜ and 1000 μΜ. Equal volumes of test media-inducer and cells are added to the wells of a 384 well microtiter plate and mixed. The cells are prepared as described above and diluted 1:100 in the appropriate media containing the test antibiotic immediately prior to addition to the microtiter plate wells. For a control, cells are also added to several wells of each media that do not contain inducer, for example 0 M IPTG. Cell growth is monitored continuously by incubation at 37°C in a microtiter plate reader monitoring the 0D600 of the wells over an 18-hour period. The percent inhibition of growth produced by each concentration of inducer is calculated by comparing the rates of logarithmic growth against that exhibited by cells growing in media without inducer. The medium yielding greatest sensitivity to inducer is selected for use in the assays described below.

# D. Measurement of Test Antibiotic Sensitivity in the Absence of Antisense Construct Induction

Two-fold dilution series of antibiotics of known mechanism of action are generated in the culture media selected for further assay development that has been supplemented with the antibiotic used to maintain the construct. A panel of test antibiotics known to act on different pathways is tested side by side with three to four wells being used to evaluate the effect of a test antibiotic on cell growth at each concentration. Equal volumes of test antibiotic and cells are added to the wells of a 384 well microtiter plate and mixed. Cells are prepared as described above using the media selected for assay development supplemented with the antibiotic required to maintain the antisense construct and are diluted 1:100 in identical media immediately prior to addition to the microtiter plate wells. For a control, cells are also added to several

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wells that contain the solvent used to dissolve the antibiotics but no antibiotic. Cell growth is monitored continuously by incubation at  $37^{\circ}$ C in a microtiter plate reader monitoring the OD600 of the wells over an 18-hour period. The percent inhibition of growth produced by each concentration of antibiotic is calculated by comparing the rates of logarithmic growth against that exhibited by cells growing in media without antibiotic. A plot of percent inhibition against log[antibiotic concentration] allows extrapolation of an  $IC_{50}$  value for each antibiotic.

# E. Measurement of Test Antibiotic Sensitivity in the Presence of Antisense Construct Inducer

The culture media selected for use in the assay is supplemented with inducer at concentrations shown to inhibit cell growth by 50 and 80% as described above and the antibiotic used to maintain the construct. Two fold dilution series of the panel of test antibiotics used above are generated in each of these media. Several antibiotics are tested side by side with three to four wells being used to evaluate the effects of an antibiotic on cell growth at each concentration, in each media. Equal volumes of test antibiotic and cells are added to the wells of a 384 well microtiter plate and mixed. Cells are prepared as described above using the media selected for use in the assay supplemented with the antibiotic required to maintain the antisense construct. The cells are diluted 1:100 into two 50 mL aliquots of identical media containing concentrations of inducer that have been shown to inhibit cell growth by 50% and 80 % respectively and incubated at 37°C with shaking for 2.5 hours. Immediately prior to addition to the microtiter plate wells, the cultures are adjusted to an appropriate  $OD_{600}$  (typically 0.002) by dilution into warm (37°C) sterile media supplemented with identical concentrations of the inducer and antibiotic used to maintain the antisense construct. For a control, cells are also added to several wells that contain solvent used to dissolve test antibiotics but which contain no antibiotic. Cell growth is monitored continuously by incubation at 37°C in a microtiter plate reader monitoring the OD600 of the wells over an 18hour period. The percent inhibition of growth produced by each concentration of antibiotic is calculated by comparing the rates of logarithmic growth against that exhibited by cells growing in media without antibiotic. A plot of percent inhibition against log(antibiotic concentration) allows extrapolation of an  $IC_{50}$  value for each antibiotic.

## F. Determining the Specificity of the Test Antibiotics

A comparison of the IC<sub>50</sub>s generated by antibiotics of known mechanism of action under antisense induced and non-induced conditions allows the pathway in which a proliferation-required nucleic acid lies to be identified. If cells expressing an antisense nucleic acid against a proliferation-required gene are selectively sensitive to an antibiotic acting via a particular pathway, then the gene against which the antisense acts is involved in the pathway in which the antibiotic acts.

## G. Identification of Pathway in which a Test Antibiotic Acts

As discussed above, the cell based assay may also be used to determine the pathway against which a test antibiotic acts. In such an analysis, the pathways against which each member of a panel of antisense nucleic acids acts are identified as described above. A panel of cells, each containing an inducible antisense vector against a gene in a known proliferation-required pathway, is contacted with a test antibiotic for which it is desired to determine the pathway

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on which it acts under inducing an non-inducing conditions. If heightened sensitivity is observed in induced cells expressing antisense against a gene in a particular pathway but not in induced cells expressing antisense against genes in other pathways, then the test antibiotic acts against the pathway for which heightened sensitivity was observed.

One skilled in the art will appreciate that further optimization of the assay conditions, such as the concentration of inducer used to induce antisense expression and/or the growth conditions used for the assay (for example incubation temperature and media components) may further increase the selectivity and/or magnitude of the antibiotic sensitization exhibited.

The following example confirms the effectiveness of the methods described above.

### **EXAMPLE 11**

# Identification of the Pathway in which a Proliferation-Required Gene Lies

Antibiotics of various chemical classes and modes of action were purchased from Sigma Chemicals (St. Louis, MO). Stock solutions were prepared by dissolving each antibiotic in an appropriate aqueous solution based on information provided by the manufacturer. The final working solution of each antibiotic contained no more than 0.2% (w/v) of any organic solvent. To determine their potency against a bacterial strain engineered for expression of an antisense against a proliferation-required 50S ribosomal protein, each antibiotic was serially diluted two or three fold in growth medium supplemented with the appropriate antibiotic for maintenance of the anti-sense construct. At least ten dilutions were prepared for each antibiotic. 25 µL aliquots of each dilution were transferred to discrete wells of a 384-well microplate (the assay plate) using a multi-channel pipette. Quadruplicate wells were used for each dilution of an antibiotic under each treatment condition (plus and minus inducer). Each assay plate contained twenty wells for cell growth controls (growth media replacing antibiotic), ten wells for each treatment (plus and minus inducer, in this example IPTG). Assay plates were usually divided into the two treatments: half the plate containing induced cells and an appropriate concentrations of inducer (in this example IPTG) to maintain the state of induction, the other half containing non-induced cells in the absence of IPTG.

Cells for the assay were prepared as follows. Bacterial cells containing a construct, from which expression of antisense nucleic acid against rplL and rplJ, which encode proliferation-required 50S ribosomal subunit proteins, is inducible in the presence of IPTG, were grown into exponential growth ( $OD_{600}$  0.2 to 0.3) and then diluted 1:100 into fresh media containing either 400  $\mu$ M or 0  $\mu$ M inducer (IPTG). These cultures were incubated at 37° C for 2.5 hr. After a 2.5 hr incubation, induced and non-induced cells were respectively diluted into an assay medium at a final  $OD_{600}$  value of 0.0004. The medium contained an appropriate concentration of the antibiotic for the maintenance of the anti-sense construct. In addition, the medium used to dilute induced cells was supplemented with 800  $\mu$ M IPTG so that addition to the assay plate would result in a final IPTG concentration of 400  $\mu$ M. Induced and non-induced cell suspensions were dispensed (25  $\mu$ I/well) into the appropriate wells of the assay plate as discussed previously. The plate was then loaded into a plate reader, incubated at constant temperature, and cell growth was monitored in each well by the measurement of

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light scattering at 595 nm. Growth was monitored every 5 minutes until the cell culture attained a stationary growth phase. For each concentration of antibiotic, a percentage inhibition of growth was calculated at the time point corresponding to mid-exponential growth for the associated control wells (no antibiotic, plus or minus IPTG). For each antibiotic and condition (plus or minus IPTG), a plot of percent inhibition versus log of antibiotic concentration was generated and the IC50 determined. A comparison of the IC50 for each antibiotic in the presence and absence of IPTG revealed whether induction of the antisense construct sensitized the cell to the mechanism of action exhibited by the antibiotic. Cells which exhibited a significant (standard statistical analysis) numerical decrease in the IC50 value in the presence of inducer were considered to have an increased sensitivity to the test antibiotic.

The results are provided in the table below, which lists the classes and names of the antibiotics used in the analysis, the targets of the antibiotics, the IC50 in the absence of IPTG, the IC50 in the presence of IPTG, the concentration units for the IC50s, the fold increase in IC50 in the presence of IPTG, and whether increased sensitivity was observed in the presence of IPTG.

TABLE IV Effect of Expression of Antisense RNA to rplL and rplJ on Antibiatic Sensitivity

AN HBIUTIC CLASS (Names	TARGET	IC50 (-IPTG)	IC50 (+ IPTG)	Conc.	Fold Increase	Sensitivity
				Unit	in Sensitivity	Increased?
PHULEIN SYNTHESIS INHIBITOR ANTIBIOTICS						
AMINOGLYCOSIDES			-			
Gentamicin	30S ribosome function	2715	19.19	lanjuu	1/1	, 5 ,
Streptomycin	30S ribosome function	11280	181	i i	02	S ;
Spectinomycin	30S ribosome function	18050	15.	ing/mi	₹	16S
Tobramycin	30S ribosome function	3594	20 70 58	illiligin Gran	ī	res
MACROLIDES	S		00.07		<u>.</u>	Yes
Erythromycin	50S ribosome function	7467	187	lanton	9	
AROMATIC POYKETIDES			2		<b>5</b>	res
Tetracycline	30S ribosome function	199.7	1.83	lu/ou	100	Vor
Minocycline	30S ribosome function	668.4	3.897	lm/uu	173	ς σ - Α
Doxycycline	30S ribosome function	413.1	27.81	Julian	10	G ;
OTHER PROTEIN SYNTHESIS INHIBITORS			2		2	Sal
Fusidic acid	Elongation Factor G function	59990	641	lmfuu	70	>
Chloramphenicol	30S ribosome function	465.4	1.516	lm/nu	307	S .
Lincomycin	50S ribosome function	47150	324.2	julia.	145	G 7
OTHER ANTIBIOTIC MECHANISMS			1:1:0	5	<u> </u>	Tes
B-LACTAMS				<u>·</u>		
Cefoxitin	Cell wall biosynthesis	2782	2484	m/mi		2
Cefotaxime	Cell wall biosynthesis	24.3	24 16	m/ou		
DNA SYNTHESIS INHIBITORS		*	2	5		2 .
Nalidixic acid	DNA Gyrase activity	6973	6025	mjuu	-	
Offoxacin	DNA Gyrase activity	49.61	45.89	in the second		2 .
ОТНЕЯ		5	2		_	2
Bacitracin	Cell membrane function	4077	4677		•	
Trimethoprim	Dihydrofolate Reductase activity	128.9		Jul/uu	- +	2 2
Vancomycin	Cell wall biosynthesis	145400	72550	lm/R:	- ~	2 2
		T			7	. 021

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The above results demonstrate that induction of an antisense RNA to genes encoding 50S ribosomal subunit proteins results in a selective and highly significant sensitization of cells to antibiotics that inhibit ribosomal function and protein synthesis. The above results further demonstrate that induction of an antisense construct to an essential gene sensitizes an organism to compounds that interfere with that gene products' biological role. This sensitization is restricted to compounds that interfere with pathways associated with the targeted gene and it's product.

Assays utilizing antisense constructs to essential genes can be used to identify compounds that specifically interfere with the activity of multiple targets in a pathway. Such constructs can be used to simultaneously screen a sample against multiple targets in one pathway in one reaction (Combinatorial HTS).

Furthermore, as discussed above, panels of antisense construct containing cells may be used to characterize the point of intervention of any compound affecting an essential biological pathway including antibiotics with no known mechanism of action.

Another embodiment of the present invention is a method for determining the pathway against which a test antibiotic compound is active in which the activity of target proteins or nucleic acids involved in proliferation-required pathways is reduced by contacting cells with a sublethal concentration of a known antibiotic which acts against the target protein or nucleic acid. In one embodiment, the target protein or nucleic acid is a target protein or nucleic acid corresponding to a proliferation-required nucleic acid identified using the methods described above. The method is similar to those described above for determining which pathway a test antibiotic acts against except that rather than reducing the activity or level of a proliferation-required gene product using a sublethal level of antisense to a proliferation-required nucleic acid, the activity or level of the proliferation-required gene product is reduced using sublethal level of a known antibiotic which acts against the proliferation required gene product.

Interactions between drugs which affect the same biological pathway has been described in the literature. For example, Mecillinam (Amdinocillin) binds to and inactivates the penicillin binding protein 2 (PBP2, product of the *mrdA* in *E. coli*). This antibiotic inteacts with other antibiotics that inhibit PBP2 as well as antibiotics that inhibit other penicillin binding proteins such as PBP3 [(Gutmann, L., Vincent, S., Billot-Klein, D., Acar, J.F., Mrena, E., and Williamson, R. (1986) Involvement of penicillin-binding protein 2 with other penicillin-binding proteins in lysis of *Escherichia coli* by some beta-lactam antibiotics alone and in synergistic lytic effect of amdinocillin (mecillinam). Antimicrobial Agents & Chemotherapy, 30:906-912), the disclosure of which is incorporated herein by reference in its entirety]. Interactions between drugs could, therefore, involve two drugs that inhibit the same target protein or nucleic acid or inhibit different proteins or nucleic acids in the same pathway [(Fukuoka, T., Domon, H., Kakuta, M., Ishii, C., Hirasawa, A., Utsui, Y., Ohya, S., and Yasuda, H. (1997) Combination effect between panipenem and vancomycin on highly methicillin-resistant Staphylococcus aureus. Japan. J. Antibio. 50:411-419; Smith, C.E., Foleno, B.E., Barrett, J.F., and Frosc, M.B. (1997) Assessment of the synergistic interactions of levofloxacin and ampicillin against Enterococcus faecium by the checkerboard agar dilution and time-kill methods. Diagnos. Microbiol. Infect. Disease 27:85-92; den Hollander, J.G., Horrevorts, A.M., van Goor, M.L.,

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Verbrugh, H.A., and Mouton, J.W. (1997) Synergism between tobramycin and ceftazidime against a resistant Pseudomonas aeruginosa strain, tested in an in vitro pharmacokinetic model. Antimicrobial Agents & Chemotherapy. 41:95-110), the disclosure of all of which are incorporated herein by reference in their entireties).

Two drugs may interact even though they inhibit different targets. For example, the proton pump inhibitor, Omeprazole, and the antibiotic, Amoxycillin, two synergistic compounds acting together, can cure *Helicobacter pylori* infection [( Gabryelewicz, A., Laszewicz, W., Dzieniszewski, J., Ciok, J., Marlicz, K., Bielecki, D., Popiela, T., Legutko, J., Knapik, Z., Poniewierka, E. (1997) Multicenter evaluation of dual-therapy (omeprazol and amoxycillin) for *Helicobacter pylori*-associated duodenal and gastric ulcer (two years of the observation). J. Physiol. Pharmacol. 48 Suppl 4:93-105), the disclosure of which is incorporated herein by reference in its entirety).

The growth inhibition from the sublethal concentration of the known antibiotic may be at least about 5%, at least about 8%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, or at least about 75%, or more.

Alternatively, the sublethal concentration of the known antibiotic may be determined by measuring the activity of the target proliferation-required gene product rather than by measuring growth inhibition.

Cells are contacted with a combination of each member of a panel of known antibiotics at a sublethal level and varying concentrations of the test antibiotic. As a control, the cells are contacted with varying concentrations of the test antibiotic alone. The  $IC_{50}$  of the test antibiotic in the presence and absence of the known antibiotic is determined. If the  $IC_{50}$ s in the presence and absence of the known drug are substantially similar, then the test drug and the known drug act on different pathways. If the  $IC_{50}$ s are substantially different, then the test drug and the known drug act on the same pathway.

Another embodiment of the present invention is a method for identifying a candidate compound for use as an antibiotic in which the activity of target proteins or nucleic acids involved in proliferation-required pathways is reduced by contacting cells with a sublethal concentration of a known antibiotic which acts against the target protein or nucleic acid. In one embodiment, the target protein or nucleic acid is a target protein or nucleic acid corresponding to a proliferation-required nucleic acid identified using the methods described above. The method is similar to those described above for identifying candidate compounds for use as antibiotics except that rather than reducing the activity or level of a proliferation-required gene product using a sublethal level of antisense to a proliferation-required nucleic acid, the activity or level of the proliferation-required gene product is reduced using a sublethal level of a known antibiotic which acts against the proliferation required gene product.

The growth inhibition from the sublethal concentration of the known antibiotic may be at least about 5%, at least about 8%, at least about 10%, at least about 20%, at least about 30%, at least about 40%, at least about 50%, at least about 60%, or at least about 75%, or more.

Alternatively, the sublethal concentration of the known antibiotic may be determined by measuring the activity of the target proliferation-required gene product rather than by measuring growth inhibition.

In order to characterize test compounds of interest, cells are contacted with a panel of known antibiotics at a sublethal level and one or more concentrations of the test compound. As a control, the cells are contacted with the same concentrations of the test compound alone. The  $IC_{50}$  of the test compound in the presence and absence of the known antibiotic is determined. If the  $IC_{50}$  of the test compound is substantially different in the presence and absence of the known drug then the test compound is a good candidate for use as an antibiotic. As discussed above, once a candidate compound is identified using the above methods its structure may be optimized using standard techniques such as combinatorial chemistry.

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Representative known antibiotics which may be used in each of the above methods are provided in the table below. However, it will be appreciated that other antibiotics may also be used.

ANTIBIOTIC	INHIBITS/TARGET	RESISTANT MUTANTS
Inhibitors of Transcription		
Rifamycin, 1959 Rifampicin Rifabutin Rifaximin	Inhibits initiation of transcription/ß-subunit RNA polymerase, <i>rpoB</i>	rpoB, crp, cyaA
Streptolydigin	Accelerates transcription chain termination/ß- subunit RNA polymerase	гроВ
Streptovaricin Actinomycin D+EDTA	an acyclic ansamycin, inhibits RNA polymerase Intercalates between 2 successive G-C pairs, rpoB, inhibits RNA synthesis	rpoB pldA
Inhibitors of Nucleic Acid Metab Quinolones, 1962 Nalidixic acid Oxolinic acid		gyrAorB, icd, sloB
Fluoroquinolones Ciprofloxacin, 1983 Norfloxacin	subunit gyrase, <i>gyrA</i> and/or topoisomerase IV_ (probable target in Staph)	gyrA norA (efflux in Staph) hipΩ
Coumerins Novobiocin	Inhibits ATPase activity of ß-subunit gyrase, gyrB	gyrB, cysB, cysE, nov,
Coumermycin	Inhibits ATPase activity of ß-subunit gyrase,	ompA gyrB, hisW
Albicidin Metronidazole	DNA synthesis Causes single-strand breaks in DNA	tsx (nucleoside channel) nar
Inhibitors of Metabolic Pathways Sulfonamides, 1932 Sulfanilamide	blocks synthesis of dihydrofolate, dihydro-	folP, gpt, pabA, pabB,
Trimethoprim, 1962 Showdomycin	pteroate synthesis, <i>folP</i> Inhibits dihydrofolate reductase, <i>folA</i> Nucleoside analogue capable of alkylating	pabC folA, thyA nupC, pnp
		··

ANTIBIOTIC	INHIBITS/TARGET	RESISTANT MUTANTS
	sulfhydryl groups, inhibitor of thymidylate	
<del></del>	synthetase	
Thiolactomycin	type II fatty acid synthase inhibitor	emrB
		fadB, emrB due to gene dosage
Psicofuranine	Adenosine glycoside antibiotic, target is GMP	guaA,B
	synthetase	92277,5
Triclosan	Inhibits fatty acid synthesis	fabl (envM)
Diazoborines Isoniazid,	heterocyclic, contains boron, inhibit fatty acid	fabl (envM)
Ethionamide	synthesis, enoyl-ACP reductase, fabl	,
Inhibitors of Translation		· .
Phenylpropanoids	Binds to ribosomal peptidyl transfer center	•
Chloramphenicol, 1947	preventing peptide translocation/ binds to S6,	rrn cml/ mos/ cm-F
•	L3, L6, L14, L16, L25, L26, L27, but	rrn, cmlA, marA, ompF, ompR
	preferentially to L16	σπρτι
Tetracyclines, 1948, type II	Binding to 30S ribosomal subunit, "A" site	clmA (cmr), mar, ompF
polyketides	on 30S subunit, blocks peptide elongation,	ommi (ami), mar, ampr
Minocycline	strongest binding to S7	
Doxycycline		
Macrolides (type I polyketides)	Binding to 50 S ribosomal subunit, 23S rRNA,	
Erythromycin, 1950	blocks peptide translocation, L15, L4, L12	
Carbomycin, Spiramycin		rrn, rpIC, rpID, rpIV , mac
etc		·
Aminoglycosides Streptomycin,	Irreversible binding to 30S ribosomal subunit,	· · · · · · · · · · · · · · · · · · ·
1944	prevents translation or causes mistranslation of	rpsL, strC,M, ubiF
Neomycin	mRNA/16S rRNA	atpA-E, ecfB,
	,	hemAC,D,E,G, topA,
Spanting-weig Kananania		rpsC,D,E, rrn, spcB
Spectinomycin Kanamycin		atpA-atpE, cpxA, ecfB,
·		hemA,B,L, topA
Kasugamycin	·	ksgA,B,C,D, rplB,K,
		rpsl,N,M,R
Gentamicin, 1963		rplF, ubiF
Amikacin		срхА
Paromycin		rpsL
Lincosamides	Binding to 50 S ribosomal subunit, blocks	
Lincomycin, 1955 Clindamycin	peptide translocation	linB, rpIN,O, rpsG
Streptogramins Virginiamycin, 1955 Pristinamycin	2 components, Streptogramins A&B, bind to	
Synercid: quinupristin	the 50S ribosomal subunit blocking peptide	
/dalfopristin	translocation and peptide bond formation	
Fusidanes	Inhibition of elongation factor G (EF-G) prevents	fusA
Fusidic Acid	peptide translocation	
Kirromycin (Mocimycin)	Inhibition of elongation factor TU (EF-Tu), prevents peptide bond formation	tufA,B

ANTIBIOTIC	INHIBITS/TARGET	RESISTANT MUTANTS
Pulvomycin .	Binds to and inhibits EF-TU	
Thiopeptin	Sulfur-containing antibiotic, inhibits protein synthesis, EF-G	rplE
Tiamulin	Inhibits protein synthesis	rpIC, rpID
Negamycin	Inhibits termination process of protein synthesis	prfB
Oxazolidinones Linezolid Isoniazid	23S rRNA	
·	• •	pdx
Nitrofurantoin	Inhibits protein synthesis, nitroreductases convert nitrofurantoin to highly reactive electrophilic intermediates which attack bacterial ribosomal proteins non-specifically	nfnA,B
Pseudomonic Acids Mupirocin (Bactroban)	Inhibition of isoleucyl tRNA synthetase-used for Staph, topical cream, nasal spray	ileS
Indolmycin	Inhibits tryptophanyl-tRNA synthetase	trpS
Viomycin		rrmA (23S rRNA methyltransferase; mutant has slow growth rate, slow chair elongation rate, and viomycin resistance)
Thiopeptides	Binds to L11-23S RNA complex	traini form tobiotalice;
Thiostrepton	Inhibits GTP hydrolysis by EF-G	
Micrococcin	Stimulates GTP hydrolysis by EF-G	

# Inhibitors of Cell Walls/Membranes

B-lactams Penicillin, 1929 Ampicillin	Inhibition of one or more cell wall transpeptidases, endopeptidases, and	
Methicillin, 1960	glycosidases (PBPs), of the 12 PBPs only 2 are essential: mrdA (PBP2) and ftsl (pbpB, PBP3)	ampC, ampD, ampE, envZ, galU, hipA, hipQ, ompC, ompF, ompR, ptsl, rfa, tolD, tolE
Cephalosporins, 1962		tonB
Mecillinam (amdinocillin)	Binds to and inactivates PBP2 (mrdA) Inactivates PBP3 (fts/)	alaS, argS, crp, cyaA, envB, mrdA,B,
Aztreonam (Furazlocillin)		mreB,C,D
Bacilysin, Tetaine	Dipeptide, inhib glucosamine synthase	dρpA
Glycopeptides Vancomycin, 1955	Inhib G+ cell wall syn, binds to terminal D- ala-D-ala of pentapeptide,	,,
Polypeptides Bacitracin	Prevents dephosphorylation and regeneration of lipid carrier	rfa
Cyclic lipopeptide Daptomycin, 1980	Disrupts multiple aspects of membrane	

function, including peptidoglycan synthesis, lipoteichoic acid synthesis, and the bacterial

membrane potential

Cyclic polypeptides Polymixin, 1939

Surfactant action disrupts cell membrane lipids, binds lipid A minety of LPS

pmrA

Fosfomycin, 1969

Analogue of P-enolpyruvate, inhibits 1st step in peptidoglycan synthesis - UDP-N-

murA, crp, cyaA glpT, hipA, ptsl, uhpT

acetylglucosamine enolpyruvyl transferase.

murA. Also acts as Immunosuppressant

Prevents formation of D-ala dimer, inhibits Dala ligase, ddlA,B

hipA, cycA

Alafosfalin

Cycloserine

phosphonodipeptide, cell wall synthesis inhibitor, potentiator of -lactams

pepA, tpp

Inhibitors of Protein Processing/Transport

Globomycin

Inhibits signal peptidase II (cleaves prolipoproteins subsequent to lipid modification, IspA

lpp, dnaE

## **EXAMPLE 12**

# Transfer of Exogenous Nucleic Acid Sequences to other Bacterial Species Using the E. coli Expression Vectors or Expression Vectors Functional in Bacterial Species other than E. coli.

The above methods were validated using antisense nucleic acids which inhibit the growth of E. coli which were identified using methods similar to those described above. Expression vectors which inhibited growth of E. coli upon induction of antisense RNA expression with IPTG were transformed directly into Enterobacter cloacae, Klebsiella pneumonia or Salmonella typhimurium. The transformed cells were then assayed for growth inhibition according to the method of Example 1. After growth in liquid culture, cells were plated at various serial dilutions and a score determined by calculating the log difference in growth for INDUCED vs. UNINDUCED antisense RNA expression as determined by the maximum 10 fold dilution at which a colony was observed. The results of these experiments are listed below in Table VI. If there was no effect of antisense RNA expression in an organism, the clone is minus in Table VI. In contrast, a positive in Table VI means that at least 10 fold more cells were required to observe a colony on the induced plate than on the noninduced plate under the conditions used and in that organism.

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Sixteen of the construts were found to inhibit growth in all the organisms tested upon induction of antisense RNA expression with IPTG. Those skilled in the art will appreciate that a negative result in a heterologous organism does not mean that that organism is missing that gene nor does it mean that the gene is unessential. However, a positive result means that the heterologous organism contains a homologous gene which is required for proliferation of that organism. The homologous gene may be obtained using the methods described herein. Those cells that are inhibited by antisense may be used in cell based assays as described herein for the identification and characterization of compounds in order to

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develop antibiotics effective in these organisms. Those skilled in the art will appreciate that an antisense molecule which works in the organism from which it was obtained will not always work in a heterologous organism.

TABLE VI
Sensitivity of Other Microorganisms to Antisense Nucleic Acids That Inhibit Proliferation in *E. coli* 

Mol. No.	S. typhimurium	E. cloacae	K. pneumoniae
EcXA001	+	+	•
EcXA004			. •
EcXA005	+	+	+
EcXA006			•
EcXA007		+	
EcXA008	+		+
EcXA010	+	. +	+
EcXA011		+	
EcXA012		+	
EcXA013	+	+	+
EcXA014	+	+	
EcXA015	:. •	+	+
EcXAD16	+	+	+
EcXA017	+	+	+
EcXA018	+	+	+
EcXA019	+	+	+
- EcXA020	+	+	+
EcXA021	+	+	+
EcXA023	+	+	+
EcXA024	+		+
EcXA025	-		
EcXA026	+	+	
EcXA027	+	+	+
EcXA028	+		т - ·
EcXA029			•

			·
Mol. No.	S. typhimurium	E. cloacae	K. pneumoniae
EcXA030	+	+	+
EcXA031	. +		
EcXA032	+		
EcXA033	+	+	+
EcXA034	. +	+	+
EcXA035			
EcXA036	+		+
EcXA037		+	
EcXA038	+	+	
· EcXA039	+	-	
EcXA041	+	+	+
EcXA042	. •	+	+
EcXA044			
EcXA045		+	
EcXA046	•		
EcXA047	+	+	
EcXA048	·		
EcXA049	+		
EcXA050			
EcXA051	+		
EcXA052	+ .		
EcXA053	+	+	+
EcXA054			+
EcXA055	+		<u>:</u>

## **EXAMPLE 13**

# Use of Identified Exogenous Nucleic Acid Sequences as Probes

The identified sequence of the present invention can be used as probes to obtain the sequence of additional genes of interest from a second organism. For example, probes to potential bacterial target proteins may be hybridized to nucleic acids from other organisms including other bacteria and higher organisms, to identify homologous sequences. Such

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hybridization might indicate that the protein encoded by the gene to which the probe corresponds is found in humans and therefore not necessarily a good drug target. Alternatively, the gene can be conserved only in bacteria and therefore would be a good drug target for a broad spectrum antibiotic or antimicrobial.

Probes derived from the identified nucleic acid sequences of interest or portions thereof can be labeled with detectable labels familiar to those skilled in the art, including radioisotopes and non-radioactive labels, to provide a detectable probe. The detectable probe can be single stranded or double stranded and can be made using techniques known in the art, including *in vitro* transcription, nick translation, or kinase reactions. A nucleic acid sample containing a sequence capable of hybridizing to the labeled probe is contacted with the labeled probe. If the nucleic acid in the sample is double stranded, it can be denatured prior to contacting the probe. In some applications, the nucleic acid sample can be immobilized on a surface such as a nitrocellulose or nylon membrane. The nucleic acid sample can comprise nucleic acids obtained from a variety of sources, including genomic DNA, cDNA libraries, RNA, or tissue samples.

Procedures used to detect the presence of nucleic acids capable of hybridizing to the detectable probe include well known techniques such as Southern blotting, Northern blotting, dot blotting, colony hybridization, and plaque hybridization. In some applications, the nucleic acid capable of hybridizing to the labeled probe can be cloned into vectors such as expression vectors, sequencing vectors, or in vitro transcription vectors to facilitate the characterization and expression of the hybridizing nucleic acids in the sample. For example, such techniques can be used to isolate, purify and clone sequences from a genomic library, made from a variety of bacterial species, which are capable of hybridizing to probes made from the sequences identified in Examples 5 and 6.

### **EXAMPLE 14**

## Preparation of PCR Primers and Amplification of DNA

The identified E. coli genes corresponding directly to or located within the operon of nucleic acid sequences required for proliferation or portions thereof can be used to prepare PCR primers for a variety of applications, including the identification or isolation of homologous sequences from other species, for example *S. typhimurium, E. cloacae, and Klebsiella pneumoniae*, which contain part or all of the homologous genes. Because homologous genes are related but not identical in sequence, those skilled in the art will often employ degenerate sequence PCR primers. Such degenerate sequence primers are designed based on conserved sequence regions, either known or suspected, such as conserved coding regions. The successful production of a PCR product using degenerate probes generated from the sequences identified herein would indicate the presence of a homologous gene sequence in the species being screened. The PCR primers are at least 10 bases, and preferably at least 20 bases in length. More preferably, the PCR primers are at least 20-30 bases in length. In some embodiments, the PCR primers can be more than 30 bases in length. It is preferred that the primer pairs have approximately the same G/C ratio, so that melting temperatures are approximately the same. A variety of PCR techniques are familiar to those skilled in the art. For a review of PCR technology, see Molecular Cloning to Genetic Engineering White, B.A. Ed. in Methods in Molecular Biology 67: Humana Press, Totowa 1997. When the entire coding sequence of the target gene is known, the 5' and 3' regions of the target gene

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can be used as the sequence source for PCR probe generation. In each of these PCR procedures, PCR primers on either side of the nucleic acid sequences to be amplified are added to a suitably prepared nucleic acid sample along with dNTPs and a thermostable polymerase such as Taq polymerase, Pfu polymerase, or Vent polymerase. The nucleic acid in the sample is denatured and the PCR primers are specifically hybridized to complementary nucleic acid sequences in the sample. The hybridized primers are extended. Thereafter, another cycle of denaturation, hybridization, and extension is initiated. The cycles

## **EXAMPLE 15**

are repeated multiple times to produce an amplified fragment containing the nucleic acid sequence between the primer sites.

## **Inverse PCR**

The technique of inverse polymerase chain reaction can be used to extend the known nucleic acid sequence identified in Examples 5 and 6. The inverse PCR reaction is described generally by Ochman et al., in Ch. 10 of PCR Technology: Principles and Applications for DNA Amplification, (Henry A. Erlich, Ed.) W.H. Freeman and Co. (1992). Traditional PCR requires two primers that are used to prime the synthesis of complementary strands of DNA. In inverse PCR, only a core sequence need be known.

Using the sequences identified as relevant from the techniques taught in Examples 5 and 6 and applied to other species of bacteria, a subset of exogenous nucleic sequences are identified that correspond to genes or operons that are required for bacterial proliferation. In species for which a genome sequence is not known, the technique of inverse PCR provides a method for obtaining the gene in order to determine the sequence or to place the probe sequences in full context to the target sequence to which the identified exogenous nucleic acid sequence binds.

To practice this technique, the genome of the target organism is digested with an appropriate restriction enzyme so as to create fragments of nucleic acid that contain the identified sequence as well as unknown sequences that flank the identified sequence. These fragments are then circularized and become the template for the PCR reaction. PCR primers are designed in accordance with the teachings of Example 15 and directed to the ends of the identified sequence are synthesized. The primers direct nucleic acid synthesis away from the known sequence and toward the unknown sequence contained within the circularized template. After the PCR reaction is complete, the resulting PCR products can be sequenced so as to extend the sequence of the identified gene past the core sequence of the identified exogenous nucleic acid sequence identified. In this manner, the full sequence of each novel gene can be identified. Additionally the sequences of adjacent coding and noncoding regions can be identified.

### **EXAMPLE 16**

## Identification of Genes Required for Staphylococcus aureus Proliferation

Genes required for proliferation in Staphylococcus aureus are identified according to the methods described above.

#### **EXAMPLE 17**

## Identification of Genes Required for Neisseria gonorrhoeae Proliferation

Genes required for proliferation in Neisseria gonorrhoeae are identified according to the methods described above.

#### **EXAMPLE 18**

## Identification of Genes Required for Pseudomonas aeruginosa Proliferation

Genes required for proliferation in *Pseudomonas aeruginosa* are identified according to the methods described above.

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### **EXAMPLE 19**

# Identification of Genes Required for Enterococcus faecalis Proliferation

Genes required for proliferation in Enterococcus faecalis are identified according to the methods described above.

### **EXAMPLE 20**

## Identification of Genes Required for Haemophilus influenzae Proliferation

Genes required for proliferation in Haemophilus influenzae are identified according to the methods described above.

## **EXAMPLE 21**

# Identification of Genes Required for Salmonella typhimurium Proliferation

Genes required for proliferation in Salmonella typhimurium are identified according to the methods described above.

#### **EXAMPLE 22**

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# Identification of Genes Required for Helicobacter pylori Proliferation

Genes required for proliferation in Helicobacter pylori are identified according to the methods described above.

#### **EXAMPLE 23**

## Identification of Genes Required for Mycoplasma pneumoniae Proliferation

Genes required for proliferation in Mycoplasma pneumoniae are identified according to the methods described

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## **EXAMPLE 24**

## Identification of Genes Required for Plasmodium ovale Proliferation

Genes required for proliferation in Plasmodium ovale are identified according to the methods described above.

## EXAMPLE 25

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# Identification of Genes Required for Saccharomyces cerevisiae Proliferation

Genes required for proliferation in Saccharomyces cerevisiae are identified according to the methods described

above.

#### **EXAMPLE 26**

# Identification of Genes Required for Entamoeba histolytica Proliferation

Genes required for proliferation in Entamoeba histolytica are identified according to the methods described above.

#### **EXAMPLE 27**

## Identification of Genes Required for Candida albicans Proliferation

Genes required for proliferation in Candida albicans are identified according to the methods described above.

#### **EXAMPLE 28**

# Identification of Genes Required for Klebsiella pneumoniae Proliferation

Genes required for proliferation in Klebsiella pneumoniae are identified according to the methods described above.

### **EXAMPLE 29**

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# Identification of Genes Required for Salmonella typhi Proliferation

Genes required for proliferation in Salmonella typhi are identified according to the methods described above.

#### **EXAMPLE 30**

## Identification of Genes Required for Salmonella paratyphi Proliferation

Genes required for proliferation in Salmonella paratyphi are identified according to the methods described above.

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### **EXAMPLE 31**

## Identification of Genes Required for Salmonella cholerasuis Proliferation

Genes required for proliferation in Salmonella cholerasuis are identified according to the methods described above.

### **EXAMPLE 32**

# Identification of Genes Required for Staphylococcus epidermis Proliferation

15 above.

Genes required for proliferation in Staphylococcus epidermis are identified according to the methods described

### **EXAMPLE 33**

## Identification of Genes Required for Mycobacterium tuberculosis Proliferation

Genes required for proliferation in Mycobacterium tuberculosis are identified according to the methods described

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#### **EXAMPLE 34**

# Identification of Genes Required for Mycobacterium leprae Proliferation

Genes required for proliferation in Mycobacterium leprae are identified according to the methods described above.

#### **EXAMPLE 35**

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## Identification of Genes Required for Treponema pallidum Proliferation

Genes required for proliferation in Treponema pallidum are identified according to the methods described above.

#### **EXAMPLE 36**

## Identification of Genes Required for Bacillus anthracis Proliferation

Genes required for proliferation in Bacillus anthracis are identified according to the methods described above.

### **EXAMPLE 37**

## Identification of Genes Required for Yersinia pestis Proliferation

Genes required for proliferation in Yersinia pestis are identified according to the methods described above.

### **EXAMPLE 38**

## Identification of Genes Required for Clostridium botulinum Proliferation

Genes required for proliferation in Clostridium botulinum are identified according to the methods described above.

### **EXAMPLE 39**

## Identification of Genes Required for Campylobacter jejuni Proliferation

Genes required for proliferation in Campylobacter jejuni are identified according to the methods described above.

#### **EXAMPLE 40**

## Identification of Genes Required for Chlamydia trachomatis Proliferation

Genes required for proliferation in *Chlamydia trachomatis* are identified according to the methods described above.

Use of Isolated Exogenous Nucleic Acid Fragments as Antisense Antibiotics

In addition to using the identified sequences to enable screening of molecule libraries to identify compounds useful to identify antibiotics, the sequences themselves can be used as therapeutic agents. Specifically, the identified exogenous sequences in an antisense orientation can be provided to an individual to inhibit the translation of a bacterial target gene.

### Generation of Antisense Therapeutics from Identified Exogenous Sequences

The sequences of the present invention can be used as antisense therapeutics for the treatment of bacterial infections or simply for inhibition of bacterial growth *in vitro* or *in vivo*. The therapy exploits the biological process in cells where genes are transcribed into messenger RNA (mRNA) that is then translated into proteins. Antisense RNA technology contemplates the use of antisense oligonucleotides directed against a target gene that will bind to its target and decrease or inhibit the translation of the target mRNA. In one embodiment, antisense oligonucleotides can be used to treat and control a bacterial infection of a cell culture containing a population of desired cells contaminated with bacteria. In another embodiment, the antisense oligonucleotides can be used to treat an organism with a bacterial infection.

Antisense oligonucleotides can be synthesized from any of the sequences of the present invention using methods well known in the art. In a preferred embodiment, antisense oligonucleotides are synthesized using artificial means. Uhlmann & Peymann, Chemical Rev. 90:543-584 (1990) review antisense oligonucleotide technology in detail. Modified or unmodified antisense oligonucleotides can be used as therapeutic agents. Modified antisense oligonucleotides are preferred since it is well known that antisense oligonucleotides are extremely unstable. Modification of the phosphate backbones of the antisense oligonucleotides can be achieved by substituting the internucleotide phosphate residues with methylphosphonates, phosphorothioates, phosphoramidates, and phosphate esters. Nonphosphate internucleotide analogs such as siloxane bridges, carbonate bridges, thioester bridges, as well as many others known in the art. The preparation of certain antisense oligonucleotides with modified internucleotide linkages is described in U.S. Patent No. 5,142,047, hereby incorporated by reference.

Modifications to the nucleoside units of the antisense oligonucleotides are also contemplated. These modifications can increase the half-life and increase cellular rates of uptake for the oligonucleotides in vivo. For example,

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 $\alpha$ -anomeric nucleotide units and modified bases such as 1,2-dideoxy-d-ribofuranose, 1,2-dideoxy-1-phenylribofuranose, and  $N^4$ ,  $N^4$ -ethano-5-methyl-cytosine are contemplated for use in the present invention.

An additional form of modified antisense molecules is found in peptide nucleic acids. Peptide nucleic acids (PNA) have been developed to hybridize to single and double stranded nucleic acids. PNA are nucleic acid analogs in which the entire deoxyribose phosphate backbone has been exchanged with a chemically completely different, but structurally homologous, polyamide (peptide) backbone containing 2-aminoethyl glycine units. Unlike DNA, which is highly negatively charged, the PNA backbone is neutral. Therefore, there is much less repulsive energy between complementary strands in a PNA-DNA hybrid than in the comparable DNA-DNA hybrid, and consequently they are much more stable. PNA can hybridize to DNA in either a Watson/Crick or Hoogsteen fashion (Demidov et al., *Proc. Natl. Acad. Sci. U.S.A.* 92:2637-2641, 1995; Egholm, *Nature* 365:566-568, 1993; Nielsen et al., *Science* 254:1497-1500, 1991; Dueholm et al., *New J. Chem.* 21:19-31, 1997).

Molecules called PNA "clamps" have been synthesized which have two identical PNA sequences joined by a flexible hairpin linker containing three 8-amino-3,6-dioxaoctanoic acid units. When a PNA clamp is mixed with a complementary homopurine or homopyrimidine DNA target sequence, a PNA-DNA-PNA triplex hybrid can form which has been shown to be extremely stable (Bentin et al., *Biochemistry* 35:8863-8869, 1996; Egholm et al., *Nucleic Acids Res.* 23:217-222, 1995; Griffith et al., *J. Am. Chem. Soc.* 117:831-832, 1995).

The sequence-specific and high affinity duplex and triplex binding of PNA have been extensively described (Nielsen et al., Science 254:1497-1500, 1991; Egholm et al., J. Am. Chem. Soc. 114:9677-9678, 1992; Egholm et al., Nature 365:566-568, 1993; Almarsson et al., Proc. Natl. Acad. Sci. U.S.A. 90:9542-9546, 1993; Demidov et al., Proc. Natl. Acad. Sci. U.S.A. 92:2637-2641, 1995). They have also been shown to be resistant to nuclease and protease digestion (Demidov et al., Biochem. Pharm. 48:1010-1313, 1994). PNA has been used to inhibit gene expression (Hanvey et al., Science 258:1481-1485,1992; Nielsen et al., Nucl. Acids. Res., 21:197-200, 1993; Nielsen et al., Gene 149:139-145, 1994; Good & Nielsen, Science, 95: 2073-2076, 1998; all of which are hereby incorporated by reference), to block restriction enzyme activity (Nielsen et al., supra., 1993), to act as an artificial transcription promoter (Mollegaard, Proc. Natl. Acad. Sci. U.S.A. 91:3892-3895, 1994) and as a pseudo restriction endonuclease (Demidov et al., Nucl. Acids. Res. 21:2103-2107, 1993). Recently, PNA has also been shown to have antiviral and antitumoral activity mediated through an antisense mechanism (Norton, Nature Biotechnol., 14:615-619, 1996; Hirschman et al., J. Investig. Med. 44:347-351, 1996). PNAs have been linked to various peptides in order to promote PNA entry into cells (Basu et al., Bioconj. Chem. 8:481-488, 1997; Pardridge et al., Proc. Natl. Acad. Sci. U.S.A. 92:5592-5596, 1995).

The antisense oligonucleotides contemplated by the present invention can be administered by direct application of oligonucleotides to a target using standard techniques well known in the art. The antisense oligonucleotides can be generated within the target using a plasmid, or a phage. Alternatively, the antisense nucleic acid may be expressed from a sequence in the chromosome of the target cell. It is further contemplated that contemplated that the antisense oligonucleotide contemplated are incorporated in a ribozyme sequence to enable the antisense to specifically bind and cleave its

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target mRNA. For technical applications of ribozyme and antisense oligonucleotides see Rossi et al., Pharmacol. Ther. 50(2):245-254, (1991), which is hereby incorporated by reference. The present invention also contemplates using a retron to introduce an antisense oligonucleotide to a cell. Retron technology is exemplified by U.S. Patent No. 5,405,775, which is hereby incorporated by reference. Antisense oligonucleotides can also be delivered using liposomes or by electroporation techniques which are well known in the art.

The antisense nucleic acids of the present invention can also be used to design antibiotic compounds comprising nucleic acids which function by intracellular triple helix formation. Triple helix oligonucleotides are used to inhibit transcription from a genome. The sequences identified as required for proliferation in the present invention, or portions thereof, can be used as templates to inhibit microorganism gene expression in individuals infected with such organisms. Traditionally, homopurine sequences were considered the most useful for triple helix strategies. However, homopyrimidine sequences can also inhibit gene expression. Such homopyrimidine oligonucleotides bind to the major groove at homopurine:homopyrimidine sequences. Thus, both types of sequences based on the sequences of the present invention that are required for proliferation are contemplated for use as antibiotic compound templates.

The antisense oligonucleotides of this example employ the identified sequences of the present invention to induce bacterial cell death or at least bacterial stasis by inhibiting target gene translation. Antisense oligonucleotides containing from about 8 to 40 bases of the sequences of the present invention have sufficient complementary to form a duplex with the target sequence under physiological conditions.

To kill bacterial cells or inhibit their growth, the antisense oligonucleotides are applied to the bacteria or to the target cells under conditions that facilitate their uptake. These conditions include sufficient incubation times of cells and oligonucleotides so that the antisense oligonucleotides are taken up by the cells. In one embodiment, an incubation period of 7-10 days is sufficient to kill bacteria in a sample. An optimum concentration of antisense oligonucleotides is selected for use.

The concentration of antisense oligonucleotides to be used can vary depending on the type of bacteria sought to be controlled, the nature of the antisense oligonucleotide to be used, and the relative toxicity of the antisense oligonucleotide to the desired cells in the treated culture. Antisense oligonucleotides can be introduced to cell samples at a number of different concentrations preferably between 1x10<sup>-10</sup>M to 1x10<sup>-4</sup>M. Once the minimum concentration that can adequately control gene expression is identified, the optimized dose is translated into a dosage suitable for use *in vivo*. For example, an inhibiting concentration in culture of 1x10<sup>-7</sup> translates into a dose of approximately 0.6 mg/kg body weight. Levels of oligonucleotide approaching 100 mg/kg body weight or higher may be possible after testing the toxicity of the oligonucleotide in laboratory animals. It is additionally contemplated that cells from the subject are removed, treated with the antisense oligonucleotide, and reintroduced into the subject. This range is merely illustrative and one of skill in the art are able to determine the optimal concentration to be used in a given case.

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After the bacterial cells have been killed or controlled in a desired culture, the desired cell population may be used for other purposes.

#### **EXAMPLE 41**

The following example demonstrates the ability of an *E. coli* antisense oligonucleotide to act as a bactericidal or bacteriostatic agent to treat a contaminated cell culture system. The application of the antisense oligonucleotides of the present invention are thought to inhibit the translation of bacterial gene products required for proliferation.

The antisense oligonucleotide of this example corresponds to a 30 base phophorothioate modified oligodeoxynucelotide complementary to a nucleic acid involved in proliferation, such as Molecule Number EcXA001. A sense oligodeoxynucelotide complementary to the antisense sequence is synthesized and used as a control. The oligonucleotides are synthesized and purified according to the procedures of Matsukura, et al., Gene 72:343 (1988). The test oligonucleotides are dissolved in a small volume of autoclaved water and added to culture medium to make a 100 micromolar stock solution.

Human bone marrow cells are obtained from the peripheral blood of two patients and cultured according standard procedures well known in the art. The culture is contaminated with the K-12 strain of *E. coli* and incubated at 37°C overnight to establish bacterial infection.

The control and antisense oligonucleotide containing solutions are added to the contaminated cultures and monitored for bacterial growth. After a 10 hour incubation of culture and oligonucleotides, samples from the control and experimental cultures are drawn and analyzed for the translation of the target bacterial gene using standard microbiological techniques well known in the art. The target *E. coli* gene is found to be translated in the control culture treated with the control oligonucleotide, however, translation of the target gene in the experimental culture treated with the antisense oligonucleotide of the present invention is not detected or reduced.

### **EXAMPLE 42**

A subject suffering from an *E. coli* infection is treated with the antisense oligonucleotide preparation of Example 39. The antisense oligonucleotide is provided in a pharmaceutically acceptable carrier at a concentration effective to inhibit the translation of the target gene. The present subject is treated with a concentration of antisense oligonucleotide sufficient to achieve a blood concentration of about 100 micromolar. The patient receives daily injections of antisense oligonucleotide to maintain this concentration for a period of 1 week. At the end of the week a blood sample is drawn and analyzed for the presence or absence using standard techniques well known in the art. There is no detectable evidence of E. coli and the treatment is terminated.

#### **EXAMPLE 43**

### Preparation and use of Triple Helix Probes

The sequences of microorganism genes required for proliferation of the present invention are scanned to identify 10mer to 20-mer homopyrimidine or homopurine stretches that could be used in triple-helix based strategies for inhibiting gene

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expression. Following identification of candidate homopyrimidine or homopurine stretches, their efficiency in inhibiting gene expression is assessed by introducing varying amounts of oligonucleotides containing the candidate sequences into a population of bacterial cells that normally express the target gene. The oligonucleotides may be prepared on an oligonucleotide synthesizer or they may be purchased commercially from a company specializing in custom oligonucleotide synthesis, such as GENSET, Paris, France.

The oligonucleotides can be introduced into the cells using a variety of methods known to those skilled in the art, including but not limited to calcium phosphate precipitation, DEAE-Dextran, electroporation, liposome-mediated transfection or native uptake.

Treated cells are monitored for a reduction in proliferation using techniques such as monitoring growth levels as compared to untreated cells using optical density measurements. The oligonucleotides that are effective in inhibiting gene expression in cultured cells can then be introduced *in vivo* using the techniques well known in that art at a dosage level shown to be effective.

In some embodiments, the natural (beta) anomers of the oligonucleotide units can be replaced with alpha anomers to render the oligonucleotide more resistant to nucleases. Further, an intercalating agent such as ethidium bromide, or the like, can be attached to the 3 end of the alpha oligonucleotide to stabilize the triple helix. For information on the generation of oligonucleotides suitable for triple helix formation see Griffin et al. (Science 245:967-971 (1989), which is hereby incorporated by this reference).

### **EXAMPLE 44**

# Identification of Bacterial Strains from Isolated Specimens by PCR

Classical bacteriological methods for the detection of various bacterial species are time consuming and costly. These methods include growing the bacteria isolated from a subject in specialized media, cultivation on selective agar media, followed by a set of confirmation assays that can take from 8 to 10 days or longer to complete. Use of the identified sequences of the present invention provides a method to dramatically reduce the time necessary to detect and identify specific bacterial species present in a sample.

In one exemplary method, bacteria are grown in enriched media and DNA samples are isolated from specimens of, for example, blood, urine, stool, saliva or central nervous system fluid by conventional methods. A panel of PCR primers based on identified sequences unique to various species of microorganisms are then utilized in accordance with Example 12 to amplify DNA of approximately 100-200 bases in length from the specimen. A separate PCR reaction is set up for each pair of PCR primers and after the PCR reaction is complete, the reaction mixtures are assayed for the presence of PCR product. The presence of bacteria from the species to which the PCR primer pairs belong is determined by the presence or absence of a PCR product in the various test PCR reaction tubes.

Although the PCR reaction is used to assay the isolated sample for the presence of various bacterial species, other assays such as the Southern blot hybridization are also contemplated.

### WHAT IS CLAIMED IS:

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- 1. A purified or isolated nucleic acid sequence consisting essentially of one of SEQ ID NOs: 405-485, wherein said nucleic acid inhibits microorganism proliferation.
- 2. The nucleic acid sequence of Claim 1, wherein said nucleic acid sequence is complementary to at least a portion of a coding sequence of a gene whose expression is required for microorganism proliferation.
- 3. The nucleic acid sequence of Claims 1 or 2, wherein said nucleic acid comprises a fragment of one of SEO ID NOs. 405-485, said fragment selected from the group consisting of fragments comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEO ID NOs: 405-485.
- 4. The nucleic acid sequence of Claim 3, wherein said nucleic acid sequence is complementary to a coding sequence of a gene whose expression is required for microorganism proliferation.
- 5. A vector comprising a promoter operably linked to a nucleic acid comprising a sequence selected from the group consisting of SEQ ID NOs. 405-485.
- 6. The vector of Claim 5, wherein said promoter is active in an organism selected from the group consisting of Escherichia coli, Staphylococcus aureus, Pseudomonas aeruginosa, Enterobacter cloacae, Helicobacter pylori, Neisseria gonorrhoeae, Enterococcus faecalis, Streptococcus pneumoniae, Haemophilus influenzae, Salmonella typhimurium, Saccharomyces cerevisiae, Candida albicans, Cryptococcus neoformans, Aspergillus fumigatus, Klebsiella pneumoniae, Salmonella typhi, Salmonella paratyphi, Salmonella cholerasuis, Staphylococcus epidermidis, Mycobacterium tuberculosis, Mycobacterium leprae, Treponema pallidum, Bacillus anthracis, Yersinia pestis, Clostridium botulinum, campylobacter jejuni, Chlamydia trachomatus, Chlamydia pneumoniae or any species falling within the genera of any of the above species.
  - 7. A host cell containing the vector of Claim 5 or Claim 6.
- 8. A purified or isolated nucleic acid consisting essentially of the coding sequence of one of SEQ ID NOs: 82-88, 90-242.
- 9. A fragment of the nucleic acid of Claim 8, said fragment comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEQ ID NOs: 82:88, 90-242.
  - A vector comprising a promoter operably linked to the nucleic acid of Claim 8 or Claim 9.
- 11. A purified or isolated nucleic acid comprising a nucleic acid sequence complementary to at least a portion of an intragenic sequence, intergenic sequence, sequences spanning at least a portion of two or more genes, 5' noncoding region, or 3' noncoding region within an operon encoding a polypeptide comprising a sequence selected from the group consisting of SEO ID NOs: 243-357, 359-398.
- 12. A purified or isolated nucleic acid comprising a nucleic acid having at least 70% homology to a sequence selected from the group consisting of SEO ID NOs 405-485, 82-88, 90-242 or the sequences complementary thereto as determined using BLASTN version 2.0 with the default parameters.

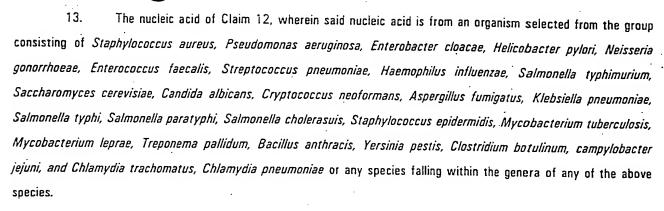
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- 14. A purified or isolated nucleic acid consisting essentially of a nucleic acid encoding a polypeptide having a sequence selected from the group consisting of SEQ ID NOs.: 243-357, 359-398.
- 15. A vector comprising a promoter operably linked to a nucleic acid encoding a polypeptide having a sequence selected from the group consisting of SEQ ID NOs.: 243-357, 359-398.
  - 16. A host cell containing the vector of Claim 15.
  - 17. A purified or isolated polypeptide comprising the sequence of one of SEQ ID NOs: 243-357, 359-398.
- 18. A purified or isolated polypeptide comprising a fragment of one of the polypeptides of SEQ ID NOs. 243-357, 359-398, said fragment selected from the group consisting of fragments comprising at least 5, at least 10, at least 20, at least 30, at least 40, at least 50, at least 60 or more than 60 consecutive amino acids of one of the polypeptides of SEQ ID NOs.: 243-357, 359-398.
  - 19. An antibody capable of specifically binding the polypeptide of Claim 17 or Claim 18.
- 20. A method of producing a polypeptide, comprising introducing a vector comprising a promoter operably linked to a nucleic acid encoding a polypeptide having a sequence selected from the group consisting of SEQ ID NOs. 243-357, 359-398 into a cell.
  - 21. The method of Claim 20, further comprising the step of isolating said protein.
- 22. A method of inhibiting proliferation comprising inhibiting the activity or reducing the amount of a polypeptide having a sequence selected from the group consisting of SEQ ID NOs. 243-357, 359-398 or inhibiting the activity or reducing the amount of a nucleic acid encoding said polypeptide.
- 23. A method for identifying compounds which influence the activity of a polypeptide required for proliferation comprising:

contacting a polypeptide having a sequence selected from the group consisting of 243-357, 359-398 with a candidate compound; and

determining whether said compound influences the activity of said polypeptide.

- 24. The method of Claim 23, wherein said activity is an enzymatic activity.
- 25. The method of Claim 23, wherein said activity is a carbon compound catabolism activity.

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- The method of Claim 23, wherein said activity is a biosynthetic activity.
- 27. The method of Claim 23, wherein said activity is a transporter activity.
- 28. The method of Claim 23, wherein said activity is a transcriptional activity.
- 29. The method of Claim 23, wherein said activity is a DNA replication activity.
- 30. The method of Claim 23, wherein said activity is a cell division activity.
- 31. A method for assaying compounds for the ability to reduce the activity or level of a polypeptide required for proliferation, comprising:

providing a target, wherein said target comprises the coding sequence of a sequence selected from the group consisting of SEO ID NOs. 82-88, 90-242;

contacting said target with a candidate compound; and measuring an activity of said target.

- 32. The method of Claim 31, wherein said target is a messenger RNA molecule transcribed from a coding region of one of SEQ ID. NOs.: 82-88, 90-242 and said activity is translation of said messenger RNA.
- 33. The method of Claim 32, wherein said target is a coding region of one of SEQ ID. NOs. 82-88, 90-242 and said activity is transcription of said messenger RNA.
  - 34. A compound identified using the method of Claim 31.
- 35. A method for identifying compounds which reduce the activity or level of a gene product required for cell proliferation comprising the steps of:

expressing an antisense nucleic acid against a nucleic acid encoding said gene product in a cell to reduce the activity or amount of said gene product in said cell, thereby producing a sensitized cell;

contacting said sensitized cell with a compound; and

determining whether said compound inhibits the growth of said sensitized cell to a greater extent than said compound inhibits the growth of a nonsensitized cell.

- 36. The method of Claim 35, wherein said cell is selected from the group consisting of bacterial cells, fungal cells, plant cells, and animal cells.
  - 37. The method of Claim 36, wherein said cell is an E. coli cell.
- 38. The method of Claim 36, wherein said cell is from an organism selected from the group consisting of Staphylococcus aureus, Pseudomonas aeruginosa, Enterobacter cloacae, Helicobacter pylori, Neisseria gonorrhoeae, Enterococcus faecalis, Streptococcus pneumoniae, Haemophilus influenzae, Salmonella typhimurium, Saccharomyces cerevisiae, Candida albicans, Cryptococcus neoformans, Aspergillus fumigatus, Klebsiella pneumoniae, Salmonella typhi, Salmonella paratyphi, Salmonella cholerasuis, Staphylococcus epidermidis, Mycobacterium tuberculosis, Mycobacterium leprae, Treponema pallidum, Bacillus anthracis, Yersinia pestis, Clostridium botulinum, campylobacter jejuni, and Chlamydia trachomatus, Chlamydia pneumoniae or any species falling within the genera of any of the above species.

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- 39. The method of Claim 35, wherein said antisense nucleic acid is transcribed from an inducible promoter.
- 40. The method of Claim 39, further comprising the step of contacting said cell with a concentration of inducer which induces said antisense nucleic acid to a sublethal level.
- 41. The method of Claim 40, wherein said sub-lethal concentration of said inducer is such that growth inhibition is 8% or more.
  - 42. The method of Claim 40, wherein said inducer is isopropyl 1-thio-β-D-galactoside.
- 43. The method of Claim 35, wherein growth inhibition is measured by monitoring optical density of a culture growth solution.
  - 44. The method of Claim 35, wherein said gene product is a polypeptide.
  - 45. The method of Claim 35, wherein said gene product is an RNA.
- 46. The method of Claim 44, wherein said gene product comprises a polypeptide having a sequence selected from the group consisting of SEQ ID NOs.: 243-357, 359-398.
  - 47. A compound identified using the method of Claim 35.
- 48. A method for inhibiting cellular proliferation comprising introducing a compound with activity against a gene corresponding to one of SEO ID NOs.: 82-88, 90-242 or with activity against the product of said gene into a population of cells expressing a gene.
- 49. The method of Claim 48, wherein said compound is an antisense oligonucleotide comprising a sequence selected from the group consisting of SEQ ID NOs.: 405-485, or a proliferation-inhibiting portion thereof.
- 50. The method of Claim 49, wherein said proliferation inhibiting portion of one of SEQ ID NOs. 405-485 is a fragment comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEQ ID NOs: 405-485.
  - 51. The method of Claim 48, wherein said compound is a triple helix oligonucleotide.
- 52. A preparation comprising an effective concentration of an antisense oligonucleotide comprising a sequence selected from the group consisting of SEO ID NOs.: 405-485, or a proliferation-inhibiting portion thereof in a pharmaceutically acceptable carrier.
- 53. The preparation of Claim 52, wherein said proliferation-inhibiting portion of one of SEO ID NOs. 405-485 comprises at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEO ID NOs: 405-485.
- 54. A method for inhibiting the expression of a gene in an operon required for proliferation comprising contacting a cell in a cell population with an antisense nucleic acid, said cell expressing a gene corresponding to one of SEQ ID NOs.: 82-88, 90-242, wherein said antisense nucleic acid comprises at least a proliferation-inhibiting portion of said operon in an antisense orientation that is effective in inhibiting expression of said gene.

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- 55. The method of Claim 54, wherein said antisense nucleic acid is complementary to a sequence of a gene comprising one or more of SEO ID NOs.: 82-88, 90-242.
- 56. The method of Claim 54, wherein said antisense nucleic acid is a sequence of one of SEQ ID NOs.: 405-485, or a portion thereof.
- 57. The method of Claim 54, wherein said cell is contacted with said antisense nucleic acid by introducing a plasmid which expresses said antisense nucleic acid into said cell population.
- 58. The method of Claim 54, wherein said cell is contacted with said antisense nucleic acid by introducing a phage which expresses said antisense nucleic acid into said cell population.
- 59. The method of Claim 54, wherein said cell is contacted with said antisense nucleic acid by introducing a sequence encoding said antisense nucleic acid into the chromosome of said cell into said cell population.
- 60. The method of Claim 54, wherein said cell is contacted with said antisense nucleic acid by introducing a retron which expresses said antisense nucleic acid into said cell population.
- 61. The method of Claim 54, wherein said cell is contacted with said antisense nucleic acid by introducing a ribozyme into said cell-population, wherein a binding portion of said ribozyme is complementary to said antisense oligonucleotide.
- 62. The method of Claim 54, wherein said cell is contacted with said antisense nucleic acid by introducing a liposome comprising said antisense oligonucleotide into said cell.
- 63. The method of Claim 54, wherein said cell is contacted with said antisense nucleic acid by electroporation.
- 64. The method of Claim 54, wherein said antisense nucleic acid is a fragment comprising at least 10, at least 20, at least 25, at least 30, at least 50 or more than 50 consecutive bases of one of SEO ID NOs: 82-88, 90-242.
  - 65. The method of Claim 54 wherein said antisense nucleic acid is an oligonucleotide.
  - 66. A method for identifying bacterial strains comprising the steps of: providing a sample containing a bacterial species; and

identifying a bacterial species using a species specific probe having a sequence selected from the group consisting of SEO ID NOs. 405-485, 82-88, 90-242.

- 67. A method for identifying a gene in a microorganism required for proliferation comprising:
- (a) identifying an inhibitory nucleic acid which inhibits the activity of a gene or gene product required for proliferation in a first microorganism;
  - (b) contacting a second microorganism with said inhibitory nucleic acid;
- (c) determining whether said inhibitory nucleic acid from said first microorganism inhibits proliferation of said second microorganism; and

- (d) identifying the gene in said second microorganism which is inhibited by said inhibitory nucleic acid.
- 68. A method for assaying a compound for the ability to inhibit proliferation of a microorganism comprising:
  - (a) identifying a gene or gene product required for proliferation in a first microorganism;
  - (b) identifying a homolog of said gene or gene product in a second microorganism;
- (c) identifying an inhibitory nucleic acid sequence which inhibits the activity of said homolog in said second microorgansim;
- (d) contacting said second microorganism with a proliferation-inhibiting amount of said inhibitory nucleic acid, thus sensitizing said second microorganism;
  - (e) contacting the sensitized microorganism of step (d) with a compound; and
- (f) determining whether said compound inhibits proliferation of said sensitized microorganism to a greater extent than said compound inhibits proliferation of a nonsensitized microorganism.
- 69. The method of Claim 68, wherein said step of identifying a gene involved in proliferation in a first microorganism comprises:

introducing a nucleic acid comprising a random genomic fragment from said first microorganism operably linked to a promoter wherein said random genomic fragment is in the antisense orientation; and

comparing the proliferation of said first microorganism transcribing a first level of said random genomic fragment to the proliferation of said first microorganism transcribing a lower level of said random genomic fragment, wherein a difference in proliferation indicates that said random genomic fragment comprises a gene involved in proliferation.

- 70. The method of Claim 69, wherein said step of identifying a homolog of said gene in a second microorganism comprises identifying a homologous nucleic acid or a nucleic acid encoding a homologous polypeptide in a database using an algorithm selected from the group consisting of BLASTN version 2.0 with the default parameters and FASTA version 3.0t78 algorithm with the default parameters.
- 71. The method of Claim 69, wherein said step of identifying a homolog of said gene in a second microorganism comprises identifying a homologous nucleic acid or a nucleic acid encoding a homologous polypeptide by identifying nucleic acids which hybridize to said first gene.
- 72. The method of Claim 69, wherein the step of identifying a homolog of said gene in a second microorganism comprises expressing a nucleic acid which inhibits the proliferation of said first microorganism in said second microorganism.
  - 73. The method of Claim 69, wherein said inhibitory nucleic acid is an antisense nucleic acid.
- 74. The method of Claim 69, wherein said inhibitory nucleic acid comprises an antisense nucleic acid to a portion of said homolog.

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- 75. The method of Claim 69, wherein said inhibitory nucleic acid comprises an antisense nucleic acid to a portion of the operon encoding said homolog.
- 76. The method of Claim 69, wherein the step of contacting the second microorganism with a proliferation-inhibiting amount of said nucleic acid sequence comprises directly contacting said second microorganism with said nucleic acid.
- 77. The method of Claim 69, wherein the step of contacting the second microorganism with a proliferation-inhibiting amount of said nucleic acid sequence comprises expressing an antisense nucleic acid to said homolog in said second microorganism.
  - 78. A compound identified using the method of Claim 68.
  - 79. A method of assaying a compound for the ability to inhibit proliferation comprising:
  - (a) identifying an inhibitory nucleic acid sequence which inhibits the activity of a gene or gene product required for proliferation in a first microorgansim;
  - (b) contacting a second microorganism with a proliferation-inhibiting amount of said inhibitory nucleic acid, thus sensitizing said second microorganism;
    - (c) contacting the proliferation inhibited microorganism of step (b) with a compound; and
- (d) determining whether said compound inhibits proliferation of said sensitized second microorganism to a greater extent than said compound inhibits proliferation of a nonsensitized second microorganism.
- 80. The method of Claim 79, wherein said inhibitory nucleic acid is an antisense nucleic acid which inhibits the proliferation of said first microorganism.
- 81. The method of Claim 79, wherein said inhibitory nucleic acid comprises a portion of an antisense nucleic acid which inhibits the proliferation of said first microorganism.
- 82. The method of Claim 79, wherein said inhibitory nucleic acid comprises an antisense molecule against the entire coding region of the gene involved in proliferation of the first microorganism.
- 83. The method of Claim 79, wherein said inhibitory nucleic acid comprises an antisense nucleic acid to a portion of the operon encoding the gene involved in proliferation of the first microorganism.
  - 84. A compound identified using the method of Claim 79.
- 85. A method for assaying compounds for activity against a biological pathway required for proliferation comprising:

sensitizing a cell by expressing an antisense nucleic acid against a nucleic acid encoding a gene product required for proliferation in a cell to reduce the activity or amount of said gene product;

contacting the sensitized cell with a compound; and

determining whether said compound inhibits the growth of said sensitized cell to a greater extent than said compound inhibits the growth of an nonsensitized cell.

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- 86. The method of Claim 85, wherein said cell is selected from the group consisting of bacterial cells, fungal cells, plant cells, and animal cells.
  - 87. The method of Claim 86, wherein said cell is an E. coli cell.
- 88. The method of Claim 85, wherein said cell is from an organism selected from the group consisting of Staphylococcus aureus, Pseudomonas aeruginosa, Enterobacter cloacae, Helicobacter pylori, Neisseria gonorrhoeae, Enterococcus faecalis, Streptococcus pneumoniae, Haemophilus influenzae, Salmonella typhimurium, Saccharomyces cerevisiae, Candida albicans, Cryptococcus neoformans, Aspergillus fumigatus, Klebsiella pneumoniae, Salmonella typhi, Salmonella paratyphi, Salmonella cholerasuis, Staphylococcus epidermidis, Mycobacterium tuberculosis, Mycobacterium leprae, Treponema pallidum, Bacillus anthracis, Yersinia pestis, Clostridium botulinum, campylobacter jejuni, and Chlamydia trachomatus, Chlamydia pneumoniae or any species falling within the genera of any of the above species.
  - 89. The method of Claim 85, wherein said antisense nucleic acid is transcribed from an inducible promoter.
- 90. The method of Claim 89, further comprising contacting the cell with an agent which induces expression of said antisense nucleic acid from said inducible promoter, wherein said antisense nucleic acid is expressed at a sublethal level.
- 91. The method of Claim 90, wherein said sublethal level of said antisense nucleic acid inhibits proliferation by 8% or more.
  - 92. The method of Claim 90, wherein said agent is isopropyl-1-thio-β-D-galactoside (IPTG).
- 93. The method of Claim 91, wherein inhibition of proliferation is measured by monitoring the optical density of a liquid culture.
- 94. The method of Claim 85, wherein said gene product comprises a polypeptide having a sequence selected from the group consisting of SEQ ID NOs: 243-357, 359-398.
  - 95. A compound identified using the method of Claim 85.
  - 96. A method for assaying a compound for the ability to inhibit cellular proliferation comprising:

    contacting a cell with an agent which reduces the activity or level of a gene product required for proliferation of said cell;

contacting said cell with said compound; and

determining whether said compound reduces proliferation to a greater extent than said compound reduces proliferation of cells which have not been contacted with said agent.

- 97. The method of Claim 96, wherein said agent which reduces the activity or level of a gene product required for proliferation of said cell comprises an antisense nucleic acid to a gene or operon required for proliferation.
- 98. The method of Claim 96, wherein said agent which reduces the activity or level of a gene product required for proliferation of said cell comprises an antibiotic.

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- 99. The method of Claim 96, wherein said cell contains a temperature sensitive mutation which reduces the activity or level of said gene product required for proliferation of said cell.
- 100. The method of Claim 99, wherein said antisense nucleic acid is directed against the nucleic acid encoding the same functional domain of said gene product required for proliferation of said cell to which said antisense nucleic acid is directed.
- 101. The method of Claim 99, wherein said antisense nucleic acid is directed against the nucleic acid a different functional domain of said gene product required for proliferation of said cell than the functional domain to which said antisense nucleic acid is directed.
  - 102. A compound identified using the method of Claim 96.
- 103. A method for identifying the pathway in which a proliferation required nucleic acid or its gene product lies comprising:

expressing a sublethal level of an antisense nucleic acid directed against said proliferation-required nucleic acid in a cell;

contacting said cell with an antibiotic, wherein the biological pathway on which said antibiotic acts is known; and

determining whether said cell has a substantially greater sensitivity to said antibiotic than a cell which does not express said sublethal level of said antisense nucleic acid.

- 104. A method for determining the pathway on which a test compound acts comprising:
- (a) expressing a sublethal level of an antisense nucleic acid directed against a proliferation-required nucleic acid in a cell, wherein the biological pathway in which said proliferation-required nucleic acid lies is known,
  - (b) contacting said cell with said test compound; and
- (c) determining whether said cell has a substantially greater sensitivity to said test compound than a cell which does not express said sublethal level of said antisense nucleic acid.
  - 105. The method of Claim 104, further comprising:
  - (d) expressing a sublethal level of a second antisense nucleic acid directed against a second proliferation-required nucleic acid in said cell, wherein said second proliferation-required nucleic acid is in a different biological pathway than said proliferation-required nucleic acid in step (a); and
  - (e) determining whether said cell has a substantially greater sensitivity to said test compound than a cell which does not express said sublethal level of said second antisense nucleic acid.
  - 106. A purified or isolated nucleic acid consisting essentially of one of SEQ ID NOs: 358, 399-402.
  - 107. A compound identified using the method of Claim 23.
- 108. A compound which interacts with the gene or gene product of a nucleic acid comprising a sequence of one of SEQ ID NOs: 82-88, 90-242 to inhibit proliferation.

- 109. A compound which interacts with a polypeptide comprising one of SEO ID NOs. 243-357, 359-398 to inhibit proliferation.
- 110. A compound which interacts with a nucleic acid comprising one of SEQ ID NOs: 358, 399-402 to inhibit proliferation.

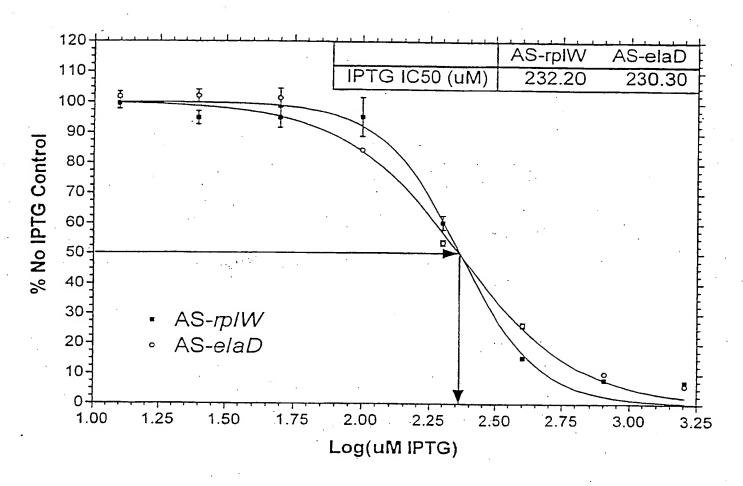
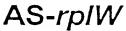


Fig. 1



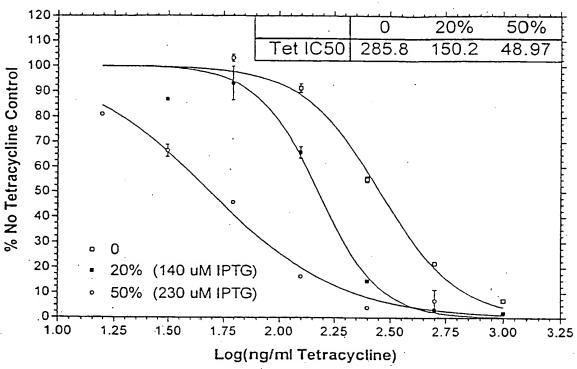


Fig. 2a



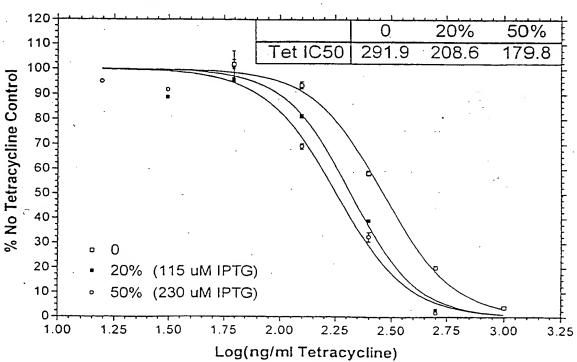


Fig. 2b

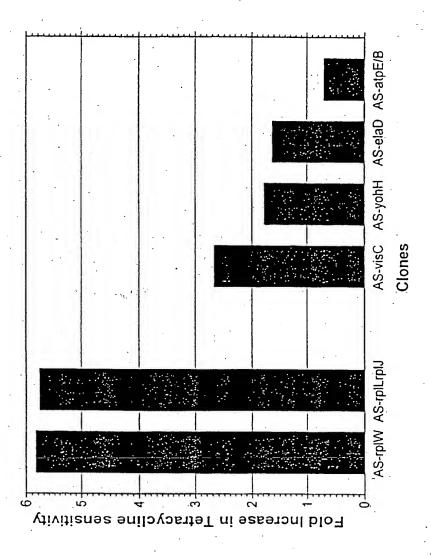


Fig. 3

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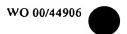
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<210> 19

```
<211> 588
       <212> DNA
       <213> E. Coli
       <220>
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       <222> (1) ... (588)
       <223> n = A, T, C \text{ or } G
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                                                                          60
cacgcggaac agttaaaacc aaaaacagtg ttatgggtgg attctctgta tccggcgtta
                                                                         120
attgttaact ggttaacgtc acctggaggc accaggcact gcatcacaaa attcattgtt
                                                                         180
gaggacgcga taatgaaaac gttattacca aacgttaata cgtctgaagg ttgttttgaa
                                                                         240
attggtgtca ctatcagtaa cccagtattt actgaagatg ccattaacaa gagaaaacaa
                                                                         300
gaacgggagc tattaaataa aatatgcatt gtttcaatgc tggctcgttt acgtctgatg
                                                                         360
ccaaaaggat gtgcacaatg aattcagcat ttgtgcttgt tctgacagtt tttcttgttt
                                                                         420
ccggagagcc agttgatatt gcagtcagtg ttcacaggac aatgcangag tgtatgactg
                                                                         480
cagcaacccg aacagaaaat toocggtaac tgttacccgg tcgataaagt tattcaccag
                                                                         540
gataatatcg aaatcccggc aggtctttaa aacagttccg taataaat
                                                                         588
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      <211> 101
      <212> DNA
      <213> E. Coli
      <400> 20
gatccagcaa gaagatgcgg ttgtaccgtc atcacgcaga tgcgcaaagc tactcagcaa
                                                                          60
ctgacctttc ttcgcaataa gcacgccatt agcgtcatag a
                                                                         101
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      <211> 465
      <212> DNA
      <213> E. Coli
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tegegtgttt acctteaaca teggtaactt tetggeggat agttteaegg taageaacet
                                                                         60
gcggtttacc tacgttcgct tcaacgttga attcacgctt catacggtca acgatgatgt
                                                                        120
cgaggtgcag ttcgcccata cccgcgatga tggtctggtt agattcttcg tcagtccata
                                                                        180
cacggaaaga cgggtcttct ttagccagac ggcccagagc cagacccatt ttttcctggt
                                                                        240
cagetttggt tttcggttca actgcgatgg agattaccgg ctcagggaat tccatacgtt
                                                                        300
ccagaatgat cggcgcatcc gggtcacaca gggtgtcacc agtggttacg tctttcagac
                                                                        360
cgatagcage agegatgteg ecegegegaa ettetttgat etetteaegt ttgttagegt
                                                                        420
gcatctgaac gatacgaccg aaacgctcac gtgcagcttt cacgg
                                                                        465
      <210> 22 -
      <211> 859
      <212> DNA
      <213> E. Coli
      <220>
      <221> misc_feature
      <222> (1)...(859)
      <223> n = A, T, C or G
      <400> 22
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                                                                         60
cagccagttt aaacgccagt tcagaggagt caacgtcatg gtaagaaccg aagtgcagac
                                                                        120
gaatacccat gtctactacc gggtagcctg ccageggacc tgctttcage tgttcctgga
                                                                        180
tacctttatc aacggccggg atgtattcgc cagggattac accaccttta atgtcgttga
                                                                        240
tgaactcgta geettteggg tttgaaceeg geteeagegg gtacatgteg ataacaacat
                                                                        300
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471

WO 00/44	906		•		· PC	T/US00/0220
gaccatactg ac tctggcggat ag attcacgctt ca tggtctggtt ag gggccanagc ca gattaccggc tc anggngtacc ag gccnaacttc tt aaaagngtta an cggggtnaac cc	tttcacgg tacggtca attcttcg gacccatt anggaatt gggggtac tggaacnn ngccantt	taagcaacct acgatgatgt tcagtccata ttttcctggt tccatacctt ntntttttaa tttaccggtt	gcggtttacc cgaggtgcag cacggaaaga cagctttggt ccaggaatga nancgattgc ggtaaccngc	tacgttcgct ttcgcccata cgggtcttct tttcggtcaa tcggcgcatt cagcancgga cttttnaacn	tcaacgttga cccgcgatga ttagccagac ctgcgatgga ccggtcaaac tntnncccgn atccaaccga	420 480 540 600 660 720 780
<210> 2 <211> 2 <212> D <213> E	69 NA . Coli		• .	·		
<400> 2 ctttcttaaa gc agtcaacgtc at ctgccagcgg ac cgccagggat ta ccggctccag cg	cttctta ggtaagaa ctgctttc caccacct	ccgaagtgca agctgttcct ttaatgtcgt	gacgaatacc ggataccttt	catgtctact atcaacggcc	accgggtagc gggatgtatt	60 120 180 240 269
<210> 2 <211> 3 <212> D <213> E	30 NA					
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<210> 2: <211> 4 <212> DI <213> E	71 NA					
<222> (	isc_featu 1)(471 = A,T,C	)	,			
<400> 2! gttttgggga gat atgactgatt gccctgatccttc tgt tcggacgcac ctt agtgaatgat tattattataatt aat	tgtaaggg cgatacct ttcttata ttaataac tgctaatg tactctac	gattaaacgg acacaaggaa tataaataag tcatcaatta ttccagagta	gtcatcaaaa acgtacttaa tgtctgggca aataaatata gaatattaaa	tcatcattgc ggtgccgtcc gatactatat atggcgttaa ttttatccgc	tgttttacag ggtgaaccag aaattaactt ggcttcccag	60 120 180 240 300 360

<210> 26 <211> 379 <212> DNA <213> E. Coli

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tantattggg ggattnggcc cncctttttg ncaggttggg gtcntctnat g

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<220>
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      <222> (1)...(379)
      <223> n = A, T, C or G
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                                                                       60
ttaaacgggt catcaaaatc atcattgctg ttttacagct gatccttctg ttcttataac
                                                                      120
acaaggaaac gtacttaagg tgcgtccggt gaaccagtcg gacgcacctt taataactat
                                                                      180
aaataagtgt ctgggcagat actatataaa ttaacttagt gaatgattat gctaatgtca
                                                                      240
tcaattaaat aaatataatg gcgttaaggc ttcccagtaa tataattaat actctacttc
                                                                      300
cagagragaa tattaaattt tatccgcgtg gtgcatcagc acaaatttat cccacaactg
                                                                      3.60
ttcttctgtc tcgacatgc
                                                                      379
      <210> 27
      <211> 799
      <212> DNA
      <213> E. Coli
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                                                                       60
caaaaaacac taaatcaaaa aataatggca ttagaaaata taatgcgaaa acggaggtga
                                                                      120
aattagttta tttcaaatga ggaaaatctc ccggcgaaaa aaccgggaga tgaaagtgtg
                                                                      180
atgggtatca aataaacaac agaggagaaa tttttaacgc agccattcag gcaaatcgtt
                                                                     240
taatcccatt gcctggcgga taagttgcgg cttaacgcca ggaagcgtgt cggccagttt
                                                                      300
caaaccaata tcacgcagca gttttttcgc cggattggta ccggaaaaca gatcgcggaa
                                                                      360
tecetgeata ceagecagea teaacgeege actgtgettg eggetaeget catagegaeg
                                                                      420
cagataaatg tactgcccga tgtctgggat ccgtcgacct qcaqccaaqc ttqqqctttt
                                                                     480
cagcctgata cagattaaat cagaacgcag aagcggtctg ataaaacaga atttgcctgg
                                                                     540
cggcagtage geggtggtee cacetgaeee catgeegaae teagaagtga aacqeeeqta
                                                                     600
gcgcccgatg gtagtgtggg gtctccccat gcgagagtag ggaactgcca ggcatcaaat
                                                                     660
aaaacgaaag gctcagtcga aagactgggc ctttcggttt atctggtggt tgtcggtgaa
                                                                     720
cgctctctga gtaggacaaa tccgccggga gcggattttg aacgttgcga aacaaccggc
                                                                     780
ccggaaaggg gtgggggct
                                                                     799
      <210> 28
      <211> 636
      <212> DNA
      <213> E. Coli
      <220>
     <221> misc_feature
     <222> (1)...(636)
     <223> n = A, T, C or G
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acaataaaca acgaatcagg gcatttgata gtcaataccg caattctatc aggagatata
                                                                     120
gtcactctaa gaggaggaga aattaggttg gtattatagc ttgtgcgcgc catgattggc
                                                                     180
gcgcaattta aacttagtgc tttacatcgc tattgtcttg atttctttga attatttat
                                                                     240
aaattaaaaa aacgactgtt atgtataagc aaaggtcgaa cgaaaaatac attccaaata
                                                                     300
aatgettget taaateteta tateetteee egaaaaatga cacataaaat tgagatatte
                                                                     360
420
caataaaaaa taataacaat gatataaatc taatgttttt aaatatattg tcttttatgt
                                                                     480
tagtaatagt cgttagtatg tttgattctc catatattac gtgtagtttt ttatatacat
                                                                     540
ggaaataatt ntctttatac tgagacatca caccatcatc aaatggaagt ttgaagatgg
                                                                     600
tgcttggttt gctaaccaat aaaaagagtg cattcg
                                                                     636
     <210> 29
     <211> 757
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<212> DNA

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WO 00/44906
      <213> E. Coli
      <220>
      <221> misc feature
      <222> (1)...(757)
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                                                                         60
gatgaaaaaa acaacgatta ttatgatggg tgtggcgatt attgtcgtac tcggcactga
                                                                        120
gctgggatgg tggtaacgtc acctctaaaa aatagcaaag gctgcctgtg tgcagccttt
                                                                        180
gtgcaattta agcgttaact tttaatcttc ctgtagataa atagcacgac aatcgcacca
                                                                        240
ataacggcaa ccacgaagct gccaaaattg aagccatcga ctttaccaaa gccaaacagc
                                                                        300
gtgctgatcc atccgccgac tacggcaccg actatcccca gcaggatagt cataaagaat
                                                                        360
ccacctccat ctttacctgg catgatccac ttcgccagaa taccggcaat aagcccaaaa
                                                                        420
ataatccatg acagaatgcc cattgtttcc tcacttatct gttttgcatt agegggttag
                                                                        480
tcgctgataa aaagcatagc acaacatcgg gagggcaaga tttgtgacga gcatcacgga
                                                                        540
ggtttttttt gcgatggcgc agaaattgcg ccatcaacga tcagtgataa ttaccaacca
                                                                        600
caaacatcat gttcgttttc cgtgtcataa gaaccgtacg ggattcacca gatcttttat
                                                                        660
cacticaage eggeactict ggeaceagea aagteategg egtetetggt teataatega
                                                                        720
ccggaaacgc cattgctggt attggtgacn gtcacgg
                                                                        757
      <210> 30
      <211> 392
      <212> DNA
      <213> E. Coli
      <400> 30
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                                                                         60
acaaatatta ttgtgctgca ggtgttttag cgggttgttg atccacaggt tctaactgga
                                                                        120
agaccacate gacetgatea teaaaetgaa tageggeetg etegtaagtt teetgggegg
                                                                       180
acaccggcgc ggcatcggct ttcatcatcc gcaccattgg gctgggctga tagttggaaa
                                                                        240
catggtagcg cacgctatat accggcccca gtttacgatg aaagccgttc gccagttcct
                                                                        300
gegeetgatg aategegtta teaategetg cettaegege titgtettta taggeateeg
                                                                        360
gctgcgccac gcccagcgac acagaacgaa tt
                                                                        392
      <210> 31
      <211> 351
      <212> DNA
      <213> E. Coli
      <400> 31
ctatccttga tgaaaccgcg agcaaagata ggtgattacg tcatggtttt acagaaaatt
                                                                         60
acagaaaaag gaggcaatat cgggtaaagg cattagcccg acgaatacgt cgggctacaa
                                                                       120
atattattgt gctgcaggtg tttttagcggg ttgttgatcc acaggttcta actggaagac
                                                                       180
cacatcgacc tgatcatcaa actgaatagc ggcctgctcg taagtttcct gggcggacac
                                                                       240
cggcgcggca tcggctttca tcatccgcac cattgggctg ggctgatagt tggaaacatg
                                                                       300
gtagogoacg ctatatacog gooccagttt acgatgaaag cogttogoca g
                                                                       351
      <210> 32
      <211> 762
      <212> DNA
      <213> E. Coli
      <220>
      <221> misc_feature
      <222> (1)...(762)
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<223> n = A, T, C or G

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gtaataacac aggaaactat tttatctacg cgttagcgat agactgcttg catggcgaaa
                                                                        120
ggaggtaagc cgacgatttc agcgggacgc tgaaacggga aagcccctcc cgaggaaggg
                                                                        180
gccataaata aggaaagggt catgatgaag ctactcatca tcgtggtgct cttagtcata
                                                                        240
agetteeceg ettactaaga etaceaggge gggggaaace eegetetace etcacteetg
                                                                        300
aaagtatgcc ttcacgataa gattgtcaat ccgcaggctt tgtagtctgc gatcctgcca
                                                                        360
gcaaatattc tttgcgagtc gttacgcaat aatcacagag gaaactattt tattcacgcg
                                                                        420
ttagcgatag actgcattca gggcgaaagg aggtaagccg atgatttcag cgggacgctg
                                                                        480
aaacgggaaa gcctctcccg gagaagaggg cttttaataa ggaaagggtt atgatgaagc
                                                                        540
acgtcatcat actggtgata ctcttagtga ttagcttcca ggcttactaa gaacaccagg
                                                                        600
gggaggggga aacctcttcc taaccctcac ttctgaaatt gggtgctatg acgctggcgt
                                                                        660
tactgcttan cgctaccagt ttgtctgccc tggcggttgt aacgccagat cggtacccgt.
                                                                        720
ttggatattt taatgaaagc cgacaaatca atcancgtga cg
                                                                        762
     . <210> 33
      <211> 293
      <212> DNA
      <213> E. Coli
      <400> 33
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gatagactgc ttgcatggcg aaaggaggta agccgacgat ttcagcggga cgctgaaacg
                                                                        120
ggaaagcccc tcccgaggaa ggggccataa ataaggaaag ggtcatgatg aagctactca
                                                                        180
tcatcgtggt gctcttagtc ataagcttcc ccgcttacta agactaccag ggcggggaa
                                                                        240
accocgetet acceteacte etgaaagtat geetteacga taagattgte aat
                                                                        293
     <210> 34
      <211> 633
      <212> DNA
      <213> E. Coli
      <220>
      <221> misc_feature /
      <222> (1)...(633)
      <223> n = A, T, C or G
      <400> 34
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                                                                         60
tatggcagga aagatatgcg actgatatta cagatcccca aagtggagag tttatgacca
                                                                        120
ttaaaaataa gatgttgctg ggtgcgcttt tgctggttac cagtgccgcc tgqgccgcac
                                                                        180
cagccaccgc gggttcgacc aatacctcgg gaatttctaa gtatgagtta agtagtttca
                                                                        240
ttgctgactt taagcatttc aaaccagggg acaccgtacc agaaatgtac cgtaccgatg
                                                                        300
agtacaacat taagcagtgg cagttgcgta acctgcccgc gcctgatgcc qqqacqcact
                                                                       360
ggacctatat gggtggcgcg tacgtgttga tcagcgacac cgacggtaaa atcattaaaq
                                                                       420
cctacgacgg tgagattttt tatcatcgct aaaaaaaagcc ccctcatcat qaqqqqaaa
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concnence conceatece					720
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12237 II - A, I, C	OI G				
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catgctgtag ttaccacgac					180
acgaggtaca gcaatagtga					240
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ggcgggcagg acgcccgcca					660
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                                                                      120
taattgaaag tattgggttg ctgataattt gagctgttct attcttttta aatatctata
                                                                      180
taggicigti aatggatiti attittacaa tittitgigt tiaggcatat aaaaatcaac
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caggagagtt atgagctggc ggcgttttta gcctgcaaat tgaaagagta agagtcttcg
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gcgggaaatt attcccgcct tacttacggc gttgcgcatt ctcattgcac ccaaatttat
                                                                     240
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                                                                         240
                                                                         300
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 qttqttcgat gacaatcgqt qccccctgat gcggtgcctt catgccgaag aatttcaccc
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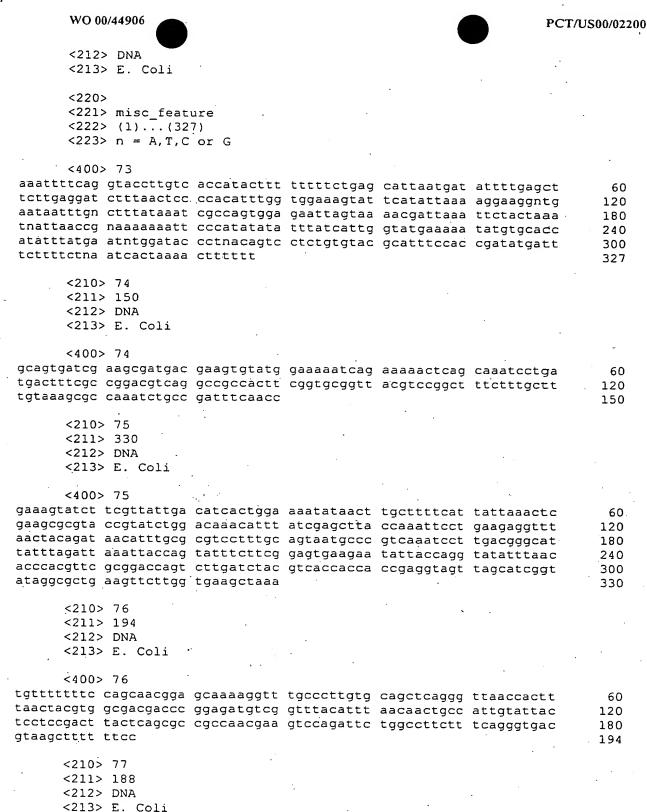
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120

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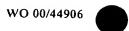
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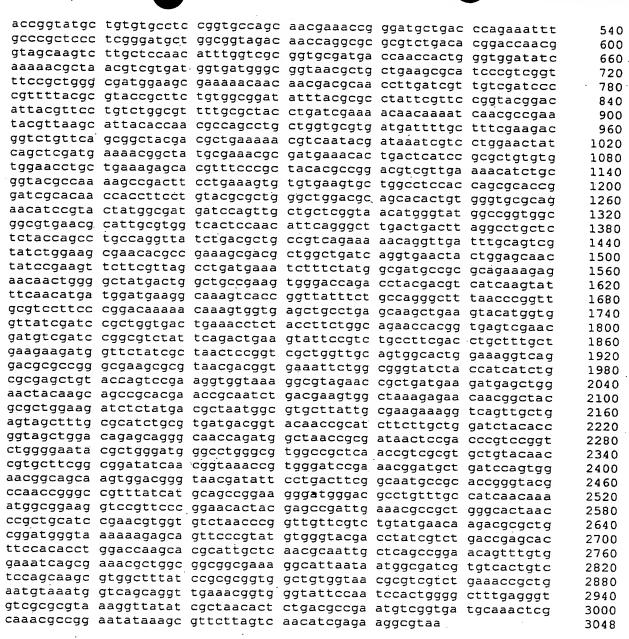
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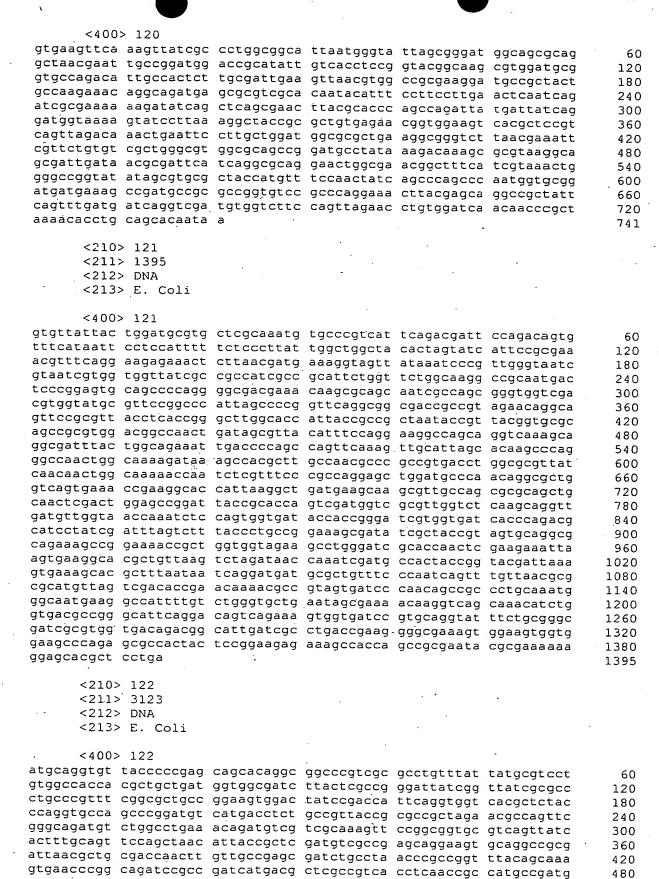
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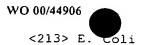
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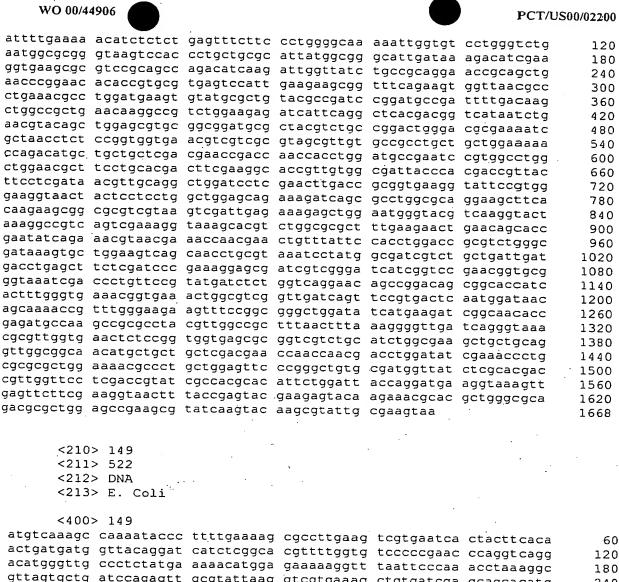
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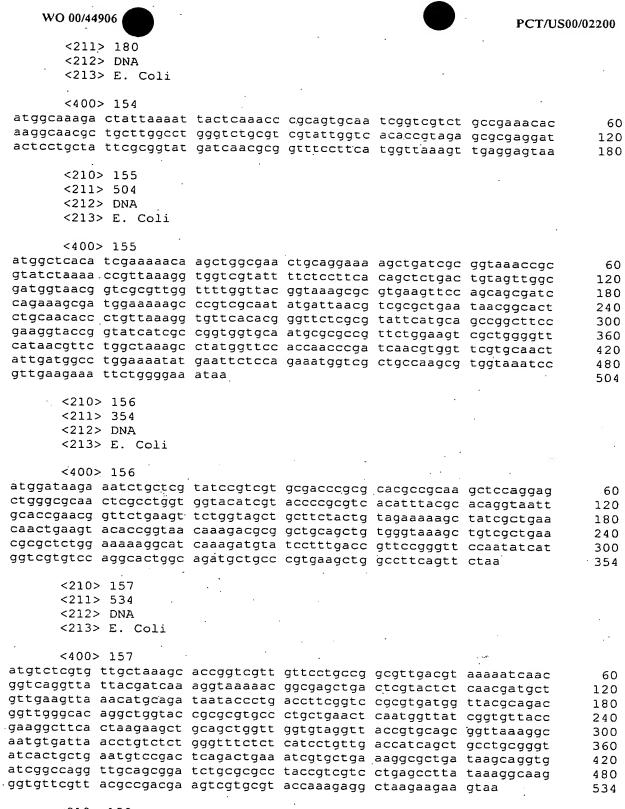
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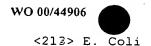
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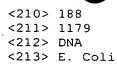
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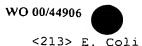
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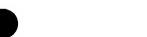
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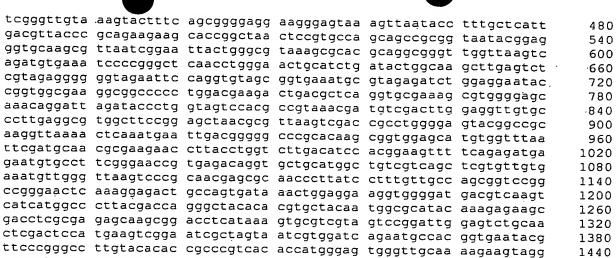
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1549



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<213> E. Coli

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<211> 203

<212> PRT

<213> E. Coli

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Gly Ser Val Ala Glu Ser Asn Ala Thr Gly Asn Pro Val Asn Leu Leu
                        55
Asp Gly Lys Leu Ser Phe Ser Leu Pro Ala Asp Met Thr Asp Gln Ser
                  . 70
                                        7.5
Gly Lys Leu Gly Thr Gln Ala Asn Asn Met His Val Trp Ser Asp Ala
                                    90
Thr Gly Gln Lys Ala Val Ile Val Ile Met Gly Asp Asp Pro Lys Glu
                               105
Asp Leu Ala Val Leu Ala Lys Arg Leu Glu Asp Gln Gln Arg Ser Arg
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Asp Pro Gln Leu Gln Val Val Thr Asn Lys Ala Ile Glu Leu Lys Gly
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                                            140
His Lys Met Gln Gln Leu Asp Ser Ile Ile Ser Ala Lys Gly Gln Thr
                    150
                                        155
Ala Tyr Ser Ser Val Ile Leu Gly Asn Val Gly Asn Gln Leu Leu Thr
                165
                                    170
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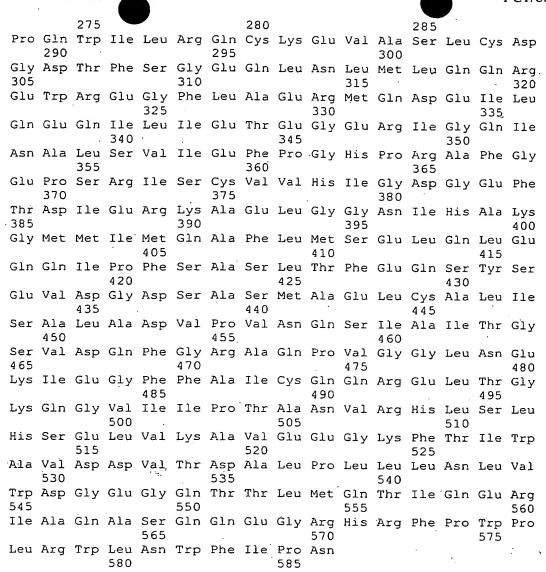


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Val	Pro	Ile	Ala	Gln 245	Asp	Gln	Val	Gly	Met 250	Tyr	Tyr	Gln	Gln	Pro 255	Gly
Gln	Gln	Leu	Ala 260	Thr	Trp	Ile	Val	Pro 265	Pro	Gly	Gln	Tyr	Phe 270	Met	Met
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Pro	Glu 290	Ala	Asn	Leu	Val	Gly 295	Arg	Ala	Thr	Ala	Ile 300	Trp	Met	Ser	Phe
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<210> 246 <211> 586 <212> PRT <213> E. Coli

<400> 246

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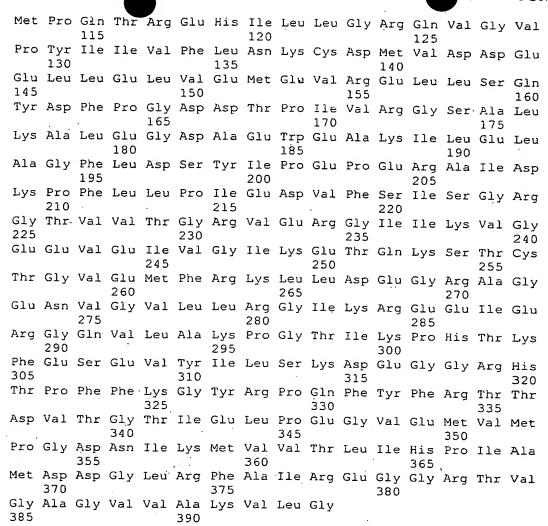


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<213> E. Coli

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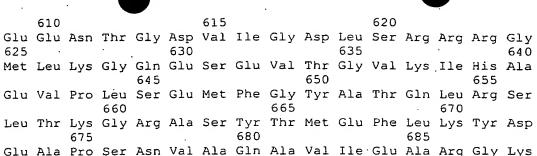
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 Glu
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 Phe
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<210> 248 <211> 704 <212> PRT <213> E. Coli

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			(									'			
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Asp 145	Arg	Met	Gly	Ala	Asn 150	Phe	Leu	Lys	Val	Val 155	Asn	Gln	Ile	Lys	Thr 160
Arg	Leu	Gly	Ala	Asn 165	Pro	Val	Pro		Gln 170	Leu	Ala	Ile	Gly	Ala 175	Glu ·
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Trp	Asn	Asp 195	Ala	Asp	Gln	Gly	Val 200	Thr	Phe	Glu	Tyr	Glu 205	Asp	Ile	Pro
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				405	Leu				410					415	
			420		Pro			425					430		
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	450					455					460		-		
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				485.					490				-	495	Gln
			500					505					510		Arg
		515					520	_		_		525			Gly
	530					535				•	540				Val
545					550				_	555	_				Gln 560
				565		•			570					575	Ile
		•	580					585		_			590		Ala
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<210> 249 <211> 179 <212> PRT <213> E. Coli

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<210> 250 <211> 124 <212> PRT <213> E. Coli

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Ala Arg Ser Lys Tyr Gly Val Lys Arg Pro Lys Ala

<210> 251 . <211> 165 <212> PRT <213> E. Coli

<400> 251

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<210> 252 <211> 121 <212> PRT <213> E. Coli

<400> 252

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 Ile
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 Glu
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 Ser

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 Asp
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 Val
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 Leu
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 Lys
 Phe
 Gly

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 Ala
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Val Leu Leu Gly Asn Pro Ala Glu Ile Asn Arg Val Ala Ala Ser Gln 440 Gly Val Glu Leu Gly Ala Gly Ile Glu Ile Val Asp Pro Glu Val Val 455 460 Arg Glu Ser Tyr Val Gly Arg Leu Val Glu Leu Arg Lys Asn Lys Gly 470 475 Met Thr Glu Thr Val Ala Arg Glu Gln Leu Glu Asp Asn Val Val Leu 490 Gly Thr Leu Met Leu Glu Gln Asp Glu Val Asp Gly Leu Val Ser Gly 500 505 Ala Val His Thr Thr Ala Asn Thr Ile Arg Pro Pro Leu Gln Leu Ile 520 Lys Thr Ala Pro Gly Ser Ser Leu Val Ser Ser Val Phe Phe Met Leu 535 540 Leu Pro Glu Gln Val Tyr Val Tyr Gly Asp Cys Ala Ile Asn Pro Asp 550 555 Pro Thr Ala Glu Gln Leu Ala Glu Ile Ala Ile Gln Ser Ala Asp Ser 565 570 Ala Ala Ala Phe Gly Ile Glu Pro Arg Val Ala Met Leu Ser Tyr Ser . 580 585 Thr Gly Thr Ser Gly Ala Gly Ser Asp Val Glu Lys Val Arg Glu Ala 600 605 Thr Arg Leu Ala Gln Glu Lys Arg Pro Asp Leu Met Ile Asp Gly Pro 615 620 , Leu Gln Tyr Asp Ala Ala Val Met Ala Asp Val Ala Lys Ser Lys Ala 630 635 Pro Asn Ser Pro Val Ala Gly Arg Ala Thr Val Phe Ile Phe Pro Asp 645 650 Leu Asn Thr Gly Asn Thr Thr Tyr Lys Ala Val Gln Arg Ser Ala Asp 660 665 Leu Ile Ser Ile Gly Pro Met Leu Gln Gly Met Arg Lys Pro Val Asn , . 680 Asp Leu Ser Arg Gly Ala Leu Val Asp Asp Ile Val Tyr Thr Ile Ala 695 Leu Thr Ala Ile Gln Ser Ala Gln Gln Gln 710

<210> 254 <211> 588 <212> PRT <213> E. Coli

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		130					135					140		-		
	Gln 145	Tyr	Leu	ı Glm	Val	Ser 150	Ser	Cys	Val	Pro	Ala 155	Leu	Glu	Gly	Cys	Asp
	Val	Asn	Gly	/ Ala	Ser 165	Phe	Thr	Leu	Glu	Gln 170	Met	Leu	Ala	Trp	Arg 175	Asp
	His	Pro	Glr	Val 180	Thr	Gly	Leu	Ala	Glu 185	Met		Asp	Tyr	Pro 190	Gly	Val
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	Leu	Thr 210	Leu	Asp	Gly	His	Cys 215	Pro		Leu	Gly	Gly 220	Lys	Glu	Leu	Asn
	225					230					235	Glu	Ser			Leu 240
					245					250					-255	Arg
				260		Arg			265					Leu -270	Ile	-
			275			Gln		280					285			
		290				Glu	295					300			_	
	305					Val 310		-			315				-	320
					325	His				330					335	
				340		Asp	•		345			•	•	350	_	
			355			Leu		360				•	365			
		370				Ser	375					380				
	385					Arg 390					395					400
					405	Lys				410			•		415	
				. 420		Ser			425				•	430		
			4.35			Ser		440		i,			445			
		450				Cys	455					460				
	465	1114	DC u	AIG	ALG	Thr 470	vaı	261	UTZ	Asp	5er 475	H1S	Asn	тте	vaı	Val 480
					485	Gļu				490	Ala				495	Ile
				500		Leu			505					510	Gln	
			515			Ile	•	520					525			
	•	530				Ile	535					540				
	545					Glu 550					555					5.60
					565					570			Leu	Phe	Asp 575	Gly.
•	Glu	Lys	Phe	Ala 580	Phe	Thr	Thr	Leu	Glu 585	Val	Thr	Glu	•			

<210> 255 <211> 408 <212> PRT <213> E. Coli

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<210> 256 <211> 299 <212> PRT

## <213> E. Coli

<400> 256 Met Ile Asp Met Thr Met Lys Val Gly Phe Ile Gly Leu Gly Ile Met Gly Lys Pro Met Ser Lys Asn Leu Leu Lys Ala Gly Tyr Ser Leu Val 25 Val Ala Asp Arg Asn Pro Glu Ala Ile Ala Asp Val Ile Ala Ala Gly 40 Ala Glu Thr Ala Ser Thr Ala Lys Ala Ile Ala Glu Gln Cys Asp Val-55 Ile Ile Thr Met Leu Pro Asn Ser Pro His Val Lys Glu Val Ala Leu 7.0 Gly Glu Asn Gly Ile Ile Glu Gly Ala Lys Pro Gly Thr Val Leu Ile 85 90 Asp Met Ser Ser Ile Ala Pro Leu Ala Ser Arg Glu Ile Ser Glu Ala 100 105 Leu Lys Ala Lys Gly Ile Asp Met Leu Asp Ala Pro Val Ser Gly Gly 120 Glu Pro Lys Ala Ile Asp Gly Thr Leu Ser Val Met Val Gly Gly Asp 135 140 Lys Ala Ile Phe Asp Lys Tyr Tyr Asp Leu Met Lys Ala Met Ala Gly 150 155 Ser Val Val His Thr Gly Glu Ile Gly Ala Gly Asn Val Thr Lys Leu 165 170 Ala Asn Gln Val Ile Val Ala Leu Asn Ile Ala Ala Met Ser Glu Ala 180 185 Leu Thr Leu Ala Thr Lys Ala Gly Val Asn Pro Asp Leu Val Tyr Gln 200 Ala Ile Arg Gly Gly Leu Ala Gly Ser Thr Val Leu Asp Ala Lys Ala 215 Pro Met Val Met Asp Arg Asn Phe Lys Pro Gly Phe Arg Ile Asp Leu 230 235 His Ile Lys Asp Leu Ala Asn Ala Leu Asp Thr Ser His Gly Val Gly 245 250 Ala Gln Leu Pro Leu Thr Ala Ala Val Met Glu Met Met Gln Ala Leu 265 Arg Ala Asp Gly Leu Gly Thr Ala Asp His Ser Ala Leu Ala Cys Tyr 280 Tyr Glu Lys Leu Ala Lys Val Glu Val Thr Arg

<210> 257 <211> 256 <212> PRT

<213> E. Coli

## <400> 257

 Met
 Asn
 Asn
 Asp
 Val
 Phe
 Pro
 Asn
 Lys
 Phe
 Lys
 Ala
 Ala
 Leu
 Ala
 Ala</th

90 Ile Pro Phe Val Glu Thr Lys Glu Glu Ala Glu Leu Ala Val Ala Ser 100 105 Thr Arg Tyr Pro Pro Glu Gly Ile Arg Gly Val Ser Val Ser His Arg 120 Ala Asn Met Phe Gly Thr Val Ala Asp Tyr Phe Ala Gln Ser Asn Lys 135 140 Asn Ile Thr Ile Leu Val Gln Ile Glu Ser Gln Gln Gly Val Asp Asn 150 155 Val Asp Ala Ile Ala Ala Thr Glu Gly Val Asp Gly Ile Phe Val Gly 170 Pro Ser Asp Leu Ala Ala Ala Leu Gly His Leu Gly Asn Ala Ser His 185 Pro Asp Val Gln Lys Ala Ile Gln His Ile Phe Asn Arg Ala Ser Ala 200 His Gly Lys Pro Ser Gly Ile Leu Ala Pro Val Glu Ala Asp Ala Arg 215 220 Arg Tyr Leu Glu Trp Gly Ala Thr Phe Val Ala Val Gly Ser Asp Leu 230 235 Gly Val Phe Arg Ser Ala Thr Gln Lys Leu Ala Asp Thr Phe Lys Lys

<210> 258 <211> 444 <212> PRT <213> E. Coli

<400> 258

Met Ile Leu Asp Thr Val Asp Glu Lys Lys Gly Val His Thr Arg Tyr Leu Ile Leu Leu Ile Ile Phe Ile Val Thr Ala Val Asn Tyr Ala Asp Arg Ala Thr Leu Ser Ile Ala Gly Thr Glu Val Ala Lys Glu Leu 40 Gln Leu Ser Ala Val Ser Met Gly Tyr Ile Phe Ser Ala Phe Gly Trp 55 Ala Tyr Leu Leu Met Gln Ile Pro Gly Gly Trp Leu Leu Asp Lys Phe 70 75 Gly Ser Lys Lys Val Tyr Thr Tyr Ser Leu Phe Phe Trp Ser Leu Phe 85 90 Thr Phe Leu Gln Gly Phe Val Asp Met Phe Pro Leu Ala Trp Ala Gly 100 105 Ile Ser Met Phe Phe Met Arg Phe Met Leu Gly Phe Ser Glu Ala Pro 120 Ser Phe Pro Ala Asn Ala Arg Ile Val Ala Ala Trp Phe Pro Thr Lys 135 140 Glu Arg Gly Thr Ala Ser Ala Ile Phe Asn Ser Ala Gln Tyr Phe Ser 150 155 Leu Ala Leu Phe Ser Pro Leu Leu Gly Trp Leu Thr Phe Ala Trp Gly 165 170 Trp Glu His Val Phe Thr Val Met Gly Val Ile Gly Phe Val Leu Thr 180 185 Ala Leu Trp Ile Lys Leu Ile His Asn Pro Thr Asp His Pro Arg Met 195 200 Ser Ala Glu Glu Leu Lys Phe Ile Ser Glu Asn Gly Ala Val Val Asp 215 220 Met Asp His Lys Lys Pro Gly Ser Ala Ala Ala Ser Gly Pro Lys Leu 230 235 His Tyr Ile Lys Gln Leu Leu Ser Asn Arg Met Met Leu Gly Val Phe 245 250

												_			
Phe	Gly	Gln	Tyr 260	Phe	Ile	Asn	Thr	Ile 265	Thr	Trp	Phe	Phe	Leu 270	Thr	Trp
Phe	Pro	Ile 275	Tyr	Leu	Val	Gln	Glu 280	Lys	Gly	Met	Ser	Ile 285	Leu	Lys	Val.
Gly	Leu 290	Val	Ala	Ser	Ile	Pro 295	Ala	Leu	Cys	Gly	Phe 300	Ala	Gly	Gly	Val
Leu 305	Gly	Gly	Val	Phe	Ser 310	Asp	Tyr	Leu	Ile	Lys 315	Arg	Gly	Leu	Ser	Leu 320
	٠			325	Leu-				330					335	
Thr	Ile	Ile	Leu 340	Cys	Asn	Tyr	Thr	Asn 345	Asn	Thr	Thr	Leu	Val 350	Val	Met
Leu	Met	Ala 355	Leu	Ala	Phe		360	Lys .	Gly	Phe	Gly	Ala 365	Leu	Gly	Trp
Pro	Val 370	Ile	Ser	Asp	Thr	Ala 375	Pro	Lys	Glu	Ile	Val 380	Gly	Leu	Cys	Gly
Gly 385	Val	Phe	Asn	Val	Phe 390	Gly	Asn	Val	Ala	Ser 395	Ile	Val	Thr	Pro	Leu 400
Val	Ile	Gly	Tyr	Leu 405	Val	Ser	Glu	Leu	His 410	Ser	Phe	Asn	Ala	Ala 415	
Val	Phe	Val	Gly 420	Cys	Ser	Ala	Leu	Met 425	Ala	Met	Val	Cys	Tyr 430	Leu	Phe
Val	Val.	Gly 435	Asp	Ile	Lys	Arg	Met 440	Glu	Leu	Gln	Lys			٠.	

<210> 259 <211> 511 <212> PRT <213> E. Coli

<400> 259

Met Gln Thr Ser Asp Thr Arg Ala Leu Pro Leu Cys Ala Arg Ser 10 Val Tyr Lys Gln Tyr Ser Gly Val Asn Val Leu Lys Gly Ile Asp Phe 20 25 Thr Leu His Gln Gly Glu Val His Ala Leu Leu Gly Gly Asn Gly Ala 40 Gly Lys Ser Thr Leu Met Lys Ile Ile Ala Gly Ile Thr Pro Ala Asp 55 Ser Gly Thr Leu Glu Ile Glu Gly Asn Asn Tyr Val Arg Leu Thr Pro 70 75 Val His Ala His Gln Leu Gly Ile Tyr Leu Val Pro Gln Glu Pro Leu 90 85 Leu Phe Pro Ser Leu Ser Ile Lys Glu Asn Ile Leu Phe Gly Leu Ala 100 105 Lys Lys Gln Leu Ser Met Gln Lys Met Lys Asn Leu Leu Ala Ala Leu 120 Gly Cys Gln Phe Asp Leu His Ser Leu Ala Gly Ser Leu Asp Val Ala 135 · 140 Asp Arg Gln Met Val Glu Ile Leu Arg Gly Leu Met Arg Asp Ser Arg 150 155 Ile Leu Ile Leu Asp Glu Pro Thr Ala Ser Leu Thr Pro Ala Glu Thr 165 170 Glu Arg Leu Phe Ser Arg Leu Gln Glu Leu Leu Ala Thr Gly Val Gly 185 Ile Val Phe Ile Ser His Lys Leu Pro Glu Ile Arg Gln Ile Ala Asp 200 205 Arg Ile Ser Val Met Arg Asp Gly Thr Ile Ala Leu Ser Gly Lys Thr 215 220 Ser Glu Leu Ser Thr Asp Asp Ile Ile Gln Ala Ile Thr Pro Ala Val

225					230					235		•			240
			Ser	245					250					255	Pro
			Pro 260					265					270		
		275	Gly				280					285			
•	290		Leu			295					300			-	
305			Thr		310		•			315					320
			Gly	325					330			_		335	
			Leu 340					345					350	_	
		355	Ala				360					365			
	370		Phe			375					380				
385			Arg		390	•				395		•			400
			Leu	405					410					415	-
			Ala 420		•			425			_		430		
		435	Vaļ				440					445		_	
	450		Gln			455					460				
Glu 465			Leu		470	•				475		•		-	480
			Ser	485					490					495	Iļe
Met	Arg	Val	Ala 500	Phe	Gly	Asp	Ser	Gln 505	Arg	Gln	Glụ	Ala	Ser 510	Cys	

<210> 260

<211> 342 <212> PRT

<213> E. Coli

<400> 260

Met Leu Lys Phe Ile Gln Asn Asn Arg Glu Ile Thr Ala Leu Leu Ala 10 Val Val Leu Phe Val Leu Pro Gly Phe Leu Asp Arg Gln Tyr Leu 25 Ser Val Gln Thr Leu Thr Met Val Tyr Ser Ser Ala Gln Ile Leu Ile 40 Leu Leu Ala Met Gly Ala Thr Leu Val Met Leu Thr Arg Asn Ile Asp 55 60 Val Ser Val Gly Ser Ile Thr Gly Met Cys Ala Val Leu Leu Gly Met 70 75 Leu Leu Asn Ala Gly Tyr Ser Leu Pro Val Ala Cys Val Ala Thr Leu 85 90 Leu Leu Gly Leu Leu Ala Gly Phe Phe Asn Gly Val Leu Val Ala Trp 100 105. Leu Lys Ile Pro Ala Ile Val Ala Thr Leu Gly Thr Leu Gly Leu Tyr 120 125 Arg Gly Ile Met Leu Leu Trp Thr Gly Gly Lys Trp Ile Glu Gly Leu 135

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Pro Ala Glu Leu Lys Gln Leu Ser Ala Pro Leu Leu Gly Val Ser
                    150
                                        155
Ala Ile Gly Trp Leu Thr Ile Ile Leu Val Ala Phe Met Ala Trp Leu
                                    170
                                                      175
Leu Ala Lys Thr Ala Phe Gly Arg Ser Phe Tyr Ala Thr Gly Asp Asn
                                185
Leu Gln Gly Ala Arg Gln Leu Gly Val Arg Thr Glu Ala Ile Arg Ile
                            200
Val Ala Phe Ser Leu Asn Gly Cys Met Ala Ala Leu Ala Gly Ile Val
                        215
                                             220
Phe Ala Ser Gln Ile Gly Phe Ile Pro Asn Gln Thr Gly Thr Gly Leu
                    230
                                        235
Glu Met Lys Ala Ile Ala Ala Cys Val Leu Gly Gly Ile Ser Leu Leu
                245
                                    250
Gly Gly Ser Gly Ala Ile Ile Gly Ala Val Leu Gly Ala Trp Phe Leu
                                265
            260
Thr Gln Ile Asp Ser Val Leu Val Leu Leu Arg Ile Pro Ala Trp Trp
                            280
Asn Asp Phe Ile Ala Gly Leu Val Leu Leu Ala Val Leu Val Phe Asp
                        295
Gly Arg Leu Arg Cys Ala Leu Glu Arg Asn Leu Arg Arg Gln Lys Tyr
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                    310
Ala Arg Phe Met Thr Pro Pro Pro Ser Val Lys Pro Ala Ser Ser Gly
                325
                                    330
Lys Lys Arg Glu Ala Ala
            340
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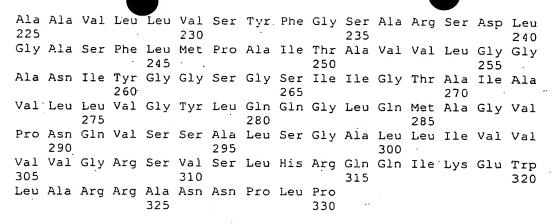
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<212> PRT

<213> E. Coli

<400> 261

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<210> 262 <211> 340 <212> PRT <213> E. Coli

<400> 262 Met Thr Leu His Arg Phe Lys Lys Ile Ala Leu Leu Ser Ala Leu Gly 5 Ile Ala Ala Ile Ser Met Asn Val Gln Ala Ala Glu Arg Ile Ala Phe 25 Ile Pro Lys Leu Val Gly Val Gly Phe Phe Thr Ser Gly Gly Asn Gly 35 40 Ala Gln Gln Ala Gly Lys Glu Leu Gly Val Asp Val Thr Tyr Asp Gly 50 \$55 \$60 Pro Thr Glu Pro Ser Val Ser Gly Gln Val Gln Leu Ile Asn Asn Phe 75 Val Asn Gln Gly Tyr Asn Ala Ile Ile Val Ser Ala Val Ser Pro Asp 90 Gly Leu Cys Pro Ala Leu Lys Arg Ala Met Gln Arg Gly Val Arg Val 105 Leu Thr Trp Asp Ser Asp Thr Lys Pro Glu Cys Arg Ser Tyr Tyr Ile 120 125 Asn Gln Gly Thr Pro Ala Gln Leu Gly Gly Met Leu Val Asp Met Ala 135 140 Ala Arg Gln Val Asn Lys Asp Lys Ala Lys Val Ala Phe Phe Tyr Ser 150 155 Ser Pro Thr Val Thr Asp Gln Asn Gln Trp Val Lys Glu Ala Lys Ala 165 170 175 Lys Ile Ala Lys Glu His Pro Gly Trp Glu Ile Val Thr Thr Gln Phe 180 185 190 Gly Tyr Asn Asp Ala Thr Lys Ser Leu Gln Thr Ala Glu Gly Ile Leu 200 205 Lys Ala Tyr Ser Asp Leu Asp Ala Ile Ile Ala Pro Asp Ala Asn Ala 215 220 Leu Pro Ala Ala Ala Gln Ala Ala Glu Asn Leu Lys Asn Asp Lys Val 230 235 Ala Ile Val Gly Phe Ser Thr Pro Asn Val Met Arg Pro Tyr Val Glu 245 250 Arg Gly Thr Val Lys Glu Phe Gly Leu Trp Asp Val Val Gln Gly 260 265 Lys Ile Ser Val Tyr Val Ala Asp Ala Leu Leu Lys Lys Gly Ser Met 280 Lys Thr Gly Asp Lys Leu Asp Ile Lys Gly Val Gly Gln Val Glu Val

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300
Ser Pro Asn Ser Val Gln Gly Tyr Asp Tyr Glu Ala Asp Gly Asn Gly
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                                        315
Ile Val Leu Leu Pro Glu Arg Val Ile Phe Asn Lys Glu Asn Ile Gly
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                325
Lys Tyr Asp Phe
            340
      <210> 263
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      <213> E. Coli
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Met Ala Asp Leu Asp Asp Ile Lys Asp Gly Lys Asp Phe Arg Thr Asp
Gln Pro Gln Lys Asn Ile Pro Phe Thr Leu Lys Gly Cys Gly Ala Leu
Asp Trp Gly Met Gln Ser Arg Leu Ser Arg Ile Phe Asn Pro Lys Thr
Gly Lys Thr Val Met Leu Ala Phe Asp His Gly Tyr Phe Gln Gly Pro
                        55
Thr Thr Gly Leu Glu Arg Ile Asp Ile Asn Ile Ala Pro Leu Phe Glu
                                        75
His Ala Asp Val Leu Met Cys Thr Arg Gly Ile Leu Arg Ser Val Val
                8.5
Pro Pro Ala Thr Asn Arg Pro Val Val Leu Arg Ala Ser Gly Ala Asn
                               105
           100
Ser Ile Leu Ala Glu Leu Ser Asn Glu Ala Val Ala Leu Ser Met Asp
                           120
       115
Asp Ala Val Arg Leu Asn Ser Cys Ala Val Ala Ala Gln Val Tyr Ile
                       135
                                           140
Gly Ser Glu Tyr Glu His Gln Ser Ile Lys Asn Ile Ile Gln Leu Val
                 . 150
                                       155
Asp Ala Gly Met Lys Val Gly Met Pro Thr Met Ala Val Thr Gly Val
                                    170
                165
Gly Lys Asp Met Val Arg Asp Gln Arg Tyr Phe Ser Leu Ala Thr Arg
           180
                               185
Ile Ala Ala Glu Met Gly Ala Gln Ile Ile Lys Thr Tyr Tyr Val Glu
       195
                            200
Lys Gly Phe Glu Arg Ile Val Ala Gly Cys Pro Val Pro Ile Val Ile
           . 215
                                            220
Ala Gly Gly Lys Lys Leu Pro Glu Arg Glu Ala Leu Glu Met Cys Trp
                   230
                                       235
Gln Ala Ile Asp Gln Gly Ala Ser Gly Val Asp Met Gly Arg Asn Ile
               245
                                    250
Phe Gln Ser Asp His Pro Val Ala Met Met Lys Ala Val Gln Ala Val
                               265
           260
Val His His Asn Glu Thr Ala Asp Arg Ala Tyr Glu Leu Tyr Leu Ser
       275
                          .280
Glu Lys Glń
    290
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      <213> E. Coli
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-101-

<400> 264

 Met
 His
 Val
 Thr
 Leu
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 Asn
 Val
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 Asp

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 Glu
 Glu

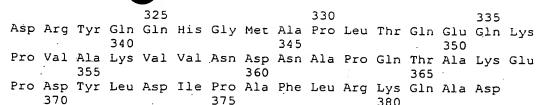
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 Ser

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 Tyr
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 Ala
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 Lys
 Asp
 Glu
 Asp
 Pro
 Ala
 Phe

 Bo
 Ileu
 Tyr
 Ileu
 Ileu</t

<210> 265 <211> 383 , <212> PRT <213> E. Coli

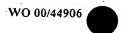
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<210> 266 <211> 1014 <212> PRT <213> E. Coli

<400> 266 Met Asp Val Ser Arg Gln Phe Phe Lys Ile Cys Ala Gly Gly Met . Ala Gly Thr Thr Val Ala Ala Leu Gly Phe Ala Pro Lys Gln Ala Leu Ala Gln Ala Arg Asn Tyr Lys Leu Leu Arg Ala Lys Glu Ile Arg Asn 40 Thr Cys Thr Tyr Cys Ser Val Gly Cys Gly Leu Leu Met Tyr Ser Leu 55 Gly Asp Gly Ala Lys Asn Ala Arg Glu Ala Ile Tyr His Ile Glu Gly 75 Asp Pro Asp His Pro Val Ser Arg Gly Ala Leu Cys Pro Lys Gly Ala 90 Gly Leu Leu Asp Tyr Val Asn Ser Glu Asn Arg Leu Arg Tyr Pro Glu 105 Tyr Arg Ala Pro Gly Ser Asp Lys Trp Gln Arg Ile Ser Trp Glu Glu 120 Ala Phe Ser Arg Ile Ala Lys Leu Met Lys Ala Asp Arg Asp Ala Asn 135 Phe Ile Glu Lys Asn Glu Gln Gly Val Thr Val Asn Arg Trp Leu Ser 150 155 Thr Gly Met Leu Cys Ala Ser Gly Ala Ser Asn Glu Thr Gly Met Leu 165 170 Thr Gln Lys Phe Ala Arg Ser Leu Gly Met Leu Ala Val Asp Asn Gln 180 185 Ala Arg Val His Gly Pro Thr Val Ala Ser Leu Ala Pro Thr Phe Gly 200 Arg Gly Ala Met Thr Asn His Trp Val Asp Ile Lys Asn Ala Asn Val 215 Val Met Val Met Gly Gly Asn Ala Ala Glu Ala His Pro Val Gly Phe 230 235 Arg Trp Ala Met Glu Ala Lys Asn Asn Asn Asp Ala Thr Leu Ile Val 250 Val Asp Pro Arg Phe Thr Arg Thr Ala Ser Val Ala Asp Ile Tyr Ala 265 Pro Ile Arg Ser Gly Thr Asp Ile Thr Phe Leu Ser Gly Val Leu Arg 280 Tyr Leu Ile Glu Asn Asn Lys Ile Asn Ala Glu Tyr Val Lys His Tyr 295 Thr Asn Ala Ser Leu Leu Val Arg Asp Asp Phe Ala Phe Glu Asp Gly 310 315 Leu Phe Ser Gly Tyr Asp Ala Glu Lys Arg Gln Tyr Asp Lys Ser Ser 325 330 Trp Asn Tyr Gln Leu Asp Glu Asn Gly Tyr Ala Lys Arg Asp Glu Thr 345 340 Leu Thr His Pro Arg Cys Val Trp Asn Leu Leu Lys Glu His Val Ser 355

•												•			
Arg	Tyr 370	Thr	Pro	Asp	Val	Val 375	Glu	Asn	Ile	Cys	Gly 380	Thr	Pro	Lys	Ala
. Asp 385		Leu	Lys	Val	Cys 390	Glu	Val	Leu`	Ala	Ser 395		Ser	Ala	Pro	Asp 400
Arg	Thr	Thr	Thr	Phe 405	Leu	Tyr	Ala	Leu	Gly 410	Trp	Thr	Gln	His	Thr 415	Val
			420		•			425					430	Leu	_
		435					440					445		His	
	450					455					460			Leu	•
465					470	1				475		•		Ser	480
				485					490					Asn 495	-
		•	500					505					510	Phe	-
		515					520					525	_	Leu	
	530					535					540			Met	-
545					550			-		555				Val	560
		٠.		565					570					Leu 575	
			580					585					590	Phe	_
	-	595					600					605		Gln	
	610					615					620		_	Gly	
625					630					635			_	Gln	640
				645					650					Ile 655 Val	
			660					665				,	670	Pro	
		675					680					685		Asp	
	690					695					700			Leu	
705					710					715				Cys	.720
				725			•		730					735 Asn	
			740					745					750	Ala	
		755					760					765		Ala	-
•	770					775					780			Trp	
785					790					795				Ala	800
				805					810					815 Met	
			820					825					830		-
	•	835					840	•				845		Glu	
ı y I	GIU	PLO	тте	.GIU	INT	L L O	ren	GTÀ	IUL	ASD	Pro	ren	Hls	Pro	Asn



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855
                                            860
Val Val Ser Asn Pro Val Val Arg Leu Tyr Glu Gln Asp Ala Leu Arg
                   870
                                        875
Met Gly Lys Lys Glu Gln Phe Pro Tyr Val Gly Thr Thr Tyr Arg Leu
               885
                                    890
Thr Glu His Phe His Thr Trp Thr Lys His Ala Leu Leu Asn Ala Ile
           900
                                905
Ala Gln Pro Glu Gln Phe Val Glu Ile Ser Glu Thr Leu Ala Ala Ala
                            920
Lys Gly Ile Asn Asn Gly Asp Arg Val Thr Val Ser Ser Lys Arg Gly
                       935
Phe Ile Arg Ala Val Ala Val Val Thr Arg Arg Leu Lys Pro Leu Asn
                   950
                                       955
Val Asn Gly Gln Gln Val Glu Thr Val Gly Ile Pro Ile His Trp Gly
                                   970
               965
Phe Glu Gly Val Ala Arg Lys Gly Tyr Ile Ala Asn Thr Leu Thr Pro
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<210> 267 <211> 294 <212> PRT <213> E. Coli

<400> 267

Met Ala Met Glu Thr Gln Asp Ile Ile Lys Arg Ser Ala Thr Asn Ser 10 Ile Thr Pro Pro Ser Gln Val Arg Asp Tyr Lys Ala Glu Val Ala Lys 20 25. Leu Ile Asp Val Ser Thr Cys Ile Gly Cys Lys Ala Cys Gln Val Ala 40 Cys Ser Glu Trp Asn Asp Ile Arg Asp Glu Val Gly His Cys Val Gly 55 Val Tyr Asp Asn Pro Ala Asp Leu Ser Ala Lys Ser Trp Thr Val Met 70 75 Arg Phe Ser Glu Thr Glu Gln Asn Gly Lys Leu Glu Trp Leu Ile Arg 8.5 90 Lys Asp Gly Cys Met His Cys Glu Asp Pro Gly Cys Leu Lys Ala Cys 100 105 Pro Ser Ala Gly Ala Ile Ile Gln Tyr Ala Asn Gly Ile Val Asp Phe 120 Gln Ser Glu Asn Cys Ile Gly Cys Gly Tyr Cys Ile Ala Gly Cys Pro 135 Phe Asn Ile Pro Arg Leu Asn Lys Glu Asp Asn Arg Val Tyr Lys Cys 150 155 Thr Leu Cys Val Asp Arg Val Ser Val Gly Gln Glu Pro Ala Cys Val 165 170 Lys Thr Cys Pro Thr Gly Ala Ile His Phe Gly Thr Lys Lys Glu Met 185 - Leu Glu Leu Ala Glu Gln Arg Val Ala Lys Leu Lys Ala Arg Gly Tyr 200 205 Glu His Ala Gly Val Tyr Asn Pro Glu Gly Val Gly Gly Thr His Val 215 220 Met Tyr Val Leu His His Ala Asp Gln Pro Glu Leu Tyr His Gly Leu 230 235 Pro Lys Asp Pro Lys Ile Asp Thr Ser Val Ser Leu Trp Lys Gly Ala 250 Leu Lys Pro Leu Ala Ala Ala Gly Phe Ile Ala Thr Phe Ala Gly Leu



260 265 270

Ile Phe His Tyr Ile Gly Ile Gly Pro Asn Lys Glu Val Asp Asp Asp 275 280 285

Glu Glu Asp His His Glu 290

<210> 268 <211> 217 <212> PRT <213> E. Coli

<400> 268 Met Ser Lys Ser Lys Met Ile Val Arg Thr Lys Phe Ile Asp Arg Ala Cys His Trp Thr Val Val Ile Cys Phe Phe Leu Val Ala Leu Ser Gly Ile Ser Phe Phe Phe Pro Thr Leu Gln Trp Leu Thr Gln Thr Phe Gly 40 Thr Pro Gln Met Gly Arg Ile Leu His Pro Phe Phe Gly Ile Ala Ile 55 Phe Val Ala Leu Met Phe Met Phe Val Arg Phe Val His His Asn Ile 75 70 . Pro Asp Lys Lys Asp Ile Pro Trp Leu Leu Asn Ile Val Glu Val Leu 85 Lys Gly Asn Glu His Lys Val Ala Asp Val Gly Lys Tyr Asn Ala Gly 105 100 Gln Lys Met Met Phe Trp Ser Ile Met Ser Met Ile Phe Val Leu Leu 120 Val Thr Gly Val Ile Ile Trp Arg Pro Tyr Phe Ala Gln Tyr Phe Pro 135 140 Met Gln Val Val Arg Tyr Ser Leu Leu Ile His Ala Ala Ala Gly Ile 150 155 Ile Leu Ile His Ala Ile Leu Ile His Met Tyr Met Ala Phe Trp Val 165 170 Lys Gly Ser Ile Lys Gly Met Ile Glu Gly Lys Val Ser Arg Arg Trp 190 185 180 Ala Lys Lys His His Pro Arg Trp Tyr Arg Glu Ile Glu Lys Ala Glu 200 Ala Lys Lys Glu Ser Glu Glu Gly Ile

<210> 269 <211> 86 <212> PRT <213> E. Coli

210

 Ala His Val
 Lys Val
 Cys Ile Asn Cys Asp Met Cys Glu 15

 Ala His Ala Asp Lys Cys Ile Asn Cys Asp Met Cys Glu 10
 Cys Glu 15

 Bro Glu Cys Pro Asn Glu Ala Ile Ser Met Gly Asp His Ile Tyr Glu 25
 Ser Met Gly Asp His Tyr Glu 30

 Bro Glu Cys Cys Thr Glu Cys Val Gly His Tyr Glu Thr Pro 35
 Ser Asp Lys Cys Thr Glu Cys Val Gly His Tyr Glu Thr Pro 45

 Bro Cys Gln Cys Cys Thr Glu Glu Cys Val Cys Tyr Asp Cys Tyr Glu Thr Pro 55
 Ser Asp Cys Cys Tyr Asp Cys Pro 55

 Bro Cys Gln Cys Cys Tyr Glu Glu Glu Glu Glu Glu Cys Cys Cys Tyr Asp Cys Phe Val Leu Met 70
 Ser Asp Cys Cys Cys Tyr Asp Cys Phe Val Leu Met 75

<210> 270 <211> 400 <212> PRT <213> E. Coli

<400> 270 Met Gln Ser Val Asp Val Ala Ile Val Gly Gly Met Val Gly Leu Ala Val Ala Cys Gly Leu Gln Gly Ser Gly Leu Arg Val Ala Val Leu 25 Glu Gln Arg Val Gln Glu Pro Leu Ala Ala Asn Ala Pro Pro Gln Leu 40 Arg Val Ser Ala Ile Asn Ala Ala Ser Glu Lys Leu Leu Thr Arg Leu Gly Val Trp Gln Asp Ile Leu Ser Arg Arg Ala Ser Cys Tyr His Gly Met Glu Val Trp Asp Lys Asp Ser Phe Gly His Ile Ser Phe Asp Asp 90 Gln Ser Met Gly Tyr Ser His Leu Gly His Ile Val Glu Asn Ser Val 100 105 Ile His Tyr Ala Leu Trp Asn Lys Ala His Gln Ser Ser Asp Ile Thr 120 Leu Leu Ala Pro Ala Glu Leu Gln Gln Val Ala Trp Gly Glu Asn Glu 135 140 Thr Phe Leu Thr Leu Lys Asp Gly Ser Met Leu Thr Ala Arg Leu Val 150 155 Ile Gly Ala Asp Gly Ala Asn Ser Trp Leu Arg Asn Lys Ala Asp Ile 165 170 Pro Leu Thr Phe Trp Asp Tyr Gln His His Ala Leu Val Ala Thr Ile 180 185 Arg Thr Glu Glu Pro His Asp Ala Val Ala Arg Gln Val Phe His Gly 200 205 Glu Gly Ile Leu Ala Phe Leu Pro Leu Ser Asp Pro His Leu Cys Ser 215 220 Ile Val Trp Ser Leu Ser Pro Glu Glu Ala Gln Arg Met Gln Gln Ala 230 235 Ser Glu Asp Glu Phe Asn Arg Ala Leu Asn Ile Ala Phe Asp Asn Arg 245 250 Leu Gly Leu Cys Lys Val Glu Ser Ala Arg Gln Val Phe Pro Leu Thr 265 270 Gly Arg Tyr Ala Arg Gln Phe Ala Ser His Arg Leu Ala Leu Val Gly 280 285 Asp Ala Ala His Thr Ile His Pro Leu Ala Gly Gln Gly Val Asn Leu 295 300 Gly Phe Met Asp Ala Ala Glu Leu Ile Ala Glu Leu Lys Arg Leu His 310 315 Arg Gln Gly Lys Asp Ile Gly Gln Tyr Ile Tyr Leu Arg Arg Tyr Glu 325 330 Arg Ser Arg Lys His Ser Ala Ala Leu Met Leu Ala Gly Met Gln Gly 340 . 345 Phe Arg Asp Leu Phe Ser Gly Thr Asn Pro Ala Lys Lys Leu Leu Arg 360 .365 Asp Ile Gly Leu Lys Leu Ala Asp Thr Leu Pro Gly Val Lys Pro Gln 375 380 Leu Ile Arg Gln Ala Met Gly Leu Asn Asp Leu Pro Glu Trp Leu Arg 385 390 395

<210> 271

<211> 392 <212> PRT <213> E. Coli

<400> 271 Met Ser Val Ile Ile Val Gly Gly Gly Met Ala Gly Ala Thr Leu Ala 10 Leu Ala Ile Ser Arg Leu Ser His Gly Ala Leu Pro Val His Leu Ile Glu Ala Thr Ala Pro Glu Ser His Ala His Pro Gly Phe Asp Gly Arg Ala Ile Ala Leu Ala Ala Gly Thr Cys Gln Gln Leu Ala Arg Ile Gly Val Trp Gln Ser Leu Ala Asp Cys Ala Thr Ala Ile Thr Thr Val His 70 Val Ser Asp Arg Gly His Ala Gly Phe Val Thr Leu Ala Ala Glu Asp 85 90 Tyr Gln Leu Ala Ala Leu Gly Gln Val Val Glu Leu His Asn Val Gly 100 105 Gln Arg Leu Phe Ala Leu Leu Arg Lys Ala Pro Gly Val Thr Leu His 120 Cys Pro Asp Arg Val Ala Asn Val Ala Arg Thr Gln Ser His Val Glu 135 Val Thr Leu Glu Ser Gly Glu Thr Leu Thr Gly Arg Val Leu Val Ala 150 155 Ala Asp Gly Thr His Ser Ala Leu Ala Thr Ala Cys Gly Val Asp Trp 165 -170 Gln Gln Glu Pro Tyr Glu Gln Leu Ala Val Ile Ala Asn Val Ala Thr 180 185 190 Ser Val Ala His Glu Gly Arg Ala Phe Glu Arg Phe Thr Gln His Gly 200 Pro Leu Ala Met Leu Pro Met Ser Asp Gly Arg Cys Ser Leu Val Trp 215 220 Cys His Pro Leu Glu Arg Arg Glu Glu Val Leu Ser Trp Ser Asp Glu 230 235 Lys Phe Cys Arg Glu Leu Gln Ser Ala Phe Gly Trp Arg Leu Gly Lys 245 Ile Thr His Ala Gly Lys Arg Ser Ala Tyr Pro Leu Ala Leu Thr His 260 265 Ala Ala Arg Ser Ile Thr His Arg Thr Val Leu Val Gly Asn Ala Ala 275 280 Gln Thr Leu His Pro Ile Ala Gly Gln Gly Phe Asn Leu Gly Met Arg 295 300 Asp Val Met Ser Leu Ala Glu Thr Leu Thr Gln Ala Gln Glu Arg Gly 310 315 Glu Asp Met Gly Asp Tyr Gly Val Leu Cys Arg Tyr Gln Gln Arg Arg 325 330 Gln Ser Asp Arg Glu Ala Thr Ile Gly Val Thr Asp Ser Leu Val His 340 345 Leu Phe Ala Asn Arg Trp Ala Pro Leu Val Val Gly Arg Asn Ile Gly 360. 365 Leu Met Thr Met Glu Leu Phe Thr Pro Ala Arg Asp Val Leu Ala Gln 375 Arg Thr Leu Gly Trp Val Ala Arg 385 390

<210> 272 <211> 441 <212> PRT <213> E. Coli

	<	400>	272												
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			Met 20					25					30		
Glu	Val	Thr 35	Arg	Ser	Ala	Asp	Ser 40	Glu	Tyr	Pro	Tyr	Arg 45	Gln	Asn	Ser
Asp	Phe 50	Trp	Tyr	Phe	Thr	Gly 55	Phe	Asn	Glu	Pro	Glu 60	Ala	Val	Leu	Val
Leu 65	Ile	Lys	Ser	Asp	Asp 70	Thr	His	Asn	His	Ser 75	Val	Leu	Phe	Asn	Arg 80
	-	•	Leu	85				_	90	_	_	_		95	
			Pro 100					105					110		
Glu	Ile	Asn 115	Gln	Gln	Leu	Tyr	Gln 120	Leu	Leu	Asn	Gly	Leu 125	Asp	Val	Val
	130		Gln			i35					140				
145			Lys		150				•	155					160
			Ile	165	•				170					175	
			Glu 180					185				_	190		
•		195	His				200					205			
٠.	210		Leu			215			•		220				
225			Pro	.,,	230					235		-			240
			His	245					250					255	_
			11e 260					265					270		
-		275	Thr				280					285			_
	290		Asp			295					300	•			
305			Gly		310					315		•			320
			Ser	325					330					335	
			340					345			٠	•	350		Gly
		355	Trp				360					365			_
	370		Ser			375		•			380				
385			Tyr		390					3.95					400
				405					410					415	Gly
			Leu 420					425	гуs	гуѕ	Pro	GIU	430	TTE	GIU
ATA	ren	435	Val	HIS	Ala	Arg	140	GID							

<210> 273

<211> 194 <212> PRT <213> E. Coli

<400> 273 Met Leu Met Ser Ile Gln Asn Glu Met Pro Gly Tyr Asn Glu Met Asn Gln Tyr Leu Asn Gln Gln Gly Thr Gly Leu Thr Pro Ala Glu Met His Gly Leu Ile Ser Gly Met Ile Cys Gly Gly Asn Asp Asp Ser Ser Trp Leu Pro Leu Leu His Asp Leu Thr Asn Glu Gly Met Ala Phe Gly His Glu Leu Ala Gln Ala Leu Arg Lys Met His Ser Ala Thr Ser Asp Ala Leu Gln Asp Asp Gly Phe Leu Phe Gln Leu Tyr Leu Pro Asp Gly Asp Asp Val Ser Val Phe Asp Arg Ala Asp Ala Leu Ala Gly Trp Val Asn 105 His Phe Leu Leu Gly Leu Gly Val Thr Gln Pro Lys Leu Asp Lys Val 120 Thr Gly Glu Thr Gly Glu Ala Ile Asp Asp Leu Arg Asn Ile Ala Gln 135 140 Leu Gly Tyr Asp Glu Asp Glu Asp Gln Glu Glu Leu Glu Met Ser Leu 150 155 Glu Glu Ile Ile Glu Tyr Val Arg Val Ala Ala Leu Leu Cys His Asp 165 170 Thr Phe Thr His Pro Gln Pro Thr Ala Pro Glu Val Gln Lys Pro Thr 180 185

<210> 274 <211> 120 <212> PRT <213> E. Coli

Leu His

<400> 274 Met Leu Lys Leu Phe Ala Lys Tyr Thr Ser Ile Gly Val Leu Asn Thr Leu Ile His Trp Val Val Phe Gly Val Cys Ile Tyr Val Ala His Thr 20 Asn Gln Ala Leu Ala Asn Phe Ala Gly Phe Val Val Ala Val Ser Phe Ser Phe Phe Ala Asn Ala Lys Phe Thr Phe Lys Ala Ser Thr Thr Met Arg Tyr Met Leu Tyr Val Gly Phe Met Gly Thr Leu Ser Ala Thr 70 Val Gly Trp Ala Ala Asp Arg Cys Ala Leu Pro Pro Met Ile Thr Leu 90 Val Thr Phe Ser Ala Ile Ser Leu Val Cys Gly Phe Val Tyr Ser Lys 100 105 Phe Ile Val Phe Arg Asp Ala Lys 115 . 120

> <210> 275 <211> 306 <212> PRT <213> E. Coli

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Met Lys Ile Ser Leu Val Val Pro Val Phe Asn Glu Glu Glu Ala Ile
Pro Ile Phe Tyr Lys Thr Val Arg Glu Phe Glu Glu Leu Lys Ser Tyr
Glu Val Glu Ile Val Phe Ile Asn Asp Gly Ser Lys Asp Ala Thr Glu
Ser Ile Ile Asn Ala Leu Ala Val Ser Asp Pro Leu Val Val Pro Leu
Ser Phe Thr Arg Asn Phe Gly Lys Glu Pro Ala Leu Phe Ala Gly Leu
Asp His Ala Thr Gly Asp Ala Ile Ile Pro Ile Asp Val Asp Leu Gln
                                     90
Asp Pro Ile Glu Val Ile Pro His Leu Ile Glu Lys Trp Gln Ala Gly
                                 105
Ala Asp Met Val Leu Ala Lys Arg Ser Asp Arg Ser Thr Asp Gly Arg
                            120
Leu Lys Arg Lys Thr Ala Glu Trp Phe Tyr Lys Leu His Asn Lys Ile
                        135
Ser Asn Pro Lys Ile Glu Glu Asn Val Gly Asp Phe Arg Leu Met Ser
                    150
                                         155
Arg Asp Val Val Glu Asn Ile Lys Leu Met Pro Glu Arg Asn Leu Phe
                165
                                    170
Met Lys Gly Ile Leu Ser Trp Val Gly Gly Lys Thr Asp Ile Val Glu
           180
                                185
Tyr Val Arg Ala Glu Arg Ile Ala Gly Asp Thr Lys Phe Asn Gly Trp
                            200
Lys Leu Trp Asn Leu Ala Leu Glu Gly Ile Thr Ser Phe Ser Thr Phe
                        215
                                            220
Pro Leu Arg Ile Trp Thr Tyr Ile Gly Leu Val Val Ala Ser Val Ala
                    230
                                        235
Phe Ile Tyr Gly Ala Trp Met Ile Leu Asp Thr Ile Ile Phe Gly Asn
                245
Ala Val Arg Gly Tyr Pro Ser Leu Leu Val Ser Ile Leu Phe Leu Gly
           260
                                265
Gly Ile Gln Met Ile Gly Ile Gly Val Leu Gly Glu Tyr Ile Gly Arg
                            280
Thr Tyr Ile Glu Thr Lys Lys Arg Pro Lys Tyr Ile Ile Lys Arg Val
  290
                        295
Lys Lys
305
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<210> 276 <211> 443 <212> PRT <213> E. Coli

 <400>
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 5
 10
 15

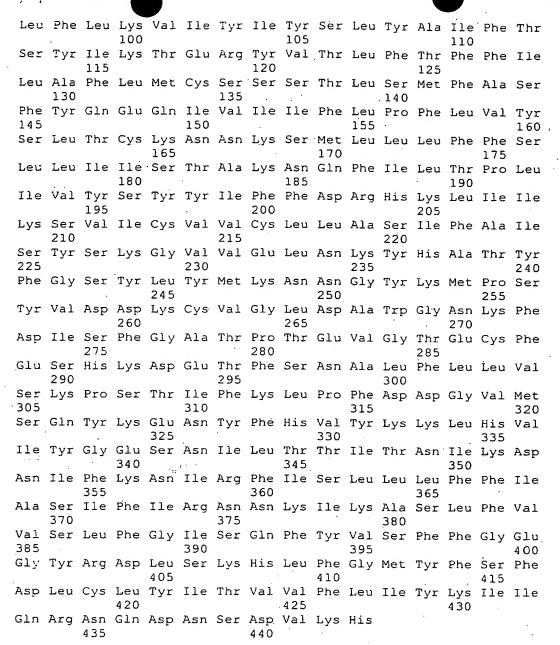
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 25
 30

 Gly Arg Ala Ile Lys Pro Leu Ile Glu Asp Ile Pro Ala Phe Thr Tyr 35
 40
 45

 Asp Leu Pro Leu Leu Tyr Lys Leu Lys Gly His Ile Asp Ser Ile Asp 50
 55
 60

 Ser Tyr Glu Tyr Ile Ser Ser Tyr Ser Tyr Ile Leu Tyr Thr Tyr Val 65
 70
 75
 80

 Leu Phe Ile Ser Ile Phe Thr Glu Tyr Leu Asp Ala Arg Val Leu Ser 90
 90
 95



<210> 277 <211> 82 <212> PRT <213> E. Coli



Val Ala Val Ile Gly Ala Ile Val Val Leu Phe Ile Tyr Arg Lys Ile Lys Ser

<210> 278 <211> 60 <212> PRT <213> E: Coli

<400> 278 Met Gly Lys Ala Thr Tyr Thr Val Thr Val Thr Asn Asn Ser Asn Gly 10 Val Ser Val Asp Tyr Glu Thr Glu Thr Pro Met Thr Leu Leu Val Pro 25 Glu Val Ala Ala Glu Val Ile Lys Asp Leu Val Asn Thr Val Arg Ser . 40 Tyr Asp Thr Glu Asn Glu His Asp Val Cys Gly Trp 55

<210> 279 <211> 119 <212> PRT <213> E. Coli

<400> 279

Met Leu Gln Ile Pro Gln Asn Tyr Ile His Thr Arg Ser Thr Pro Phe Trp Asn Lys Gln Thr Ala Pro Ala Gly Ile Phe Glu Arg His Leu Asp Lys Gly Thr Arg Pro Gly Val Tyr Pro Arg Leu Ser Val Met His Gly Ala Val Lys Tyr Leu Gly Tyr Ala Asp Glu His Ser Ala Glu Pro Asp Gln Val Ile Leu Ile Glu Ala Gly Gln Phe Ala Val Phe Pro Pro Glu 75 Lys Trp His Asn Ile Glu Ala Met Thr Asp Asp Thr Tyr Phe Asn Ile Asp Phe Phe Val Ala Pro Glu Val Leu Met Glu Gly Ala Gln Gln Arg 100 105 Lys Val Ile His Asn Gly Lys

<210> 280 <211> 246 <212> PRT <213> E. Coli

<400> 280

115

Met Lys Phe Lys Val Ile Ala Leu Ala Ala Leu Met Gly Ile Ser Gly 10 Met Ala Ala Gln Ala Asn Glu Leu Pro Asp Gly Pro His Ile Val Thr 25 Ser Gly Thr Ala Ser Val Asp Ala Val Pro Asp Ile Ala Thr Leu Ala 40 Ile Glu Val Asn Val Ala Ala Lys Asp Ala Ala Thr Ala Lys Lys Gln 55 Ala Asp Glu Arg Val Ala Gln Tyr Ile Ser Phe Leu Glu Leu Asn Gln

65					70					75					80
Ile	Ala	Lys	Lys	Asp 85	Ile	Ser	Ser	Ala	Asn 90	Leu	Arg	Thr	Gln	Pro 95	Asp
Tyr	Asp	Tyr	Gln 100	Asp	Gly.	Lys	Ser	Ile 105	Leu	Lys	Gly	Tyr	Arg 110	Ala	Val
Arg	Thr	Val. 115	Glu	Val	Thr	Leu	Arg 120	Gln	Leu	Asp	Lys	Leu 125	Asn	Ser	Leu
Leu	Asp 130	Gly	Ala	Leu	Lys	Ala 135	Gly	Leu	Asn	Glu	Ile 140	Arg	Ser	Val	Ser
Leu 145	Gly	Val	Ala	Gln	Pro 150	Asp	Ala	Tyr	Lys	Asp 155	Lys	Ala	Arg	Lys	Ala 160
Ala	Ile	Asp	Asn	Ala 165	Ile	His	Gln	Ala	Gln 170	Glu	Leu	Ala	Asn	Gly 175	Phe
His	Arg	Lys	Leu 180	Gly	Pro	Val	Tyr	Ser 185	Val	Arg	Tyr	His	Val 190	Ser	Asn
Tyr	Gln	Pro 195	Ser	Pro	Met	Val	Arg 200	Met	Met	Lys	Ala	Asp 205	Ala	Ala	Pro
Val	Ser 210		Gln	Glu	Thr	Tyr 215	Glu	Gln	Ala	Ala	Ile 220	Gln	Phe	Asp	Asp
Gln 225	Val	Asp	Val	Val	Phe 230	Gln	Leu	Glu 	Pro	Val 235	Asp	Gln	Gln	Pro	Ala 240
Lys	Thr	Pro	Ala	Ala 245	Gln									• •	•

<210> 281 <211> 464 <212> PRT <213> E. Coli

<400> 281 Met Leu Leu Asp Ala Cys Ser Gln Met Cys Pro Ser Phe Arg Arg . 10 Phe Gln Thr Val Phe His Asn Ser Ser Ile Phe Leu Pro Tyr Trp Leu 20 Ala Thr Leu Val Ser Phe Arg Glu Thr Phe Gln Glu Glu Lys Leu Leu 40 Thr Met Lys Gly Ser Tyr Lys Ser Arg Trp Val Ile Val Ile Val Val 55 Val Ile Ala Ala Ile Ala Ala Phe Trp Phe Trp Gln Gly Arg Asn Asp 70 Ser Arg Ser Ala Ala Pro Gly Ala Thr Lys Gln Ala Gln Gln Ser Pro 90 Ala Gly Gly Arg Arg Gly Met Arg Ser Gly Pro Leu Ala Pro Val Gln 100 105 110 Ala Ala Thr Ala Val Glu Gln Ala Val Pro Arg Tyr Leu Thr Gly Leu 120 125 Gly Thr Ile Thr Ala Ala Asn Thr Val Thr Val Arg Ser Arg Val Asp 135 140 Gly Gln Leu Ile Ala Leu His Phe Gln Glu Gly Gln Gln Val Lys Ala 150 155 Gly Asp Leu Leu Ala Glu Ile Asp Pro Ser Gln Phe Lys Val Ala Leu 165 170 Ala Gln Ala Gln Gly Gln Leu Ala Lys Asp Lys Ala Thr Leu Ala Asn 180 185 190 Ala Arg Arg Asp Leu Ala Arg Tyr Gln Gln Leu Ala Lys Thr Asn Leu 200 205 Val Ser Arg Gln Glu Leu Asp Ala Gln Gln Ala Leu Val Ser Glu Thr 215 220 Glu Gly Thr Ile Lys Ala Asp Glu Ala Ser Val Ala Ser Ala Gln Leu 230 235



Gln Leu Asp Trp Ser Arg Ile Thr Ala Pro Val Asp Gly Arg Val Gly 245 · 250 Leu Lys Gln Val Asp Val Gly Asn Gln Ile Ser Ser Gly Asp Thr Thr 260 265 Gly Ile Val Val Ile Thr Gln Thr His Pro Ile Asp Leu Val Phe Thr 280 Leu Pro Glu Ser Asp Ile Ala Thr Val Val Gln Ala Gln Lys Ala Gly 295 Lys Pro Leu Val Val Glu Ala Trp Asp Arg Thr Asn Ser Lys Lys Leu 310 315 Ser Glu Gly Thr Leu Leu Ser Leu Asp Asn Gln Ile Asp Ala Thr Thr 330 Gly Thr Ile Lys Val Lys Ala Arg Phe Asn Asn Gln Asp Asp Ala Leu 345 Phe Pro Asn Gln Phe Val Asn Ala Arg Met Leu Val Asp Thr Glu Gln 360 Asn Ala Val Val Ile Pro Thr Ala Ala Leu Gln Met Gly Asn Glu Gly . 375 380 His Phe Val Trp Val Leu Asn Ser Glu Asn Lys Val Ser Lys His Leu 390 395 Val Thr Pro Gly Ile Gln Asp Ser Gln Lys Val Val Ile Arg Ala Gly 405 410 Ile Ser Ala Gly Asp Arg Val Val Thr Asp Gly Ile Asp Arg Leu Thr 420 425 Glu Gly Ala Lys Val Glu Val Glu Ala Gln Ser Ala Thr Thr Pro 435 440 Glu Glu Lys Ala Thr Ser Arg Glu Tyr Ala Lys Lys Gly Ala Arg Ser 455

<210> 282 <211> 1040 <212> PRT <213> E. Coli

<400> 282 Met Gln Val Leu Pro Pro Ser Ser Thr Gly Gly Pro Ser Arg Leu Phe 10 Ile Met Arg Pro Val Ala Thr Thr Leu Leu Met Val Ala Ile Leu Leu 25 20 Ala Gly Ile Ile Gly Tyr Arg Ala Leu Pro Val Ser Ala Leu Pro Glu 40 . Val Asp Tyr Pro Thr Ile Gln Val Val Thr Leu Tyr Pro Gly Ala Ser 55 60 Pro Asp Val Met Thr Ser Ala Val Thr Ala Pro Leu Glu Arg Gln Phe 70 · 75 Gly Gln Met Ser Gly Leu Lys Gln Met Ser Ser Gln Ser Ser Gly Gly 85 90 Ala Ser Val Ile Thr Leu Gln Phe Gln Leu Thr Leu Pro Leu Asp Val 100 105 Ala Glu Gln Glu Val Gln Ala Ala Ile Asn Ala Ala Thr Asn Leu Leu 115 120 Pro Ser Asp Leu Pro Asn Pro Pro Val Tyr Ser Lys Val Asn Pro Ala 135 140 Asp Pro Pro Ile Met Thr Leu Ala Val Thr Ser Thr Ala Met Pro Met . 150 Thr Gln Val Glu Asp Met Val Glu Thr Arg Val Ala Gln Lys Ile Ser 170 Gln Ile Ser Gly Val Gly Leu Val Thr Leu Ser Gly Gly Gln Arg Pro 185 Ala Val Arg Val Lys Leu Asn Ala Gln Ala Ile Ala Ala Leu Gly Leu

•	•															
			195					200		•			205			
		Ser 210	Glu	Thr	Val	Arg	Thr 215	Ala	Ile	Thr	Gly	Ala 220	Asn	Val	Asn	Ser
			Gly	Ser	Leu	Asp 230		Pro	Ser	Arg	Ala 235	Val	Thr	Leu	Ser	Ala 240
		Asp	Gľn	Met	Gln. 245		Ala	Glu	Glu	Tyr 250		Gln	Leu	Ile	Ile 255	
	Tyr	Gln	Asn	Gly 260	Ala	Pro	Ile	Arg	Leu 265	Gly	Asp	Val	Ala	Thr 270		Glu
			275					280		Ala			285			
		290			•		295			Pro		300				·
	305					310				Pro	315					320
					325		•			Ser 330					335	
				340					345	Glu				350	:	٠.
			355					360		Leu			365			
		370					375	:		Ser		380				
	385	nec	VAI	rile	rea	390	·	261	116	Asn	395	reu	Inr	ren	мет	400
				•	405		-			Asp 410					415	
				420					425	Gly				430		
			435					440	•	Phe			445	-		
		450			-	:	455			Leu		460				•
4	465					470				Ile	475					480
					485					Leu 490 Lys					495	
				500					505	Ile		•	•	510		_
			515					520		Leu			525			
		530					535.					540				
	45					550	•			Trp	555					560
					565					Ile 570					575	
				580					585	Ala				590		
			595	•				600		Val Leu			605			
		610					615			Asp		620				
	25		200	2,5	110	630	nsp	· ·	: ·	ASP	635	Ary.	vaı	GIII	гуз	640
					645					Lys 650	Val				655	Leu
		•		660					665	Ile		•	•	670		
Τ	hr'	Gln	Tyr 675	Gln	Phe	Thr	Leu	Gln 680	Ala	Thr	Ser	Leu	Asp 685	Ala	Leu	Ser



Thr Trp 690		Pro	Gln	Leu	Met 695	Glu	Lys	Leu	Gln	Gln 700	Ľeu	Pro	Gln	Leu
Ser Asp			•	710					715				-	720
Asn Val			725					730					735	-
Val Asp		740					745					750		
Ile Tyr	755					760		•			765			
Glu Asn 770					775			•		780				
Ser Asp 785		_		790		·			795		_			800
Arg Phe			805					810					815	
Thr Ile		820					825				•	830		
Gln Ala	835		_			840					845		-	
Thr Thr 850				_	855					860				-
Ser Thr 865				870					875					880
Leu Gly			885					890					895	
Thr Leu		900		_			905					910		
Gly Ser	915					920					925			
Gly Ile 930		-			935	•				940				
Ala Glu 945	_			950				_	955			_		960
Cys Leu			965	_				970					975	•
Leu Gly		980					985		_		_	990	,	
Arg Arg	995		_		•	1000	)	_	_		1005	5 .		
Val Leu 1010	)		•		1015	5				1020	)		-	
Leu Ala 1025	Leu	Trp	Thr	Lys 1030		Arg	Phe	Ala	Arg 1035		Glu	Glu	Glu	Ala 1040

<210> 283

<211> 1025

<212> PRT

<213> E. Coli

<400> 283

 Met
 Lys
 Phe
 Ala
 Leu
 Phe
 Ile
 Tyr
 Arg
 Pro
 Val
 Ala
 Thr
 Ile
 Leu

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 Ser
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 Ala
 Ile
 Thr
 Leu
 Cys
 Gly
 Ile
 Leu
 Gly
 Phe
 Arg
 Met
 Leu

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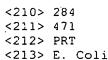
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 Ile
 Ala
 Gly
 Val
 Ser
 Glu
 Met

	65			•		70					75					80
		Ser			85					90					95	Asp
		Asp		100					105					110		
		Ala	115					120					125	_		
		Arg 130					135					140				
	145	Ser				150					155					160
		Leu		٠.	165					170					175	_
		Gly		180			•		1:85					190		
		Leu	195					200					205			
		Asn 210					215					220			_	
	225	Arg	•			230					.235					240
		Gln			245	•				250					255	
		Asp		260					265				•	270		
		Met	275					280					285	_	_	
-		Glu 290 Glu					295					300				
	305	Asp				310					315		•			320
	•	Leu			325					330					335	
		Arg		340					345					350		
		Leu	355					360					3.65			
		370 Asn					375					380				
	385	Asp				390	-				395		-			400
		Met			405					410					415	
		Thr		420					425					430		-
		Leu	435					440					445			
		450 Thr					455					460	-			
	465	Thr		•		470					475					480
		Gln			485					490					495	_
		Gly		500					505					510		
		Gly	515					·520					525		_	
		530 Ser		•			535					540				-
	545	•			-	550					555	•		•		560

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	•				U

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Met	Gly	Gly	Ile	Gln 565	Ala	Asp	Gln	Ser	Ile 570		Phe	Gln	Ala	Met 575	Arg
Gly	Lys	Leu	Gln 580	Asp	Phe	Met	Lys	Ile 585	Ile	Arg	Asp	Asp	Pro 590		Val
Asp	Asn	Val 595	Thr	Gly	Phe	Thr	Gly 600	Gly	Ser	Arg	Val	Asn 605		Gly	Met
Met	Phe 610	Ile	Thr	Leu	Lys	Pro 615	Arg	Asp	Glu	Arg	Ser 620		Thr	Ala	Gln
Gln 625	Ile	Ile	Asp	Arg	Leu 630	Arg	Val	Lys	Leu	Ala 635	Lys	Glu	Pro	Gly	Ala 640
•				645					650			_		655	Gln
	Asn		660			•		665					670		
	Arg	675					680					685			
	Leu 690					695	•				700				
705	Leu				710					715					720
	Ala			725					730					735	
	Thr		740					745					750		
	Pro	755					760			٠.		765			•
	770			•		775					780				-
785	Pro				790					795	٠.				800
	Ser	:		805					810		•			815	
	Ser		820			-		825			•	_	830		
	Val	835					840					845			
	Asn 850					855					860				_
865	Val				870	•				875				٠.	880
	Ser			885				•	890					895	
			900					905					910		Leu.
	Ile	915					920					925			
	Glu 930					935			•		940				
945	Ala				950					955					960
	Leu			965					970					975	
	Leu		980					985					990		
	Gln	995					1000	) _				1005	<b>5</b>		
	Arg 1010		Arg	Leu	Arg	Phe 1015		Arg	Lys	Pro	Lys 1020		Thr	Val	Thr
Glu 1025												•			



<400> 284 Met Thr Asp Leu Pro Asp Ser Thr Arg Trp Gln Leu Trp Ile Val Ala Phe Gly Phe Phe Met Gln Ser Leu Asp Thr Thr Ile Val Asn Thr Ala 20 . Leu Pro Ser Met Ala Gln Ser Leu Gly Glu Ser Pro Leu His Met His Met Val Ile Val Ser Tyr Val Leu Thr Val Ala Val Met Leu Pro Ala Ser Gly Trp Leu Ala Asp Lys Val Gly Val Arg Asn Ile Phe Phe Thr Ala Ile Val Leu Phe Thr Leu Gly Ser Leu Phe Cys Ala Leu Ser Gly Thr Leu Asn Glu Leu Leu Ala Arg Ala Leu Gln Gly Val Gly Gly 105 Ala Met Met Val Pro Val Gly Arg Leu Thr Val Met Lys Ile Val Pro 120 Arg Glu Gln Tyr Met Ala Ala Met Thr Phe Val Thr Leu Pro Gly Gln 135 140 Val Gly Pro Leu Gly Pro Ala Leu Gly Gly Leu Leu Val Glu Tyr 150 155 Ala Ser Trp His Trp Ile Phe Leu Ile Asn Ile Pro Val Gly Ile Ile 165 170 Gly Ala Ile Ala Thr Leu Leu Leu Met Pro Asn Tyr Thr Met Gln Thr 185 Arg Arg Phe Asp Leu Ser Gly Phe Leu Leu Leu Ala Val Gly Met Ala 195 200 Val Leu Thr Leu Ala Leu Asp Gly Ser Lys Gly Thr Gly Leu Ser Pro 215 220 Leu Thr Ile Ala Gly Leu Val Ala Val Gly Val Val Ala Leu Val Leu 230 235 Tyr Leu Leu His Ala Arg Asn Asn Arg Ala Leu Phe Ser Leu Lys 245 250 Leu Phe Arg Thr Arg Thr Phe Ser Leu Gly Leu Ala Gly Ser Phe Ala 265 Gly Arg Ile Gly Ser Gly Met Leu Pro Phe Met Thr Pro Val Phe Leu 280. Gln Ile Gly Leu Gly Phe Ser Pro Phe His Ala Gly Leu Met Met Ile . 300 295 Pro Met Val Leu Gly Ser Met Gly Met Lys Arg Ile Val Val Gln Val 310 315 Val Asn Arg Phe Gly Tyr Arg Arg Val Leu Val Ala Thr Thr Leu Gly 325 330 Leu Ser Leu Val Thr Leu Leu Phe Met Thr Thr Ala Leu Leu Gly Trp 345 Tyr Tyr Val Leu Pro Phe Val Leu Phe Leu Gln Gly Met Val Asn Ser 360 Thr Arg Phe Ser Ser Met Asn Thr Leu Thr Leu Lys Asp Leu Pro Asp . 375 380 Asn Leu Ala Ser Ser Gly Asn Ser Leu Leu Ser Met Ile Met Gln Leu 395 390 Ser Met Ser Ile Gly Val Thr Ile Ala Gly Leu Leu Gly Leu Phe 410 405 Gly Ser Gln His Val Ser Val Asp Ser Gly Thr Thr Gln Thr Val Phe . 425



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        Met
        Tyr
        Thr Trp
        Leu
        Ser
        Met
        Ala
        Leu
        Ile
        Ile
        Ala
        Leu
        Pro
        Ala
        Leu
        Ile
        Ile
        Ala
        Leu
        Pro
        Ala
        Leu
        Ile
        Ile
        Ala
        Ile
        Pro
        Ala
        Ala
        Ala
        Ile
        Ser

        Ala
        Arg
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        Arg
        Arg
        Ala
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        Ala
        Ala</
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<210> 285 <211> 344 <212> PRT <213> E. Coli

<400> 285 Met Glu Ile Arg Ile Met Leu Phe Ile Leu Met Met Val Met Pro -10 Val Ser Tyr Ala Ala Cys Tyr Ser Glu Leu Ser Val Gln His Asn Leu 20 25 Val Val Gln Gly Asp Phe Ala Leu Thr Gln Thr Gln Met Ala Thr Tyr 40 Glu His Asn Phe Asn Asp Ser Ser Cys Val Ser Thr Asn Thr Ile Thr 5.5 Pro Met Ser Pro Ser Asp Ile Ile Val Gly Leu Tyr Asn Asp Thr Ile 70 75 Lys Leu Asn Leu His Phe Glu Trp Thr Asn Lys Asn Asn Ile Thr Leu 85 90 Ser Asn Asn Gln Thr Ser Phe Thr Ser Gly Tyr Ser Val Thr Val Thr 100 105 Pro Ala Ala Ser Asn Ala Lys Val Asn Val Ser Ala Gly Gly Gly 115 120 Ser Val Met Ile Asn Gly Val Ala Thr Leu Ser Ser Ala Ser Ser Ser ... 135 Thr Arg Gly Ser Ala Ala Val Gln Phe Leu Leu Cys Leu Leu Gly Gly 150 155 Lys Ser Trp Asp. Ala Cys Val Asn Ser Tyr Arg Asn Ala Leu Ala Gln 165 170 Asn Ala Gly Val Tyr Ser Phe Asn Leu Thr Leu Ser Tyr Asn Pro Ile 180 185 Thr Thr Thr Cys Lys Pro Asp Asp Leu Leu Ile Thr Leu Asp Ser Ile 200 Pro Val Ser Gln Leu Pro Ala Thr Gly Asn Lys Ala Thr Ile Asn Ser 215 Lys Gln Gly Asp Ile Ile Leu Arg Cys Lys Asn Leu Leu Gly Gln Gln 230 235 Asn Gln Thr Ser Arg Lys Met Gln Val Tyr Leu Ser Ser Ser Asp Leu . 245 250 Leu Thr Asn Ser Asn Thr Ile Leu Lys Gly Ala Glu Asp Asn Gly Val 260 265 Gly Phe Ile Leu Glu Ser Asn Gly Ser Pro Val Thr Leu Leu Asn Ile 280 Thr Asn Ser Ser Lys Gly Tyr Thr Asn Leu Lys Glu Val Ala Ala Lys 295 300 Ser Lys Leu Thr Asp Thr Thr Val Ser Ile Pro Ile Thr Ala Ser Tyr 310 315 Tyr Val Tyr Asp Thr Asn Lys Val Lys Ser Gly Ala Leu Glu Ala Thr 325 330 Ala Leu Ile Asn Val Lys Tyr Asp

<210> 286



<211> 826 <212> PRT <213> E. Coli

<400> 286 Met Leu Arg Met Thr Pro Leu Ala Ser Ala Ile Val Ala Leu Leu Leu 10 Gly Ile Glu Ala Tyr Ala Ala Glu Glu Thr Phe Asp Thr His Phe Met 25 Ile Gly Gly Met Lys Asp Gln Gln Val Ala Asn Ile Arg Leu Asp Asp 40 Asn Gln Pro Leu Pro Gly Gln Tyr Asp Ile Asp Ile Tyr Val Asn Lys 55 -Gln Trp Arg Gly Lys Tyr Glu Ile Ile Val Lys Asp Asn Pro Gln Glu 70 Thr Cys Leu Ser Arg Glu Val Ile Lys Arg Leu Gly Ile Asn Ser Asp 85 90 Asn Phe Ala Ser Gly Lys Gln Cys Leu Thr Phe Glu Gln Leu Val Gln 100 105 Gly Gly Ser Tyr Thr Trp Asp Ile Gly Val Phe Arg Leu Asp Phe Ser 115 120 Val Pro Gln Ala Trp Val Glu Glu Leu Glu Ser Gly Tyr Val Pro Pro 135 Glu Asn Trp Glu Arg Gly Ile Asn Ala Phe Tyr Thr Ser Tyr Tyr Leu 150 155 Ser Gln Tyr Tyr Ser Asp Tyr Lys Ala Ser Gly Asn Asn Lys Ser Thr 165 170 Tyr Val Arg Phe Asn Ser Gly Leu Asn Leu Leu Gly Trp Gln Leu His 180 185 Ser Asp Ala Ser Phe Ser Lys Thr Asn Asn Asn Pro Gly Val Trp Lys 200 Ser Asn Thr Leu Tyr Leu Glu Arg Gly Phe Ala Gln Leu Leu Gly Thr 215 220. Leu Arg Val Gly Asp Met Tyr Thr Ser Ser Asp Ile Phe Asp Ser Val 230 235 Arg Phe Arg Gly Val Arg Leu Phe Arg Asp Met Gln Met Leu Pro Asn 245 250 Ser Lys Gln Asn Phe Thr Pro Arg Val Gln Gly Ile Ala Gln Ser Asn 260 265 Ala Leu Val Thr Ile Glu Gln Asn Gly Phe Val Val Tyr Gln Lys Glu 280 Val Pro Pro Gly Pro Phe Ala Ile Thr Asp Leu Gln Leu Ala Gly Gly 295 Gly Ala Asp Leu Asp Val Ser Val Lys Glu Ala Asp Gly Ser Val Thr 310 315 Thr Tyr Leu Val Pro Tyr Ala Ala Val Pro Asn Met Leu Gln Pro Gly 325 330 Val Ser Lys Tyr Asp Leu Ala Ala Gly Arg Ser His Ile Glu Gly Ala 340 345 Ser Lys Gln Ser Asp Phe Val Gln Ala Gly Tyr Gln Tyr Gly Phe Asn 360 Asn Leu Leu Thr Leu Tyr Gly Gly Ser Met Val Ala Asn Asn Tyr Tyr 375 Ala Phe Thr Leu Gly Ala Gly Trp Asn Thr Arg Ile Gly Ala Ile Ser 390 Val Asp Ala Thr Lys Ser His Ser Lys Gln Asp Asn Gly Asp Val Phe 410 Asp Gly Gln Ser Tyr Gln Ile Ala Tyr Asn Lys Phe Val Ser Gln Thr 425 Ser Thr Arg Phe Gly Leu Ala Ala Trp Arg Tyr Ser Ser Arg Asp Tyr 440

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Arg Thr Phe Asn Asp His Val Trp Ala Asn Asn Lys Asp Asn Tyr Arg
                        455
 Arg Asp Glu Asn Asp Val Tyr Asp Ile Ala Asp Tyr Tyr Gln Asn Asp
                    470
                                        475
 Phe Gly Arg Lys Asn Ser Phe Ser Ala Asn Met Ser Gln Ser Leu Pro
                 485
                                    490
 Glu Gly Trp Gly Ser Val Ser Leu Ser Thr Leu Trp Arg Asp Tyr Trp
             500
                                505
 Gly Arg Ser Gly Ser Ser Lys Asp Tyr Gln Leu Ser Tyr Ser Asn Asn
               520
                                               525
 Leu Arg Arg Ile Ser Tyr Thr Leu Ala Ala Ser Gln Ala Tyr Asp Glu
                        535
                                           540
 Asn His His Glu Glu Lys Arg Phe Asn Ile Phe Ile Ser Ile Pro Phe
                 · 550
                                        555
 Asp Trp Gly Asp Asp Val Ser Thr Pro Arg Arg Gln Ile Tyr Met Ser
                565
                                    570
 Asn Ser Thr Thr Phe Asp Asp Gln Gly Phe Ala Ser Asn Asn Thr Gly
             580
                                585
 Leu Ser Gly Thr Val Gly Ser Arg Asp Gln Phe Asn Tyr Gly Val Asn
         595
                            600
 Leu Ser His Gln His Gln Gly Asn Glu Thr Thr Ala Gly Ala Asn Leu
                    . 615
                                            620
 Thr Trp Asn Ala Pro Val Ala Thr Val Asn Gly Ser Tyr Ser Gln Ser
                    630
                                        635
 Ser Thr Tyr Arg Gln Ala Gly Ala Ser Val Ser Gly Gly Ile Val Ala
                645
                                    650
 Trp Ser Gly Gly Val Asn Leu Ala Asn Arg Leu Ser Glu Thr Phe Ala
            660
                                665
 Val Met Asn Ala Pro Gly Ile Lys Asp Ala Tyr Val Asn Gly Gln Lys
                           680
 Tyr Arg Thr Thr Asn Arg Asn Gly Val Val Ile Tyr Asp Gly Met Thr
    690 695
 Pro Tyr Arg Glu Asn His Leu Met Leu Asp Val Ser Gln Ser Asp Ser
                   710
                                        715
· Glu Ala Glu Leu Arg Gly Asn Arg Lys Ile Ala Ala Pro Tyr Arg Gly
                725
                                    730
 Ala Val Val Leu Val Asn Phe Asp Thr Asp Gln Arg Lys Pro Trp Phe
            740 . .
                                745
 Ile Lys Ala Leu Arg Ala Asp Gly Gln Ser Leu Thr Phe Gly Tyr Glu
                            760
 Val Asn Asp Ile His Gly His Asn Ile Gly Val Val Gly Gln Gly Ser
                        775
                                            780
 Gln Leu Phe Ile Arg Thr Asn Glu Val Pro Pro Ser Val Asn Val Ala
                   .790
                                        795
 Ile Asp Lys Gln Gln Gly Leu Ser Cys Thr Ile Thr Phe Gly Lys Glu
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                                    810
 Ile Asp Glu Ser Arg Asn Tyr Ile Cys Gln
            820
                                825
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<210> 287 <211> 239

<212> PRT

<213> E. Coli

<400> 287

 Met Ala Ala Ile Pro Trp Arg Pro Phe Asn Leu Arg Gly Ile Lys Met

 1
 5
 10
 15

 Lys Gly Leu Leu Ser Leu Leu Ile Phe Ser Met Val Leu Pro Ala His
 20
 25
 30

 Ala Gly Ile Val Ile Tyr Gly Thr Arg Ile Ile Tyr Pro Ala Glu Asn

40 Lys Glu Val Met Val Gln Leu Met Asn Gln Gly Asn Arg Ser Ser Leu 55 Leu Gln Ala Trp Ile Asp Asp Gly Asp Thr Ser Leu Pro Pro Glu Lys 70 Ile Gln Val Pro Phe Met Leu Thr Pro Pro Val Ala Lys Ile Gly Ala Asn Ser Gly Gln Gln Val Lys Ile Lys Ile Met Pro Asn Lys Leu Pro 100 105 Thr Asn Lys Glu Ser Ile Phe Tyr Leu Asn Val Leu Asp Ile Pro Pro 115 120 Asn Ser Pro Glu Gln Glu Gly Lys Asn Ala Leu Lys Phe Ala Met Gln 135 Asn Arg Ile Lys Leu Phe Tyr Arg Pro Ala Gly Ile Ala Pro Val Asn 150 -Lys Ala Thr Phe Lys Lys Leu Leu Val Asn Arg Ser Gly Asn Gly Leu 165 170 Val Ile Lys Asn Asp Ser Ala Asn Trp Val Thr Ile Ser Asp Val Lys 185 Ala Asn Asn Val Lys Val Asn Tyr Glu Thr Ile Met Ile Ala Pro Leu 200 Glu Ser Gln Ser Val Asn Val Lys Ser Asn Asn Ala Asn Asn Trp His 215 Leu Thr Ile Ile Asp Asp His Gly Asn Tyr Ile Ser Asp Lys Ile 230

<210> 288 <211> 180 <212> PRT <213> E. Coli

<400> 288 Met Lys Arg Ser Ile Ile Ala Ala Ala Val Phe Ser Ser Phe Phe Met Ser Ala Gly Val Phe Ala Ala Asp Val Asp Thr Gly Thr Leu Thr Ile Lys Gly Asn Ile Ala Glu Ser Pro Cys Lys Phe Glu Ala Gly Gly Asp Ser Val Ser Ile Asn Met Pro Thr Val Pro Thr Ser Val Phe Glu Gly Lys Ala Lys Tyr Ser Thr Tyr Asp Asp Ala Val Gly Val Thr Ser Ser 75 Met Leu Lys Ile Ser Cys Pro Lys Glu Val Ala Gly Val Lys Leu Ser 90 Leu Ile Thr Asn Asp Lys Ile Thr Gly Asn Asp Lys Ala Ile Ala Ser 100 105 Ser Asn Asp Thr Val Gly Tyr Tyr Leu Tyr Leu Gly Asp Asn Ser Asp 120 ٠., 125 Val Leu Asp Val Ser Ala Pro Phe Asn Ile Glu Ser Tyr Lys Thr Ala 135 Glu Gly Gln Tyr Ala Ile Pro Phe Lys Ala Lys Tyr Leu Lys Leu Thr 150 . 155 Asp Asn Ser Val Gln Ser Gly Asp Val Leu Ser Ser Leu Val Met Arg 170 Val Ala Gln Asp

<210> 289

180



/44906

<211> 112 <212> PRT <213> E. Coli

<400> 289 let Ser Ser Glu

Met Ser Ser Glu Arg Asp Leu Val Asn Phe Leu Gly Asp Phe Ser Met 10 Asp Val Ala Lys Ala Val Ile Ala Gly Gly Val Ala Thr Ala Ile Gly 20 Ser Leu Ala Ser Phe Ala Cys Val Ser Phe Gly Phe Pro Val Ile Leu 40 Val Gly Gly Ala Ile Leu Leu Thr Gly Ile Val Cys Thr Val Val Leu 55 Asn Glu Ile Asp Ala Gln Cys His Leu Ser Glu Lys Leu Lys Tyr Ala 70 75 Ile Arg Asp Gly Leu Lys Arg Gln Glu Leu Asp Lys Trp Lys Arg 85 90 Glu Asn Met Thr Pro Phe Met Tyr Val Leu Asn Thr Pro Pro Val Ile 100 105

<210> 290 <211> 193 <212> PRT <213> E. Coli

<400> 290

Met Thr Asp Tyr Leu Leu Phe Val Gly Thr Val Leu Val Asn Asn 10 Phe Val Leu Val Lys Phe Leu Gly Leu Cys Pro Phe Met Gly Val Ser 20 25 Lys Lys Leu Glu Thr Ala Met Gly Met Gly Leu Ala Thr Thr Phe Val 40 Met Thr Leu Ala Ser Ile Cys Ala Trp Leu Ile Asp Thr Trp Ile Leu Ile Pro Leu Asn Leu Ile Tyr Leu Arg Thr Leu Ala Phe Ile Leu Val 70 75 Ile Ala Val Val Gln Phe Thr Glu Met Val Val Arg Lys Thr Ser 85 90 Pro Val Leu Tyr Arg Leu Leu Gly Ile Phe Leu Pro Leu Ile Thr Thr 100 105 Asn Cys Ala Val Leu Gly Val Ala Leu Leu Asn Ile Asn Leu Gly His 115 120 Asn Phe Leu Gln Ser Ala Leu Tyr Gly Phe Ser Ala Ala Val Gly Phe 135 Ser Leu Val Met Val Leu Phe Ala Ala Ile Arg Glu Arg Leu Ala Val 150 155 Ala Asp Val Pro Ala Pro Phe Arg Gly Asn Ala Ile Ala Leu Ile Thr 165 170 Ala Gly Leu Met Ser Leu Ala Phe Met Gly Phe Ser Gly Leu Val Lys Len

<210> 291 <211> 192 <212> PRT <213> E. Coli

<400> 291 Met Asn Ala Ile Trp Ile Ala Val Ala Ala Val Ser Leu Leu Gly Leu . 10 Ala Phe Gly Ala Ile Leu Gly Tyr Ala Ser Arg Arg Phe Ala Val Glu Asp Asp Pro Val Val Glu Lys Ile Asp Glu Ile Leu Pro Gln Ser Gln Cys Gly Gln Cys Gly Tyr Pro Gly Cys Arg Pro Tyr Ala Glu Ala Ile Ser Cys Asn Gly Glu Lys Ile Asn Arg Cys Ala Pro Gly Gly Glu Ala 70 Val Met Leu Lys Ile Ala Glu Leu Leu Asn Val Glu Pro Gln Pro Leu 85 , 90 : Asp Gly Glu Ala Gln Glu Ile Thr Pro Ala Arg Met Val Ala Val Ile 100 Asp Glu Asn Asn Cys Ile Gly Cys Thr Lys Cys Ile Gln Ala Cys Pro 120 Val Asp Ala Ile Val Gly Ala Thr Arg Ala Met His Thr Val Met Ser 135 140 Asp Leu Cys Thr Gly Cys Asn Leu Cys Val Asp Pro Cys Pro Thr His 150 155 Cys Ile Ser Leu Gln Pro Val Ala Glu Thr Pro Asp Ser Trp Lys Trp 165 170 Asp Leu Asn Thr Ile Pro Val Arg Ile Ile Pro Val Glu His His Ala 180 185 .

<210> 292 <211> 740 <212> PRT <213> E. Coli

<400> 292

Met Leu Lys Leu Phe Ser Ala Phe Arg Lys Asn Lys Ile Trp Asp Phe Asn Gly Gly Ile His Pro Pro Glu Met Lys Thr Gln Ser Asn Gly Thr Pro Leu Arg Gln Val Pro Leu Ala Gln Arg Phe Val Ile Pro Leu Lys Gln His Ile Gly Ala Glu Gly Glu Leu Cys Val Ser Val Gly Asp Lys Val Leu Arg Gly Gln Pro Leu Thr Arg Gly Arg Gly Lys Met Leu Pro 75 Val His Ala Pro Thr Ser Gly Thr Val Thr Ala Ile Ala Pro His Ser 90 Thr Ala His Pro Ser Ala Leu Ala Glu Leu Ser Val Ile Ile Asp Ala 105 Asp Gly Glu Asp Cys Trp Ile Pro Arg Asp Gly Trp Ala Asp Tyr Arg 120 125 Thr Arg Ser Arg Glu Glu Leu Ile Glu Arg Ile His Gln Phe Gly Val 135 140 Ala Gly Leu Gly Gly Ala Gly Phe Pro Thr Gly Val Lys Leu Gln Gly 150 155 Gly Gly Asp Lys Ile Glu Thr Leu Ile Ile Asn Ala Ala Glu Cys Glu 165 170 Pro Tyr Ile Thr Ala Asp Asp Arg Leu Met Gln Asp Cys Ala Ala Gln 180 185. Val Val Glu Gly Ile Arg Ile Leu Ala His Ile Leu Gln Pro Arg Glu 200 Ile Leu Ile Gly Ile Glu Asp Asn Lys Pro Gln Ala Ile Ser Met Leu

215

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					_
	•				-
					-

Arg 225		Val	Leu	Ala	Asp 230	Ser	Asn	Asp	Ile	Ser 235	Leu	Arg	Val	Ile	Pro 240
				245			·		250				•	255	Thr
			Val 260					265			_		270		
		275	Val				280					285			_
•	290		Ile			295					300	-			
305			Gly		310					315				_	320
			Asp	325					330		•			335	
			Pro 340					345					350		
	•	355	Ile				360					365			
	370	•	Gln			375					380		•		
385			Pro		390					395		_			400
			Gln	405					410				•	415	_
			Gly 420 Phe					425					430		
		435	Arg				440					445		_	
	450		Glu			455				_	460			_	
465			Gln		470				_	475		_		-	480
			Val	485					490					495	
	•		500 Gly					505					510		
		515	Lys				520	•				525			
	530					535					540				
545			Ala		550					555					560
				565					570					575	Ala
	•		Pro 580					585					590		
		595	Ala				600					605			
	610		Pro			615					620				
625			Ala -		630					635					640
			Pro	645	•				650					655	
			Ala 660					665					670		
		675	Pro				680					685			
	690		Ala			695					700				
Asn	Ala	Val	Pro	Glu	Glu	Gln	Val	Asp	Pro	Arg	Lys	Ala	Ala	Val	Ala

705 710 715 720
Ala Ala Ile Ala Arg Ala Gln Ala Lys Lys Ala Ala Gln Gln Lys Val
725 730 735
Val Asn Glu Asp
740

<210> 293 <211> 352 <212> PRT <213> E. Coli

<400> 293 Met Val Phe Arg Ile Ala Ser Ser Pro Tyr Thr His Asn Gln Arg Gln 10 Thr Ser Arg Ile Met Leu Leu Val Leu Leu Ala Ala Val Pro Gly Ile 20 Ala Ala Gln Leu Trp Phe Phe Gly Trp Gly Thr Leu Val Gln Ile Leu 40 Leu Ala Ser Val Ser Ala Leu Leu Ala Glu Ala Leu Val Leu Lys Leu Arg Lys Gln Ser Val Ala Ala Thr Leu Lys Asp Asn Ser Ala Leu Leu 70 75 Thr Gly Leu Leu Ala Val Ser Ile Pro Pro Leu Ala Pro Trp 85 . 90 Met Val Val Leu Gly Thr Val Phe Ala Val Ile Ile Ala Lys Gln Leu 105 Tyr Gly Gly Leu Gly Gln Asn Pro Phe Asn Pro Ala Met Ile Gly Tyr 120 115 125 Val Val Leu Leu Ile Ser Phe Pro Val Gln Met Thr Ser Trp Leu Pro  $130\,$   $\,$   $135\,$   $\,$   $140\,$  Pro His Glu Ile Ala Val Asn Ile Pro Gly Phe Ile Asp Ala Ile Gln 150 Val Ile Phe Ser Gly His Thr Ala Ser Gly Gly Asp Met Asn Thr Leu 170 Arg Leu Gly Ile Asp Gly Ile Ser Gln Ala Thr Pro Leu Asp Thr Phe 185 Lys Thr Ser Val Arg Ala Gly His Ser Val Glu Gln Ile Met Gln Tyr 200 Pro Ile Tyr Ser Gly Ile Leu Ala Gly Ala Gly Trp Gln Trp Val Asn 215 220 Leu Ala Trp Leu Ala Gly Gly Val Trp Leu Leu Trp Gln Lys Ala Ile **.** . . 230 235 Arg Trp His Ile Pro Leu Ser Phe Leu Val Thr Leu Ala Leu Cys Ala 250 245 Met Leu Gly Trp Leu Phe Ser Pro Glu Thr Leu Ala Ala Pro Gln Ile 265 His Leu Leu Ser Gly Ala Thr Met Leu Gly Ala Phe Phe Ile Leu Thr 280 285 Asp Pro Val Thr Ala Ser Thr Thr Asn Arg Gly Arg Leu Ile Phe Gly 295 300 Ala Leu Ala Gly Leu Leu Val Trp Leu Ile Arg Ser Phe Gly Gly Tyr 310 315 Pro Asp Gly Val Ala Phe Ala Val Leu Leu Ala Asn Ile Thr Val Pro 330 Leu Ile Asp Tyr Tyr Thr Arg Pro Arg Val Tyr Gly His Arg Lys Gly

<210> 294

00/44906

<211> 206
<212> PRT
<213> E. Coli

<400> 294

Met Leu Lys Thr Ile Arg Lys His Gly Ile Thr Leu Ala Leu Phe Ala Ala Gly Ser Thr Gly Leu Thr Ala Ala Ile Asn Gln Met Thr Lys Thr 25 Thr Ile Ala Glu Gln Ala Ser Leu Gln Gln Lys Ala Leu Phe Asp Gln 40 Val Leu Pro Ala Glu Arg Tyr Asn Asn Ala Leu Ala Gln Ser Cys Tyr Leu Val Thr Ala Pro Glu Leu Gly Lys Gly Glu His Arg Val Tyr Ile 75 Ala Lys Gln Asp Asp Lys Pro Val Ala Ala Val Leu Glu Ala Thr Ala 90 Pro Asp Gly Tyr Ser Gly Ala Ile Gln Leu Leu Val Gly Ala Asp Phe 100 105 Asn Gly Thr Val Leu Gly Thr Arg Val Thr Glu His His Glu Thr Pro 120 Gly Leu Gly Asp Lys Ile Glu Leu Arg Leu Ser Asp Trp Ile Thr His 135 Phe Ala Gly Lys Lys Ile Ser Gly Ala Asp Asp Ala His Trp Ala Val 150 -155 Lys Lys Asp Gly Gly Asp Phe Asp Gln Phe Thr Gly Ala Thr Ile Thr 165 170 Pro Arg Ala Val Val Asn Ala Val Lys Arg Ala Gly Leu Tyr Ala Gln 180 185 Thr Leu Pro Ala Gln Leu Ser Gln Leu Pro Ala Cys Gly Glu 200 195

<210> 295 <211> 231 <212> PRT

<213> E. Coli

<400> 295 Met Ser Glu Ile Lys Asp Val Ile Val Gln Gly Leu Trp Lys Asn Asn 10 Ser Ala Leu Val Gln Leu Leu Gly Leu Cys Pro Leu Leu Ala Val Thr . 25 Ser Thr Ala Thr Asn Ala Leu Gly Leu Gly Leu Ala Thr Thr Leu Val 40 Leu Thr Leu Thr Asn Leu Thr Ile Ser Thr Leu Arg His Trp Thr Pro 55 60 Ala Glu Ile Arg Ile Pro Ile Tyr Val Met Ile Ile Ala Ser Val Val 70 75 Ser Ala Val Gln Met Leu Ile Asn Ala Tyr Ala Phe Gly Leu Tyr Gln 8.5 90 Ser Leu Gly Ile Phe Ile Pro Leu Ile Val Thr Asn Cys Ile Val Val 100 105 Gly Arg Ala Glu Ala Phe Ala Ala Lys Lys Gly Pro Ala Leu Ser Ala 120 125 Leu Asp Gly Phe Ser Ile Gly Met Gly Ala Thr Cys Ala Met Phe Val 135 140 Leu Gly Ser Leu Arg Glu Ile Ile Gly Asn Gly Thr Leu Phe Asp Gly 150 155 Ala Asp Ala Leu Leu Gly Ser Trp Ala Lys Val Leu Arg Val Glu Ile

 Phe His Thr Asp Ser Pro Phe Leu Leu Ala Met Leu Pro Pro Gly Ala 180
 185
 190

 Phe Ile Gly Leu Gly Leu Met Leu Ala Gly Lys Tyr Leu Ile Asp Glu 195
 200
 205

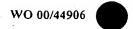
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 215
 220

 Asn Gly Glu Thr Gly Asn Val 225
 230
 230

<210> 296 <211> 211 <212> PRT <213> E. Coli

<400> 296 Met Asn Lys Ala Lys Arg Leu Glu Ile Leu Thr Arg Leu Arg Glu Asn Asn Pro His Pro Thr Thr Glu Leu Asn Phe Ser Ser Pro Phe Glu Leu Leu Ile Ala Val Leu Leu Ser Ala Gln Ala Thr Asp Val Ser Val Asn Lys Ala Thr Ala Lys Leu Tyr Pro Val Ala Asn Thr Pro Ala Ala Met Leu Glu Leu Gly Val Glu Gly Val Lys Thr Tyr Ile Lys Thr Ile Gly Leu Tyr Asn Ser Lys Ala Glu Asn Ile Ile Lys Thr Cys Arg Ile Leu Leu Glu Gln His Asn Gly Glu Val Pro Glu Asp Arg Ala Ala Leu Glu 105 Ala Leu Pro Gly Val Gly Arg Lys Thr Ala Asn Val Val Leu Asn Thr 120 125 Ala Phe Gly Trp Pro Thr Ile Ala Val Asp Thr His Ile Phe Arg Val 135 Cys Asn Arg Thr Gln Phe Ala Pro Gly Lys Asn Val Glu Gln Val Glu 150 155 Glu Lys Leu Leu Lys Val Val Pro Ala Glu Phe Lys Val Asp Cys His 165 170 175 His Trp Leu Ile Leu His Gly Arg Tyr Thr Cys Ile Ala Arg Lys Pro 180 185 Arg Cys Gly Ser Cys Ile Ile Glu Asp Leu Cys Glu Tyr Lys Glu Lys 195 200 Val Asp Ile 210

<210> 297 <211> 167 <212> PRT <213> E. Coli



65					70		•			75					80
Thr	Ser	Ala	Val	Thr 85	Ala	Ile	Val	Thr	Gly 90	Ser	Thr	Asp	Asn	Thr 95	Gly
Tyr	Tyr	Lys	Asn 100	Glu	Gly	Thr	Ala	Glu 105	Asn	Ile	Gln	Ile	Glu 110	Leu	Arg
Asp	-	Gln 115	Asp	Ala	Ala	Leu	Lys 120	Asn	Gly	Asp	Ser	Lys 125	Thr	Val	Ile
Val	Asp 130	Glu	Ile	Thr	Arg	Asn 135	Ala	Gln	Phe	Pro	Leu 140	Lys	Ala	Arg	Ala
Ile 145	Thr	Val	Asn	Gly	Asn 150	Ala	Ser	Gln	Gly	Thr 155	Ile	Glu	Ala	Leu	Ile 160
Asn	Val	Ile	Tyr	Thr 165	Trp	Gln									

<210> 298 <211> 176 <212> PRT <213> E. Coli

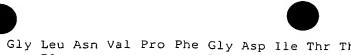
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 Tyr
 Thr
 Val
 Lys
 Pro
 Pro
 Thr
 Gly
 Asn
 Glu
 Gln
 In
 Asn
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 Gln
 In
 In
 Asn
 In
 <th



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Leu Asn Tyr Ser Tyr Ser Asn Asn Ile Trp Gln Asn Asp Arg Asp His
                85
                                     90
Leu Leu Ala Phe Thr Leu Asn Val Pro Phe Ser His Trp Met Arg Thr
            100
                                105
Asp Ser Gln Ser Ala Phe Arg Asn Ser Asn Ala Ser Tyr Ser Met Ser
                             120
                                                 125
Asn Asp Leu Lys Gly Gly Met Thr Asn Leu Ser Gly Val Tyr Gly Thr
                        135
                                             140
Leu Leu Pro Asp Asn Asn Leu Asn Tyr Ser Val Gln Val Gly Asn Thr
                    150
                                         155
His Gly Gly Asn Thr Ser Ser Gly Thr Ser Gly Tyr Ser Ser Leu Asn
                165
                                     170
Tyr Arg Gly Ala Tyr Gly Asn Thr Asn Val Gly Tyr Ser Arg Ser Gly
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                                185
Asp Ser Ser Gln Ile Tyr Tyr Gly Met Ser Gly Gly Ile Ile Ala His
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Ala Asp Gly Ile Thr Phe Gly Gln Pro Leu Gly Asp Thr Met Val Leu
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                                             220
Val Lys Ala Pro Gly Ala Asp Asn Val Lys Ile Glu Asn Gln Thr Gly
                    230
                                         235
Ile His Thr Asp Trp Arg Gly Tyr Ala Ile Leu Pro Phe Ala Thr Glu
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                                     250
Tyr Arg Glu Asn Arg Val Ala Leu Asn Ala Asn Ser Leu Ala Asp Asn
            260
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Val Glu Leu Asp Glu Thr Val Val Thr Val Ile Pro Thr His Gly Ala
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Ile Ala Arg Ala Thr Phe Asn Ala Gln Ile Gly Gly Lys Val Leu Met
                        295
                                             300
Thr Leu Lys Tyr Gly Asn Lys Ser Val Pro Phe Gly Ala Ile Val Thr
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His Gly Glu Asn Lys Asn Gly Ser Ile Val Ala Glu Asn Gly Gln Val
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Tyr Leu Thr Gly Leu Pro Gln Ser Gly Gln Leu Gln Val Ser Trp Gly
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                                345
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<212> PRT

<213> E. Coli

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 Leu Leu Asp Leu Val Cys Ala Lys Ile Thr Ala Trp Glu Glu Ser Ala 50

 Pro Glu Phe Ala Glu Phe Asn Ala Met Ala Gln Ala Met Pro Gly Gly Glu Glu Asn Pro 75

 Ile Ala Val Ile Arg Thr Leu Met Asp Gln Tyr Gly Leu Thr Leu Ser 85

 Asp Leu Pro Glu Ile Gly Ser Lys Ser Met Val Ser Arg Val Leu Ser



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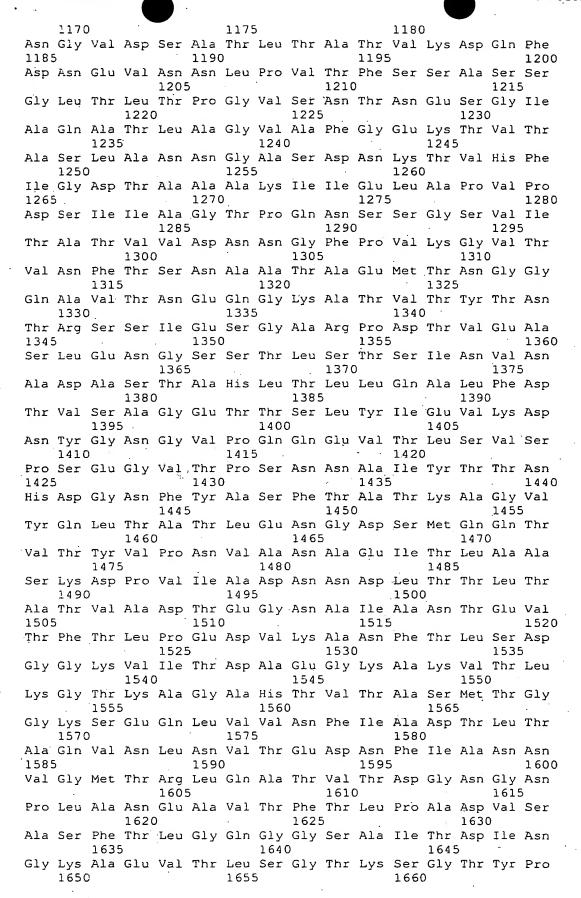
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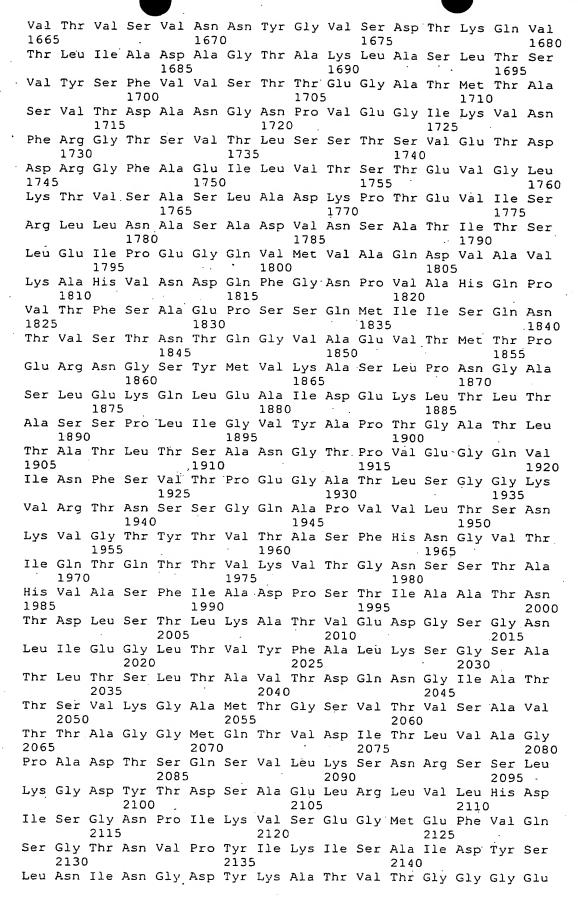
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		195			•		200					205			
Phe	Gly 210		Ala	Arg	Ile	Thr 215	Leu	Gly	Val	Asp	Glu 220	Asp	Phe	Ser	Leu
Lys 225	Asn	Ser	Gln	Phe	Asp 230	Phe	Leu	His	Pro	Trp 235	Tyr	Glu	·Thr	Pro	Asp 240
Asn	Leu	Phe	Phe	Ser 245	Gln	His	Thr	Leu	His 250	Arg	Thr	Asp	Glu	Arg 255	
Gln	Ile	Asn	Asn 260	Gly	Leu	Gly	Trp	Arg 265	His	Phe	Thr	Pro	Thr 270	Trp	Met
Ser	Gly	11e 275	Asn	Phe	Phe	Phe	Asp 280	His	Asp	Leu	Ser	Arg 285	Tyr	His	Ser
	Ala 290					295					300		•		
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	Asn			325					330					335	
	Ser		340					345	-			-	350		-
	Gln	355					360					365			_
	Ser 370					375					380				
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	Asp			405	•				410					415	
	Gln		420					425				_	430		
	Gly	435					440					445			
	Tyr 450			::		455					460		-		
465	Gly Tyr				470					475					480
	Gly			485					490			•		495	
	Ala		500					505	_	-			510		
		515					520					525			Glu
	530					535					540				Ser
545					550					555				_	560
	Val Thr			565					570				•	575	
	Leu		580					585			_		590		
	Asp	595					600					605			
	610 Gly					615					620				
625					630					635					640
	Asp	•		645 <sup>.</sup>					650					655	
	Ser		660					665		-	_	_	670		
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			_												
Lys	Pro 690	val	Lys	Glu	Gln	Lys 695	Gln	Gln	Leu	Asn	Asn 700	Ala	Val	Ser	Ile
Asp 705	Asn	Val	Lys	Pro	Gly 710	Val	Thr	Thr	Asp	Trp 715	Lys	Glu	Thr	Ala	Asp 720
Gly	Val	Tyr	Lys	Ala 725	Thr	Tyr	Thr	Ala	Tyr 730	Thr	Lys	Gly	Ser	Gly 735	
Thr	Ala	Lys	Leu 740	Leu	Met	Gln	Asn	Trp 745	Asn	Glu	Asp	Leu	His 750	Thr	Ala
Gly	Phe	Ile 755	Ile	Asp	Ala	Asn	Pro 760	Gln	Ser	Ala	Lys	Ile 765	Ala	Thr	Leu
	770				•	Val 775					780				
Val 785	Ser	Val	Asn	Val ·	Ala 790	Asp	Glu	Gly	Ser	Asn 795	Pro	Ile	Asn	Asp	His 800
				805		Leu			810					815	
			820	=		Asp		825	_				830	•	
_		835				Asp	840					845			
	850					11e 855					860				•
865			_		870	Lys				875				•	880
				885		Thr			890	_				895	
			900			Val		905					910		
		915				Val	920					925			
	930				·	Asn 935					940				
945					950	Asn				955			=	_	960
				965		Leu			970					975	
			980			Tyr		985					990		
_		995		_	-	Lys	1000	)				1005	<u>,</u>	٠.	_
	1010	)				1015	•				1020	)			Thr
1025					1030		•			1035	<u> </u>				1040
				1045	)	Asp			1050	)				1055	5
			1060	)		Ile		1065	5				1070	)	
		1075	5		•		1080	)				1085	<u>,</u>		
	1090	)				1095 Gln	5				1100	)			
1105	,				1110			•		1115	<b>.</b>				1120
_				1125	,	Gly			1130	)				1135	5
			1140	)		Ser		1145	5				1150	)	_
		1155	<b>.</b>			Leu	1160	)				1165	ò		
a	J L L			- 4 1	- 4 -	u	11	- J. C	<u> </u>	- y 3	٦.٠٠٢	O L U	- T E	TIIT	O T Y





2150 2155 2160 Gly Ile Ala Thr Leu Ile Pro Val Leu Asn Gly Val His Gln Ala Gly 2165 2170 Leu Ser Thr Thr Ile Gln. Phe Thr Arg Ala Glu Asp Lys Ile Met Ser 2180 2185 Gly Thr Val Ser Val Asn Gly Thr Asp Leu Pro Thr Thr Thr Phe Pro 2195 2200 2205 Ser Gln Gly Phe Thr Gly Ala Tyr Tyr Gln Leu Asn Asn Asp Asn Phe 2215 2220 Ala Pro Gly Lys Thr Ala Ala Asp Tyr Glu Phe Ser Ser Ala Ser 2230 2235 Trp Val Asp Val Asp Ala Thr Gly Lys Val Thr Phe Lys Asn Val Gly 2245 2250 Ser Asn Ser Glu Arg Ile Thr Ala Thr Pro Lys Ser Gly Gly Pro Ser 2260 2265 2270 Tyr Val Tyr Glu Ile Arg Val Lys Ser Trp Trp Val Asn Ala Gly Glu 2275 2280 2285 Ala Phe Met Ile Tyr Ser Leu Ala Glu Asn Phe Cys Ser Ser Asn Gly 2295 2300 Tyr Thr Leu Pro Arg Ala Asn Tyr Leu Asn His Cys Ser Ser Arg Gly 2310 2315 Ile Gly Ser Leu Tyr Ser Glu Trp Gly Asp Met Gly His Tyr Thr Thr 2325 2330 Asp Ala Gly Phe Gln Ser Asn Met Tyr Trp Ser Ser Ser Pro Ala Asn . 2340 2345 2350 Ser Ser Glu Gln Tyr Val Val Ser Leu Ala Thr Gly Asp Gln Ser Val 2360 2365 Phe Glu Lys Leu Gly Phe Ala Tyr Ala Thr Cys Tyr Lys Asn Leu 2375 2380

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 Tyr
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                                    90
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Trp Glu Trp Gln Thr Gln Ala Ala Leu Asn Thr Val Leu Gln Gln Ile
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Glu Lys Glu Gln Asn Thr Ile Ile Ala Thr Asp Arg Gln Leu Tyr Gln
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Asn Gly Ile Thr Ser Ser Val Glu Gly Val Glu Thr Asp Ile Asn Ala
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                                        155
Ser Lys Thr Arg Gln Gln Leu Asn Asp Val Ala Gly Lys Met Lys Ile
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                                   170
Ile Glu Ala Arg Leu Ser Ala Leu Thr Asn Asn Gln Thr Lys Ser Leu
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Lys Leu Lys Pro Val Ala Leu Pro Lys Val Ala Ser Gln Leu Pro Asp
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Glu Leu Gly Tyr Ser Leu Leu Ala Arg Arg Ala Asp Leu Gln Ala Ala
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His Trp Tyr Val Glu Ser Ser Leu Ser Thr Ile Asp Ala Ala Lys Ala
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                                        235
Ala Phe Tyr Pro Asp Ile Asn Leu Met Ala Phe Leu Gln Gln Asp Ala
                245
                                    250
Leu His Leu Ser Asp Leu Phe Arg His Ser Ala Gln Gln Met Gly Val
                               265
Thr Ala Gly Leu Thr Leu Pro Ile Phe Asp Ser Gly Arg Leu Asn Ala
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Asn Leu Asp Ile Ala Lys Ala Glu Ser Asn Leu Ser Ile Ala Ser Tyr
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Asn Lys Ala Val Val Glu Ala Val Asn Asp Val Ala Arg Ala Ala Ser
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Gln Val Gln Thr Leu Ala Glu Lys Asn Gln His Gln Ala Gln Ile Glu
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                                    330
Arg Asp Ala Leu Arg Val Val Gly Leu Ala Gln Ala Arg Phe Asn Ala
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Gly Ile Ile Ala Gly Ser Arg Val Ser Glu Ala Arg Ile Pro Ala Leu
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Ser Ile Gln Leu Thr Gly Ala Leu Gly Gly Gly Tyr Lys Arg
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<212> PRT

<213> E. Coli

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<212> PRT

<213> E. Coli

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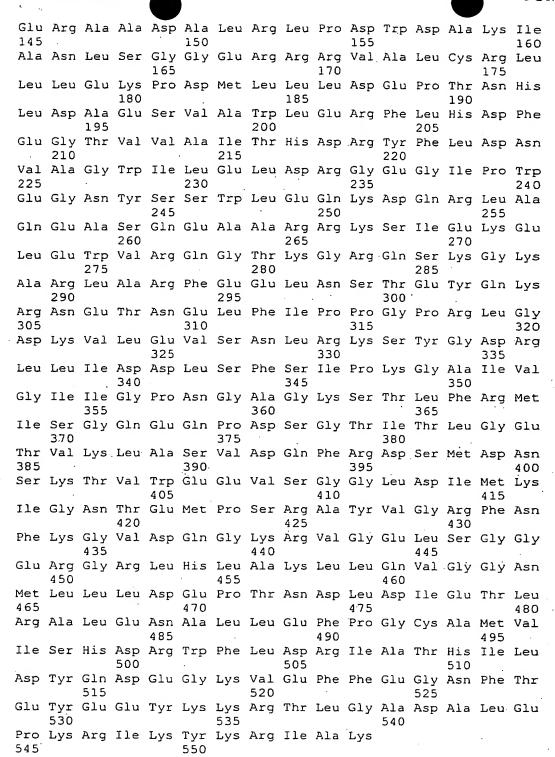


Pro Ala Pro Leu Arg Lys Leu Cys Asp Leu Thr Thr Leu Lys Leu Glu Pro Asn Ser Phe Ile Asp Glu Asp Leu Arg Gln Tyr Tyr Ser Asp Leu . 55 Leu Trp Ser Val Lys Thr Gln Glu Gly Val Gly Tyr Ile Tyr Val Val Ile Glu His Gln Ser Lys Pro Glu Glu Leu Met Ala Phe Arg Met Met 90 Arg Tyr Ser Ile Ala Ala Met Gln Asn His Leu Asp Ala Gly Tyr Lys 100 105 Glu Leu Pro Leu Val Leu Pro Met Leu Phe Tyr His Gly Cys Arg Ser 115 120 -125 Pro Tyr Pro Tyr Ser Leu Cys Trp Leu Asp Glu Phe Ala Glu Pro Ala . 130 135 140 Ile Ala Arg Lys Ile Tyr Ser Ser Ala Phe Pro Leu Val Asp Ile Thr 150 155 Val Val Pro Asp Asp Glu Ile Met Gln His Arg Lys Met Ala Leu Leu 165 170 Glu Leu Ile Gln Lys His Ile Arg Gln Arg Asp Leu Leu Gly Leu Val 180 185 Asp Gln Ile Val Ser Leu Leu Val Thr Gly Asn Thr Asn Asp Arg Gln 200 Leu Lys Ala Leu Phe Asn Tyr Val Leu Gln Thr Gly Asp Ala Gln Arg 215 220 Phe Arg Ala Phe Ile Gly Glu Ile Ala Glu Arg Ala Pro Gln Glu Lys 230 235 Glu Lys Leu Met Thr Ile Ala Asp Arg Leu Arg Glu Glu Gly Ala Met 245 250 Gln Gly Lys His Glu Glu Ala Leu Arg Ile Ala Gln Glu Met Leu Asp 260 265 Arg Gly Leu Asp Arg Glu Leu Val Met Met Val Thr Arg Leu Ser Pro 275 280 Asp Asp Leu Ile Ala Gln Ser His 295

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<400> 309



<210> 310 <211> 283 <212> PRT <213> E. Coli

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165

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Cys Phe Phe Gly Thr Leu Lys Ser Glu Cys Phe Tyr Leu Asp Glu Phe

Ser Asn Ile Ser Glu Leu Lys Asp Ala Val Thr Glu Tyr Ile Glu Tyr

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275 280 Thr Ile Ala Ser Trp Phe Gly Gly Gly Thr Gly Trp Asn Trp Leu Thr 295 Thr Ile Ser Leu Tyr Leu Gln Pro Gly Gln Pro Leu Tyr Val Leu Leu 310 315 Tyr Ala Ser Ala Ile Ile Phe Phe Cys Phe Phe Tyr Thr Ala Leu Val 325 330 Phe Asn Pro Arg Glu Thr Ala Asp Asn Leu Lys Lys Ser Gly Ala Phe 345 Val Pro Gly Ile Arg Pro Gly Glu Gln Thr Ala Lys Tyr Ile Asp Lys 360 Val Met Thr Arg Leu Thr Leu Val Gly Ala Leu Tyr Ile Thr Phe Ile 375 380 Cys Leu Ile Pro Glu Phe Met Arg Asp Ala Met Lys Val Pro Phe Tyr 390 395 Phe Gly Gly Thr Ser Leu Leu Ile Val Val Val Ile Met Asp Phe 405 410 Met Ala Gln Val Gln Thr Leu Met Met Ser Ser Gln Tyr Glu Ser Ala 420 425 Leu Lys Lys Ala Asn Leu Lys Gly Tyr Gly Arg 435

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Leu Pro Lys His Lys Ala Thr Leu Leu Gly Leu Gly Leu Arg Arg Ile

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Gly His Thr Val Glu Arg Glu Asp Thr Pro Ala Ile Arg Gly Met Ile

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Asn Ala Val Ser Phe Met Val Lys Val Glu Glu

<210> 315 <211> 167 <212> PRT <213> E. Colì

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<210> 316 <211> 117 <212> PRT <213> E. Coli

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<210> 317 <211> 177



<212> PRT <213> E. Coli

<400> 317 Met Ser Arg Val Ala Lys Ala Pro Val Val Val Pro Ala Gly Val Asp 10 Val Lys Ile Asn Gly Gln Val Ile Thr Ile Lys Gly Lys Asn Gly Glu Leu Thr Arg Thr Leu Asn Asp Ala Val Glu Val Lys His Ala Asp Asn Thr Leu Thr Phe Gly Pro Arg Asp Gly Tyr Ala Asp Gly Trp Ala Gln Ala Gly Thr Ala Arg Ala Leu Leu Asn Ser Met Val Ile Gly Val Thr Glu Gly Phe Thr Lys Lys Leu Gln Leu Val Gly Val Gly Tyr Arg Ala Ala Val Lys Gly Asn Val Ile Asn Leu Ser Leu Gly Phe Ser His Pro 105 Val Asp His Gln Leu Pro Ala Gly Ile Thr Ala Glu Cys Pro Thr Gln 120 Thr Glu Ile Val Leu Lys Gly Ala Asp Lys Gln Val Ile Gly Gln Val 135 Ala Ala Asp Leu Arg Ala Tyr Arg Arg Pro Glu Pro Tyr Lys Gly Lys 150 155 Gly Val Arg Tyr Ala Asp Glu Val Val Arg Thr Lys Glu Ala Lys Lys 165 · 170 Lys

<210> 318 <211> 130 <212> PRT <213> E. Coli

<400> 318 Met Ser Met Gln Asp Pro Ile Ala Asp Met Leu Thr Arg Ile Arg Asn 10 Gly Gln Ala Ala Asn Lys Ala Ala Val Thr Met Pro Ser Ser Lys Leu 25 Lys Val Ala Ile Ala Asn Val Leu Lys Glu Glu Gly Phe Ile Glu Asp 40. Phe Lys Val Glu Gly Asp Thr Lys Pro Glu Leu Glu Leu Thr Leu Lys 55 Tyr Phe Gln Gly Lys Ala Val Val Glu Ser Ile Gln Arg Val Ser Arg 70 75 Pro Gly Leu Arg Ile Tyr Lys Arg Lys Asp Glu Leu Pro Lys Val Met 90 Ala Gly Leu Gly Ile Ala Val Val Ser Thr Ser Lys Gly Val Met Thr 100 105 Asp Arg Ala Ala Arg Gln Ala Gly Leu Gly Gly Glu Ile Ile Cys Tyr Val Ala

<210> 319 <211> 101 <212> PRT <213> E. Coli

. <400> 319 Met Ala Lys Gln Ser Met Lys Ala Arg Glu Val Lys Arg Val Ala Leu 10 Ala Asp Lys Tyr Phe Ala Lys Arg Ala Glu Leu Lys Ala Ile Ile Ser 20 25. Asp Val Asn Ala Ser Asp Glu Asp Arg Trp Asn Ala Val Leu Lys Leu 40 Gln Thr Leu Pro Arg Asp Ser Ser Pro Ser Arg Gln Arg Asn Arg Cys 55 Arg Gln Thr Gly Arg Pro His Gly Phe Leu Arg Lys Phe Gly Leu Ser 70 75 Arg Ile Lys Val Arg Glu Ala Ala Met Arg Gly Glu Ile Pro Gly Leu 85 90 Lys Lys Ala Ser Trp 100 -

<210> 320 <211> 179 <212> PRT <213> E. Coli

<400> 320 Met Ala Lys Leu His Asp Tyr Tyr Lys Asp Glu Val Val Lys Lys Leu Met Thr Glu Phe Asn Tyr Asn Ser Val Met Gln Val Pro Arg Val Glu Lys Ile Thr Leu Asn Met Gly Val Gly Glu Ala Ile Ala Asp Lys 40 Leu Leu Asp Asn Ala Ala Ala Asp Leu Ala Ala Ile Ser Gly Gln Lys 55 Pro Leu Ile Thr Lys Ala Arg Lys Ser Val Ala Gly Phe Lys Ile Arg 70 Gln Gly Tyr Pro Ile Gly Cys Lys Val Thr Leu Arg Gly Glu Arg Met 90 Trp Glu Phe Phe Glu Arg Leu Ile Thr Ile Ala Val Pro Arg Ile Arg 105 Asp Fhe Arg Gly Leu Ser Ala Lys Ser Phe Asp Gly Arg Gly Asn Tyr 120 Ser Met Gly Val Arg Glu Gln Ile Ile Phe Pro Glu Ile Asp Tyr Asp 135 140 Lys Val Asp Arg Val Arg Gly Leu Asp Ile Thr Ile Thr Thr Thr Ala 150 155 Lys Ser Asp Glu Glu Gly Arg Ala Leu Leu Ala Ala Phe Asp Phe Pro 170

<210> 321Z <211> 104 <212> PRT <213> E. Coli

Phe Arg Lys



 Lys
 Val
 Ile
 Val
 Glu
 Gly
 Ile
 Asn
 Leu
 Val
 Lys
 Lys
 His
 Gln
 Lys
 Pro
 45

 Val
 Pro
 Ala
 Leu
 Asn
 Gln
 Pro
 Gly
 Gly
 Ile
 Val
 Glu
 Lys
 Glu
 Ala
 Ala

 50
 55
 60
 60
 60
 60
 1
 Ile
 Ala
 Ala

<210> 322 <211> 123 <212> PRT <213> E. Coli

 Met
 Ile
 Gln
 Gln
 Thr
 Met
 Leu
 Asn
 Val
 Ala
 Asp
 Asn
 Ser
 Gly
 Ala

 Arg
 Arg
 Val
 Met
 Cys
 Ile
 Lys
 Val
 Leu
 Gly
 Gly
 Ser
 His
 Arg
 Arg
 Tyr

 Arg
 Arg
 Val
 Met
 Cys
 Ile
 Lys
 Val
 Leu
 Gly
 Ser
 His
 Arg
 Arg
 Tyr

 Arg
 Gly
 Val
 Gly
 Asp
 Ile
 Lys
 Ile
 Thr
 Ile
 Lys
 Glu
 Ala
 Ile
 Pro

 Arg
 Gly
 Lys
 Lys
 Lys
 Arg
 Pro
 Asp
 Gly
 Ser
 Val
 Ile
 Arg
 Pro
 Asp

 Gly
 Asn
 Ala
 Cys
 Val
 Leu
 Asn
 Asn
 Asn
 Ser
 Glu
 Glu
 Pro
 Ile
 Gly
 Pro
 Ile
 Gly
 Pro
 Ile
 Ile
 Ile
 Ile
 Ile
 Ile
 I

<210> 323 <211> 188 <212> PRT <213> E. Coli

<400> 323 e Lvs Glv (

Met Phe Lys Gly Gln Lys Thr Leu Ala Ala Leu Ala Val Ser Leu Leu Phe Thr Ala Pro Val Tyr Ala Ala Asp Glu Gly Ser Gly Glu Ile His Phe Lys Gly Glu Val Ile Glu Ala Pro Cys Glu Ile His Pro Glu Asp Ile Asp Lys Asn Ile Asp Leu Gly Gln Val Thr Thr His Ile Asn Arg Glu His His Ser Asn Lys Val Ala Val Asp Ile Arg Leu Ile Asn 7.5 . Cys Asp Leu Pro Ala Ser Asp Asn Gly Ser Gly Met Pro Val Ser Lys 85 90 Val Gly Val Thr Phe Asp Ser Thr Ala Lys Thr Thr Gly Ala Thr Pro 105 Leu Leu Ser Asn Thr Ser Ala Gly Glu Ala Thr Gly Val Gly Val Arg 120 Leu Met Asp Lys Asn Asp Gly Asn Ile Val Leu Gly Ser Ala Ala Pro 135 140 Asp Leu Asp Leu Asp Ala Ser Ser Ser Glu Gln Thr Leu Asn Phe Phe



> <210> 324 <211> 427 <212> PRT <213> E. Coli

<400> 324 Met Ala Asp Thr Lys Ala Lys Leu Thr Leu Asn Gly Asp Thr Ala Val Glu Leu Asp Val Leu Lys Gly Thr Leu Gly Gln Asp Val Ile Asp Ile Arg Thr Leu Gly Ser Lys Gly Val Phe Thr Phe Asp Pro Gly Phe Thr Ser Thr Ala Ser Cys Glu Ser Lys Ile Thr Phe Ile Asp Gly Asp Glu Gly Ile Leu Leu His Arg Gly Phe Pro Ile Asp Gln Leu Ala Thr Asp 75 Ser Asn Tyr Leu Glu Val Cys Tyr Ile Leu Leu Asn Gly Glu Lys Pro 90 Thr Gln Glu Gln Tyr Asp Glu Phe Lys Thr Thr Val Thr Arg His Thr 105 Met Ile His Glu Gln Ile Thr Arg Leu Phe His Ala Phe Arg Arg Asp 120 125 Ser His Pro Met Ala Val Met Cys Gly Ile Thr Gly Ala Leu Ala Ala 135 140 Phe Tyr His Asp Ser Leu Asp Val Asn Asn Pro Arg His Arg Glu Ile 150 155 Ala Ala Phe Arg Leu Leu Ser Lys Met Pro Thr Met Ala Ala Met Cys 165 170 Tyr Lys Tyr Ser Ile Gly Gln Pro Phe Val Tyr Pro Arg Asn Asp Leu 185 Ser Tyr Ala Gly Asn Phe Leu Asn Met Met Phe Ser Thr Pro Cys Glu 200 Pro Tyr Glu Val Asn Pro Ile Leu Glu Arg Ala Met Asp Arg Ile Leu 215 220 Ile Leu His Ala Asp His Glu Gln Asn Ala Ser Thr Ser Thr Val Arg 230 235 Thr Ala Gly Ser Ser Gly Ala Asn Pro Phe Ala Cys Ile Ala Ala Gly 245 250 Ile Ala Ser Leu Trp Gly Pro Ala His Gly Gly Ala Asn Glu Ala Ala 265 270 Leu Lys Met Leu Glu Glu Ile Ser Ser Val Lys His Ile Pro Glu Phe 280 285 Val Arg Arg Ala Lys Asp Lys Asn Asp Ser Phe Arg Leu Met Gly Phe 295 300 Gly His Arg Val Tyr Lys Asn Tyr Asp Pro Arg Ala Thr Val Met Arg 310 315 Glu Thr Cys His Glu Val Leu Lys Glu Leu Gly Thr Lys Asp Asp Leu 325 330 Leu Glu Val Ala Met Glu Leu Glu Asn Ile Ala Leu Asn Asp Pro Tyr . 345 Phe Ile Glu Lys Lys Leu Tyr Pro Asn Val Asp Phe Tyr Ser Gly Ile 360

Ile Leu Lys Ala Met Gly Ile Pro Ser Ser Met Phe Thr Val Ile Phe



Ala Met Ala Arg Thr Val Gly Trp Ile Ala His Trp Ser Glu Met His 385 390 395 395 400 Ser Asp Gly Met Lys Ile Ala Arg Pro Arg Gln Leu Tyr Thr Gly Tyr 405 415 Glu Lys Arg Asp Phe Lys Ser Asp Ile Lys Arg 425

<210> 325 <211> 477 <212> PRT <213> E. Coli

<400> 325 Met Lys Val Thr Leu Pro Glu Phe Glu Arg Ala Gly Val Met Val Val 10 Gly Asp Val Met Leu Asp Arg Tyr Trp Tyr Gly Pro Thr Ser Arg Ile Ser Pro Glu Ala Pro Val Pro Val Val Lys Val Asn Thr Ile Glu Glu 40 Arg Pro Gly Gly Ala Ala Asn Val Ala Met Asn Ile Ala Ser Leu Gly 55 Ala Asn Ala Arg Leu Val Gly Leu Thr Gly Ile Asp Asp Ala Ala Arg 70 Ala Leu Ser Lys Ser Leu Ala Asp Val Asn Val Lys Cys Asp Phe Val 85 90 Ser Val Pro Thr His Pro Thr Ile Thr Lys Leu Arg Val Leu Ser Arg 100 105 Asn Gln Gln Leu Ile Arg Leu Asp Phe Glu Glu Gly Phe Glu Gly Val 115. 120 Asp Pro Gln Pro Leu His Glu Arg Ile Asn Gln Ala Leu Ser Ser Ile 135 140 Gly Ala Leu Val Leu Ser Asp Tyr Ala Lys Gly Ala Leu Ala Ser Val 150 155 Gln Gln Met Ile Gln Leu Ala Arg Lys Ala Gly Val Pro Val Leu Ile 165 170 Asp Pro Lys Gly Thr Asp Phe Glu Arg Tyr Arg Gly Ala Thr Leu Leu · 180 185 Thr Pro Asn Leu Ser Glu Phe Glu Ala Val Val Gly Lys Cys Lys Thr 200 Glu Glu Glu Ile Val Glu Arg Gly Met Lys Leu Ile Ala Asp Tyr Glu 215 220 Leu Ser Ala Leu Leu Val Thr Arg Ser Glu Gln Gly Met Ser Leu Leu 230 235 Gln Pro Gly Lys Ala Pro Leu His Met Pro Thr Gln Ala Gln Glu Val 245 250 Tyr Asp Val Thr Gly Ala Gly Asp Thr Val Ile Gly Val Leu Ala Ala 260 265 Thr Leu Ala Ala Gly Asn Ser Leu Glu Glu Ala Cys Phe Phe Ala Asn 280 285 Ala Ala Ala Gly Val Val Gly Lys Leu Gly Thr Ser Thr Val Ser 295 300 Pro Ile Glu Leu Glu Asn Ala Val Arg Gly Arg Ala Asp Thr Gly Phe 310 315 Gly Val Met Thr Glu Glu Glu Leu Lys Leu Ala Val Ala Ala Ala Arg 325 330 Lys Arg Gly Glu Lys Val Val Met Thr Asn Gly Val Phe Asp Ile Leu 340 345 His Ala Gly His Val Ser Tyr Leu Ala Asn Ala Arg Lys Leu Gly Asp 360 Arg Leu Ile Val Ala Val Asn Ser Asp Ala Ser Thr Lys Arg Leu Lys

375 380 370 Gly Asp Ser Arg Pro Val Asn Pro Leu Glu Gln Arg Met Ile Val Leu 390 395 Gly Ala Leu Glu Ala Val Asp Trp Val Val Ser Phe Glu Glu Asp Thr 405 410 Pro Gln Arg Leu Ile Ala Gly Ile Leu Pro Asp Leu Leu Val Lys Gly 425 Gly Asp Tyr Lys Pro Glu Glu Ile Ala Gly Ser Lys Glu Val Trp Ala 440 Asn Gly Gly Glu Val Leu Val Leu Asn Phe Glu Asp Gly Cys Ser Thr 455 460 Thr Asn Ile Ile Lys Lys Ile Gln Gln Asp Lys Gly 470

<210> 326 <211> 946 <212> PRT <213> E. Coli

<400> 326

Met Lys Pro Leu Ser Ser Pro Leu Gln Gln Tyr Trp Gln Thr Val Val 10 Glu Arg Leu Pro Glu Pro Leu Ala Glu Glu Ser Leu Ser Ala Gln Ala 25 Lys Ser Val Leu Thr Phe Ser Asp Phe Val Gln Asp Ser Val Ile Ala 40 His Pro Glu Trp Leu Thr Glu Leu Glu Ser Gln Pro Pro Gln Ala Asp 55 Glu Trp Gln His Tyr Ala Ala Trp Leu Gln Glu Ala Leu Cys Asn Val 70 . 75 Ser Asp Glu Ala Gly Leu Met Arg Glu Leu Arg Leu Phe Arg Arg Arg 85 . 90 Ile Met Val Arg Ile Ala Trp Ala Gln Thr Leu Ala Leu Val Thr Glu 105 100 Glu Ser Ile Leu Gln Gln Leu Ser Tyr Leu Ala Glu Thr Leu Ile Val 120 Ala Ala Arg Asp Trp Leu Tyr Asp Ala Cys Cys Arg Glu Trp Gly Thr 135 140 Pro Cys Asn Ala Gln Gly Glu Ala Gln Pro Leu Leu Ile Leu Gly Met 150 155 Gly Lys Leu Gly Gly Gly Glu Leu Asn Phe Ser Ser Asp Ile Asp Leu 165 170 Ile Phe Ala Trp Pro Glu His Gly Cys Thr Gln Gly Gly Arg Arg Glu 180 185 Leu Asp Asn Ala Gln Phe Phe Thr Arg Met Gly Gln Arg Leu Ile Lys 200 Val Leu Asp Gln Pro Thr Gln Asp Gly Phe Val Tyr Arg Val Asp Met 215 220 Arg Leu Arg Pro Phe Gly Glu Ser Gly Pro Leu Val Leu Ser Phe Ala 230 % Ala Leu Glu Asp Tyr Tyr Gln Glu Gln Gly Arg Asp Trp Glu Arg Tyr 245 Ala Met Val Lys Ala Arg Ile Met Gly Asp Ser Glu Gly Val Tyr Ala 260 265 Asn Glu Leu Arg Ala Met Leu Arg Pro Phe Val Phe Arg Arg Tyr Ile 280 Asp Phe Ser Val Ile Gln Ser Leu Arg Asn Met Lys Gly Met Ile Ala 295 Arg Glu Val Arg Arg Gly Leu Thr Asp Asn Ile Lys Leu Gly Ala 305 310 320



			7												
Gl	y Gly	Ile	Arg	Glu 325	Ile	Glu	Phe	Ile	Val 330		Val	Phe	Gln	Leu 335	Ile
Ar	g Gly	Gly	/ Arg 340	Glu		Ser	Leu	Gln 345			Ser	Leu	Leu 350	Pro	Thr
Let	ı Ser	Ala 355	. Ile		Glu	Leu	His 360	Leu	Leu	Ser	Glu	Asn 365	Asp	Ala	Glu
GÌI	1 Leu 370	Arg		Ala	Tyr	Leu 375			Arg	Arg	Leu 380		Asn	Leu	Leu
Glr 385	Ser		Asn	Asp	Glu 390	Gln	Thr	Gln	Thr	Leu 395		Ser	Asp	Glu	Leu 400
	Arg	Ala	Arg	Leu 405	Ala		Ala	Met	Asp 410	Phe	Ala	Asp	Trp	Pro 415	Gln
Lev	Thr	Gly	Ala 420	Leu		Ala	His	Met 425			Val	Arg	Arg 430		Phe
Asr	Glu	Leu 435		Gly	Asp	Asp	Glu 440		Glu	Thr	Gln	Glu 445		Ser	Leu
Ser	Glu 450		Trp	Arg	Glu	Leu 455	Trp	Gln	Asp	Ala	Leu 460	Gln	Glu	Asp	Asp
465		•			470					475					480
	Thr			485					490				_	495	
Gly	Pro	Arg	Gly 500	Arg	Gln	Val	Leu	Asp 505	His	Leu	Met	Pro	His 510	Leu	Leu
	Asp	515		•			520					525	•		
	Ala 530					535					540	-			
545					550					555					560
	Ser			565	. •		•		570					575	•
	Glu		580					585					590		
		595					600					605			
	Glu 610					615					620				
625	Leu				630					635					640
	.Val			645			•		650					655	
	Val		660					665					670	_	
	His	675					680					685			
	Lys 690					695					700			_	
705					710					715					720
	Ile			725					730					735	
	Leu		740					745					750		_
	Arg	755					760					765			
	Ala 770					775					780				
785	Ala				790					795					800
17 d	His	FILE	vah	VIG	AGT	wr à	ur d	OTU.	116	MEL	INT	ьeп	Pro	Arg	GTU

810 815 Gly Lys Thr Leu Gln Thr Glu Val Arg Glu Met Arg Glu Lys Met Arg 825 Ala His Leu Gly Asn Lys His Arg Asp Arg Phe Asp Ile Lys Ala Asp 840 Glu Gly Gly Ile Thr Asp Ile Glu Phe Ile Thr Gln Tyr Leu Val Leu . . 855 860 Arg Tyr Ala His Glu Lys Pro Lys Leu Thr Arg Trp Ser Asp Asn Val 870 875 Arg Ile Leu Glu Leu Leu Ala Gln Asn Asp Ile Met Glu Glu Gln Glu 885 890 Ala Met Ala Leu Thr Arg Ala Tyr Thr Thr Leu Arg Asp Glu Leu His 905 910 His Leu Ala Leu Gln Glu Leu Pro Gly His Val Ser Glu Asp Cys Phe 920 Thr Ala Glu Arg Glu Leu Val Arg Ala Ser Trp Gln Lys Trp Leu Val 935 930 940 Glu Glu 945

<210> 327 <211> 433 <212> PRT <213> E. Coli

<400> 327

Met Ala Gln Glu Ile Glu Leu Lys Phe Ile Val Asn His Ser Ala Val Glu Ala Leu Arg Asp His Leu Asn Thr Leu Gly Gly Glu His His Asp - 20 Pro Val Gln Leu Leu Asn Ile Tyr Tyr Glu Thr Pro Asp Asn Trp Leu 40 Arg Gly His Asp Met Gly Leu Arg Ile Arg Gly Glu Asn Gly Arg Tyr Glu Met Thr Met Lys Val Ala Gly Arg Val Thr Gly Gly Leu His Gln 70 Arg Pro Glu Tyr Asn Val Ala Leu Ser Glu Pro Thr Leu Asp Leu Ala 90 Gln Leu Pro Thr Glu Val Trp Pro Asn Gly Glu Leu Pro Ala Asp Leu 100 105 Ala Ser Arg Val Gln Pro Leu Phe Ser Thr Asp Phe Tyr Arg Glu Lys 115 . 120 Trp Leu Val Ala Val Asp Gly Ser Gln Ile Glu Ile Ala Leu Asp Gln 135 Gly Glu Val Lys Ala Gly Glu Phe Ala Glu Pro Ile Cys Glu Leu Glu 150 155 Leu Glu Leu Leu Ser Gly Asp Thr Arg Ala Val Leu Lys Leu Ala Asn 170 Gln Leu Val Ser Gln Thr Gly Leu Arg Gln Gly Ser, Leu Ser Lys Ala 180 Ala Arg Gly Tyr His Leu Ala Gln Gly Asn Pro Ala Arg Glu Ile Lys Pro Thr Thr Ile Leu His Val Ala Ala Lys Ala Asp Val Glu Gln Gly

Leu Glu Ala Ala Leu Glu Leu Ala Leu Ala Gln Trp Gln Tyr His Glu

Glu Leu Trp Val Arg Gly Asn Asp Ala Ala Lys Glu Gln Val Leu Ala

Ala Ile Ser Leu Val Arg His Thr Leu Met Leu Phe Gly Gly Ile Val

215

230

245

250

.220



260 265 Pro Arg Lys Ala Ser Thr His Leu Arg Asp Leu Leu Thr Gln Cys Glu 275 280 285 Ala Thr Ile Ala Ser Ala Val Ser Ala Val Thr Ala Val Tyr Ser Thr 290 295 Glu Thr Ala Met Ala Lys Leu Ala Leu Thr Glu Trp Leu Val Ser Lys 310 315 Ala Trp Gln Pro Phe Leu Asp Ala Lys Ala Gln Gly Lys Ile Ser Asp 330 Ser Phe Lys Arg Phe Ala Asp Ile His Leu Ser Arg His Ala Ala Glu 345 350 Leu Lys Ser Val Phe Cys Gln Pro Leu Gly Asp Arg Tyr Arg Asp Gln 360 365 Leu Pro Arg Leu Thr Arg Asp Ile Asp Ser Ile Leu Leu Ala Gly 375 . 380 Tyr Tyr Asp Pro Val Val Ala Gln Ala Trp Leu Glu Asn Trp Gln Gly 390 395 Leu His His Ala Ile Ala Thr Gly Gln Arg Ile Glu Ile Glu His Phe 405 410 Arg Asn Glu Ala Asn Asn Gln Glu Pro Phe Trp Leu His Ser Gly Lys 425 Arq

<210> 328 <211> 70 <212> PRT <213> E. Coli

<400> 328

 Met
 Ser
 Gly
 Lys
 Met
 Thr
 Gly
 Ile
 Val
 Lys
 Trp
 Phe
 Asn
 Ala
 Asp
 Lys

 Gly
 Phe
 Gly
 Phe
 Ile
 Thr
 Pro
 Asp
 Asp
 Gly
 Ser
 Lys
 Asp
 Val
 Phe
 Val

 His
 Phe
 Ser
 Ala
 Ile
 Gln
 Asn
 Asp
 Gly
 Tyr
 Lys
 Ser
 Leu
 Asp
 Gly
 Gly
 Ala
 Lys
 Gly
 Pro
 Ala
 Ala

<210> 329 <211> 523 <212> PRT <213> E. Coli

<400> 329

 Met
 Arg
 Asp
 Ile
 Val
 Asp
 Pro
 Val
 Phe
 Ser
 Ile
 Gly
 Ile
 Ser
 Ser
 Leu

 Trp
 Asp
 Glu
 Leu
 Arg
 His
 Met
 Pro
 Ala
 Gly
 Gly
 Val
 Trp
 Trp
 Phe
 Asn

 Val
 Asp
 Arg
 His
 Glu
 Asp
 Ala
 Ile
 Ser
 Leu
 Ala
 Asp
 Gln
 Thr
 Ala
 Ala

 Ser
 Gln
 Ala
 Glu
 Thr
 Ala
 His
 Val
 Ala
 Val
 Ile
 Ser
 Met
 Asp
 Ser
 Asp

 Ser
 Gln
 Ala
 Glu
 Thr
 Ala
 His
 Val
 Ala
 Val
 Ile
 Ser
 Met
 Asp
 Ser
 Asp

 Ser
 Ala
 Lys
 Ile
 Asp
 Ser
 Gln
 Gln
 Pro
 Asp
 Asp
 Ser

" .			1									•			
Lys	Leu	Phe	Ser	Met 85	Leu	Asn	His	Glu	Lys 90	Gly	Leu	Tyr	Tyr	Leu 95	Thr
	Asp		100					105					110	Ile	
	C);s	115					120					125			_
	Trp 130					135					140			_	
145					150					155					160
				165					170					175	Phe ·
	Gly		180					185					190		_
•	Val Thr	195			•		200					205			
	210 Lys					215					220				_
225	Ser				230					235					240
	Ala			245					250	•				255	
	Ala		260					265					270		
	Arg	275					280					.285			
Leu	290 Arg	Ala	Thr	Asp	Glu	295 Arg	Leu	Leu	Leu	Ala	300 Cys	Gly	Ala	Asn	Met
305 Val	Ile	Pro	Trp	Asn	310 Ala	Pro	Leu	Ser	Arg	315 Cys	Leu	Thr	Met	Ile	320 Glu
Ser	Val	Gln		325 Gln	Lys	Phe	Ser		330 Tyr	Val	Pro	Glu		335 Ile	Thr
Thr	Leu		340 Ser	Met	Thr	Gln		345 Leu	Lys	Leu	Arg		350 Phe	Gln	Lys
Trp	Asp 370	355 Val	Phe	Cys	Asn	Ala 375	360 Val	Asn	Asn	Met	Met 380	365 Asn	Asn	Pro	Leu
Leu 385	Pro	Ala	His	Gly	Lys 390		Val	Leu	Val	Ala 395		Arg	Pro	Val	Pro-
	Ile	Arg	Val	Glu 405		Ala		Thr	Leu 410		Arg	Pro	Asn	Arg 415	
Gly	Asp	Ile	Met 420		Ile	Gly	Gly.	Asn 425		Leu	Val	Leu	Phe 430		Ser
	Cys	435					440					445	lle		
	Pro 450					455					460				
465	Gln				470					475					480
	Trp			485					490					495	
	Ala		500					505			Ile	Pro	Glu 510	Pro	Met
Arg	Leu	Leu 515	Asp	Asp	Ala	Val	Glu 520	Arg	Ser	Ser					

<210> 330

<211> 62 <212> PRT

<213> E. Coli

<400> 330

 Met Thr Ile Ser Asp Ile Ile Glu Ile Ile Val Val Cys Ala Leu Ile

 1
 5
 10
 15

 Phe Phe Pro Leu Gly Tyr Leu Ala Arg His Ser Leu Arg Arg Ile Arg 20
 25
 30

 Asp Thr Leu Arg Leu Phe Phe Ala Lys Pro Arg Tyr Val Lys Pro Ala 35
 40
 45

 Gly Thr Leu Arg Arg Thr Glu Lys Ala Arg Ala Thr Lys Lys 50
 55

<210> 331 <211> 559 <212> PRT <213> E. Coli

<400> 331

Met Thr Gln Phe Thr Gln Asn Thr Ala Met Pro Ser Ser Leu Trp Gln 10 Tyr Trp Arg Gly Leu Ser Gly Trp Asn Phe Tyr Phe Leu Val Lys Phe Gly Leu Leu Trp Ala Gly Tyr Leu Asn Phe His Pro Leu Leu Asn Leu Val Phe Ala Ala Phe Leu Leu Met Pro Leu Pro Arg Tyr Ser Leu His Arg Leu Arg His Trp Ile Ala Leu Pro Ile Gly Phe Ala Leu Phe Trp His Asp Thr Trp Leu Pro Gly Pro Glu Ser Ile Met Ser Gln Gly Ser 85 90 Gln Val Ala Gly Phe Ser Thr Asp Tyr Leu Ile Asp Leu Val Thr Arg 105 Phe Ile Asn Trp Gln Met Ile Gly Ala Ile Phe Val Leu Leu Val Ala 120 Trp Leu Phe Leu Ser Gln Trp Ile Arg Ile Thr Val Phe Val Val Ala 135 Ile Leu Leu Trp Leu Asn Val Leu Thr Leu Ala Gly Pro Ser Phe Ser 150 155 Leu Trp Pro Ala Gly Gln Pro Thr Thr Thr Val Thr Thr Gly Gly 165 170 Asn Ala Ala Ala Thr Val Ala Ala Thr Gly Gly Ala Pro Val Val Gly 180 185 Asp Met Pro Ala Gln Thr Ala Pro Pro Thr Thr Ala Asn Leu Asn Ala 200 205 Trp Leu Asn Asn Phe Tyr Asn Ala Glu Ala Lys Arg Lys Ser Thr Phe 215 220 Pro Ser Ser Leu Pro Ala Asp Ala Gln Pro Phe Glu Leu Leu Val Ile 230 235 Asn Ile Cys Ser Leu Ser Trp Ser Asp Ile Glu Ala Ala Gly Leu Met 245 250 Ser His Pro Leu Trp Ser His Phe Asp Ile Glu Phe Lys Asn Phe Asn 265 Ser Ala Thr Ser Tyr Ser Gly Pro Ala Ala Ile Arg Leu Leu Arg Ala 280 Ser Cys Gly Gln Thr Ser His Thr Asn Leu Tyr Gln Pro Ala Asn Asn 295 300 Asp Cys Tyr Leu Phe Asp Asn Leu Ser Lys Leu Gly Phe Thr Gln His - 310 315 Leu Met Met Gly His Asn Gly Gln Phe Gly Gly Phe Leu Lys Glu Val 330



Arg Glu Asn Gly Gly Met Gln Ser Glu Leu Met Asp Gln Thr Asn Leu 340 345 Pro Val Ile Leu Leu Gly Phe Asp Gly Ser Pro Val Tyr Asp Asp Thr 360 365 Ala Val Leu Asn Arg Trp Leu Asp Val Thr Glu Lys Asp Lys Asn Ser 375 380 Arg Ser Ala Thr Phe Tyr Asn Thr Leu Pro Leu His Asp Gly Asn His 390 395 Tyr Pro Gly Val Ser Lys Thr Ala Asp Tyr Lys Ala Arg Ala Gln Lys 405 410 Phe Phe Asp Glu Leu Asp Ala Phe Phe Thr Glu Leu Glu Lys Ser Gly 420 425 430 Arg Lys Val Met Val Val Val Pro Glu His Gly Gly Ala Leu Lys 440 445 Gly Asp Arg Met Gln Val Ser Gly Leu Arg Asp Ile Pro Ser Pro Ser 455 460 Ile Thr Asp Val Pro Val Gly Val Lys Phe Phe Gly Met Lys Ala Pro 470 475 His Gln Gly Ala Pro Ile Val Ile Glu Gln Pro Ser Ser Phe Leu Ala 485 490 Ile Ser Asp Leu Val Val Arg Val Leu Asp Gly Lys Ile Phe Thr Glu 500 505 Asp Asn Val Asp Trp Lys Lys Leu Thr Ser Gly Leu Pro Gln Thr Ala 515 520 525 Pro Val Ser Glu Asn Ser Asn Ala Val Val Ile Gln Tyr Gln Asp Lys 535 540 Pro Tyr Val Arg Leu Asn Gly Gly Asp Trp Val Pro Tyr Pro Gln . 550

<210> 332 <211> 127 <212> PRT <213> E. Coli

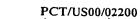
<400> 332

Met Glu Gly Ser Arg Met Lys Tyr Arg Ile Ala Leu Ala Val Ser Leu 10 Phe Ala Leu Ser Ala Gly Ser Tyr Ala Thr Thr Leu Cys Gln Glu Lys 20 25 Glu Gln Asn Ile Leu Lys Glu Ile Ser Tyr Ala Glu Lys His Gln Asn 40 Gln Asn Arg Ile Asp Gly Leu Asn Lys Ala Leu Ser Glu Val Arg Ala 55 Asn Cys Ser Asp Ser Gln Leu Arg Ala Asp His Gln Lys Lys Ile Ala 70 75 Lys Gln Lys Asp Glu Val Ala Glu Arg Gln Gln Asp Leu Ala Glu Ala 8.5 90 Lys Gln Lys Gly Asp Ala Asp Lys Ile Ala Lys Arg Glu Arg Lys Leu 100 105 Ala Glu Ala Gln Glu Glu Leu Lys Lys Leu Glu Ala Arg Asp Tyr 120

<210> 333 <211> 101 <212> PRT <213> E. Coli

<400> 333

Met Ser Lys Glu His Thr Thr Glu His Leu Arg Ala Glu Leu Lys Ser



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Leu Ser Asp Thr Leu Glu Glu Val Leu Ser Ser Ser Gly Glu Lys Ser
                                25
Lys Glu Glu Leu Ser Lys Ile Arg Ser Lys Ala Glu Gln Ala Leu Lys
Gln Ser Arg Tyr Arg Leu Gly Glu Thr Gly Asp Ala Ile Ala Lys Gln
Thr Arg Val Ala Ala Ala Arg Ala Asp Glu Tyr Val Arg Glu Asn Pro
Trp Thr Gly Val Gly Ile Gly Ala Ala Ile Gly Val Val Leu Gly Val
Leu Leu Ser Arg Arg
            100
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<210> 334 <211> 134 <212> PRT <213> E. Coli

<400> 334

Met Ala Asp Thr His His Ala Gln Gly Pro Gly Lys Ser Val Leu Gly Ile Gly Gln Arg Ile Val Ser Ile Met Val Glu Met Val Glu Thr Arg 25 Leu Arg Leu Ala Val Val Glu Leu Glu Glu Glu Lys Ala Asn Leu Phe 40 Gln Leu Leu Met Leu Gly Leu Thr Met Leu Phe Ala Ala Phe Gly 55 Leu Met Ser Leu Met Val Leu Ile Ile Trp Ala Val Asp Pro Gln Tyr 70 75 Arg Leu Asn Ala Met Ile Ala Thr Thr Val Val Leu Leu Leu Ala 85 90 Leu Ile Gly Gly Ile Trp Thr Leu Arg Lys Ser Arg Lys Ser Thr Leu 100 105 Leu Arg His Thr Arg His Glu Leu Ala Asn Asp Arg Gln Leu Leu Glu 115 120 Glu Glu Ser Arg Glu Gln 130

<210> 335 <211> 99 <212> PRT <213> E. Coli

<400> 335 Met Ser Ser Lys Val Glu Arg Glu Arg Lys Ala Gln Leu Leu Ser 10 Gln Ile Gln Gln Arg Leu Asp Leu Ser Ala Ser Arg Arg Glu Trp 25 Leu Glu Thr Thr Gly Ala Tyr Asp Arg Arg Trp Asn Met Leu Leu Ser 40 Leu Arg Ser Trp Ala Leu Val Gly Ser Ser Val Met Ala Ile Trp Thr 55 Ile Arg His Pro Asn Met Leu Val Arg Trp Ala Arg Arg Gly Phe Gly 70 Val Trp Ser Ala Trp Arg Leu Val Lys Thr Thr Leu Lys Gln Gln Leu Arg Gly

<210> 336 <211> 160 <212> PRT <213> E. Coli

<400> 336 Met Ile Leu Ser Ile Asp Ser Asn Asp Ala Asn Thr Ala Pro Leu His Lys Lys Thr Ile Ser Ser Leu Ser Gly Ala Val Glu Ser Met Met Lys 25 Lys Leu Glu Asp Val Gly Val Leu Val Ala Arg Ile Leu Met Pro Ile Leu Phe Ile Thr Ala Gly Trp Gly Lys Ile Thr Gly Tyr Ala Gly Thr Gln Gln Tyr Met Glu Ala Met Gly Val Pro Gly Phe Met Leu Pro Leu Val Ile Leu Leu Glu Phe Gly Gly Gly Leu Ala Ile Leu Phe Gly Phe Leu Thr Arg Thr Thr Ala Leu Phe Thr Ala Gly Phe Thr Leu Leu Thr 100 105 Ala Phe Leu Phe His Ser Asn Phe Ala Glu Gly Val Asn Ser Leu Met 120 Phe Met Lys Asn Leu Thr Ile Ser Gly Gly Phe Leu Leu Leu Ala Ile 135 Thr Gly Pro Gly Ala Tyr Ser Ile Asp Arg Leu Leu Asn Lys Lys Trp 150 . 155

<210> 337 <211> 296 <212> PRT <213> E. Coli

<400> 337

Met Ile Lys Lys Thr Thr Glu Ile Asp Ala Ile Leu Leu Asn Leu Asn Lys Ala Ile Asp Ala His Tyr Gln Trp Leu Val Ser Met Phe His Ser Val Val Ala Arg Asp Ala Ser Lys Pro Glu Ile Thr Asp Asn His Ser 40 Tyr Gly Leu Cys Gln Phe Gly Arg Trp Ile Asp His Leu Gly Pro Leu Asp Asn Asp Glu Leu Pro Tyr Val Arg Leu Met Asp Ser Ala His Gln His Met His Asn Cys Gly Arg Glu Leu Met Leu Ala Ile Val Glu Asn His Trp Gln Asp Ala His Phe Asp Ala Phe Gln Glu Gly Leu Leu Ser 100 105 Phe Thr Ala Ala Leu Thr Asp Tyr Lys Ile Tyr Leu Leu Thr Ile Arg 120 Ser Asn Met Asp Val Leu Thr Gly Leu Pro Gly Arg Arg Val Leu Asp 135 1.40 Glu Ser Phe Asp His Gln Leu Arg Asn Ala Glu Pro Leu Asn Leu Tyr 155 Leu Met Leu Leu Asp Ile Asp Arg Phe Lys Leu Val Asn Asp Thr Tyr 170

Gly His Leu Ile Gly Asp Val Val Leu Arg Thr Leu Ala Thr Tyr Leu 180 185 Ala Ser Trp Thr Arg Asp Tyr Glu Thr Val Tyr Arg Tyr Gly Glu 200 Glu Phe Ile Ile Val Lys Ala Ala Asn Asp Glu Glu Ala Cys Arg 215 220 Ala Gly Val Arg Ile Cys Gln Leu Val Asp Asn His Ala Ile Thr His 230 235 Ser Glu Gly His Ile Asn Ile Thr Val Thr Ala Gly Val Ser Arg Ala 245 250 Phe Pro Glu Glu Pro Leu Asp Val Val Ile Gly Arg Ala Asp Arg Ala 260 265 Met Tyr Glu Gly Lys Gln Thr Gly Arg Asn Arg Cys Met Phe Ile Asp 275 280 Glu Gln Asn Val Ile Asn Arg Val

<210> 338 <211> 203 <212> PRT <213> E. Coli

<400> 338 Met Arg Leu Arg Val Val Pro Gly Phe Ile Ser Pro Pro Pro Gly Phe .10 Gly Gly Leu Gly Tyr Thr Pro Thr Ala Arg Ala Cys Val Asn Ile Ser 25 Ile Pro Leu Gln Leu Arg Val Ile Asp Met Leu Asp Val Phe Thr Pro 40 Leu Leu Lys Leu Phe Ala Asn Glu Pro Leu Glu Arg Leu Met Tyr Thr 55 60 Ile Ile Ile Phe Gly Leu Thr Leu Trp Leu Ile Pro Lys Glu Phe Thr 75 Val Ala Phe Asn Ala Tyr Thr Glu Ile Pro Trp Leu Phe Gln Ile Ile 85 90 Val Phe Ala Phe Ser Phe Val Val Ala Ile Ser Phe Ser Arg Leu Arg 105 Ala His Ile Gln Lys His Tyr Ser Leu Leu Pro Glu Gln Arg Val Leu 120 . Leu Arg Leu Ser Glu Lys Glu Ile Ala Val Phe Lys Asp Phe Leu Lys 135 Thr Gly Asn Leu Ile Ile Thr Ser Pro Cys Arg Asn Pro Val Met Lys 150 155 Lys Leu Glu Arg Lys Gly Ile Ile Gln His Gln Ser Asp Ser Ala Asn 165 170 Cys Ser Tyr Tyr Leu Val Thr Glu Lys Tyr Ser His Phe Met Lys Leu . 180 185 Phe Trp Asn Ser Arg Ser Arg Arg Phe Asn Arg 200

<210> 339 <211> 58 <212> PRT <213> E. Coli

<400> 339
Met Leu Gln Pro Ser Ala Arg Thr Ser Phe Gly Phe Lys Cys Phe

Ala Phe Gly Ile Arg His Gly Ser Glu Arg Ser Ile Leu Val Gly Glu 20 25 30

His Ala Ala His Gln Gly Phe Val Val Ala Glu Val Asp Phe Leu His 35 40 45

Phe Ala Asn Leu Thr Ser Cys Cys Tyr Val 50 55

<210> 340 <211> 1426 <212> PRT <213> E. Coli

<400> 340 Met Ser Gly Lys Pro Ala Ala Arg Gln Gly Asp Met Thr Gln Tyr Gly Gly Pro Ile Val Gln Gly Ser Ala Gly Val Arg Ile Gly Ala Pro Thr Gly Val Ala Cys Ser Val Cys Pro Gly Gly Met Thr Ser Gly Asn Pro Val Asn Pro Leu Leu Gly Ala Lys Val Leu Pro Gly Glu Thr Asp Leu Ala Leu Pro Gly Pro Leu Pro Phe Ile Leu Ser Arg Thr Tyr Ser Ser Tyr Arg Thr Lys Thr Pro Ala Pro Val Gly Val Phe Gly Pro Gly Trp 90 Lys Ala Pro Ser Asp Ile Arg Leu Gln Leu Arg Asp Asp Gly Leu Ile 105 Leu Asn Asp Asn Gly Gly Arg Ser Ile His Phe Glu Pro Leu Leu Pro 120 Gly Glu Ala Val Tyr Ser Arg Ser Glu Ser Met Trp Leu Val Arg Gly 135 140 Gly Lys Ala Ala Gln Pro Asp Gly His Thr Leu Ala Arg Leu Trp Gly 150 155 Ala Leu Pro Pro Asp Ile Arg Leu Ser Pro His Leu Tyr Leu Ala Thr 170 Asn Ser Ala Gln Gly Pro Trp Trp Ile Leu Gly Trp Ser Glu Arg Val 185 Pro Gly Ala Glu Asp Val Leu Pro Ala Pro Leu Pro Pro Tyr Arg Val 200 Leu Thr Gly Met Ala Asp Arg Phe Gly Arg Thr Leu Thr Tyr Arg Arg 215 220 Glu Ala Ala Gly Asp Leu Ala Gly Glu Ile Thr Gly Val Thr Asp Gly 230 235 Ala Gly Arg Glu Phe Arg Leu Val Leu Thr Thr Gln Ala Gln Arg Ala 245 250 Glu Glu Ala Arg Thr Ser Ser Leu Ser Ser Ser Asp Ser Ser Arg Pro 265 Leu Ser Ala Ser Ala Phe Pro Asp Thr Leu Pro Gly Thr Glu Tyr Gly 280 Pro Asp Arg Gly Ile Arg Leu Ser Ala Val Trp Leu Met His Asp Pro 295 300 Ala Tyr Pro Glu Ser Leu Pro Ala Ala Pro Leu Val Arg Tyr Thr Tyr 310 315 Thr Glu Ala Gly Glu Leu Leu Ala Val Tyr Asp Arg Ser Asn Thr Gln 325 330 Val Arg Ala Phe Thr Tyr Asp Ala Gln His Pro Gly Arg Met Val Ala 345 . His Arg Tyr Ala Gly Arg Pro Glu Met Arg Tyr Arg Tyr Asp Asp Thr 360

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	385	)				390	)	•			395	,				Arç 400
					40	5				410	)				415	Lys
				420	)				425	)				430	)	Ala
•	•		43.	)				440	)				445	5		Tyr
		450	)			•	455	<b>,</b>				460	)			Asp
	465					470	)				475		-			Ala 480
					485	•				490					495	Pro
				. 500	)		•		505					510		Tyr
			515	)				520	٠.				525			Ala
		530					535					540				Leu
	242					550					555					Arg 560
					565					570					575	Tyr
				580		Arg			585					590		
			595			Tyr		600					605			
		610			٠.	Gly	615					620	•			_
	625					630 Arg					635					Glu 640
					645					650°					655	Ser
		•		660		Thr			665					670		
			675			Asp		680					685			
		690					695					700				Glu
	705					710 Gly	•				715					720
					725	Val				730					735	
				/4 U		Cys			745					750		
			155			Glu		760					765			
		770				Pro	775					780			-	
	/85					790 Leu					795					800
					805	Asp				810					815	
				820		Ser			825					830		
			835			Leu		840					845			
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	850	)				855					. 860				
Asp 865	Arg		Tyr	Ġly	Trp 870	Ser				Asp 875	Leu	Val	Arg	Ile	Ser 880
Gly	Pro	Arg	Gln	Thr 885	Arg	Glu	Tyr	Gly	Tyr 890	Ser	Ala	Thr	Gly	Arg 895	Leu
Glu	Ser	Val	Arg 900	Thr	Leu	Ala	Pro	Asp 905	Leu		Ile	Arg	Ile 910	Pro	Tyr
Ala	Thr	Asp 915	Pro	Ala	Gly	Asn	Arg 920	Leu	Pro	Asp	Pro	Glu 925	Leu	His	Pro
Asp	Ser 930	Thr	Leu	Thr	Val	Trp 935	Pro	Asp	Asn	Arg	Ile 940	Ala	Glu	Asp	Ala
945					950					955					Thr 960.
				965			Ile		970					975	His
			980				His	985					990	_	
		995					Glu 1000	)				100	5		
	101	0				101					102	0			
102	5				1030	)				103	5				Trp 1040
				1045	5		Val		105	0				105	5
			1060	)			Phe	1065	5.				1076	0	
		1075	5				Ala 1080	)				1089	5 .		
	1090	)				109					110	0			
110	5			•	1110	)	Arg			111!	5				1120
				1125	5		Ala		1130	9				1139	5
			1140	)			Val	1145	5				1150	)	
		1155	5				Asp 1160	)				1169	5		
	1170	)				1175					1180	)			
1185	)				1190	)	Asn			1195	5				1200
•				1205	<b>)</b>		Asp		1210	)				1215	•
			1220	)				1225	1				1230	)	
		1235	,				Trp 1240					1245	,		
	1250	)				1255					1260	)			
1265	)				1270	)	Gly			1275	<u>,                                      </u>				1280
				1285			Asp		1290	)				1295	
			1300					1305					1310	)	
		1315					Thr 1320					1325			_
ATG	GIu 1330	Tnr	Asp	Asn	Leu	Arg 1335	Lys	Glu	Ser	Tyr	Pro 1340		Lys	Arg	Lys

Cys Pro Thr Gly Thr Asp Arg Val Ala Ala Tyr His Thr His Gly Ala 1350 1355 Asp Ser His Gly Asp Tyr Val Asp Glu Phe Phe Ser Ser Ser Asp Lys . 1365 1370 Asn Leu Val Arg Ser Lys Asp Asn Asn Leu Glu Ala Phe Tyr Leu Ala 1380 1385 Thr Pro Asp Gly Arg Phe Glu Ala Leu Asn Asn Lys Gly Glu Tyr Ile 1395 1400 Phe Ile Arg Asn Ser Val Pro Gly Leu Ser Ser Val Cys Ile Pro Tyr 1415 His Asp 1425

<210> 341 <211> 122 <212> PRT <213> E. Coli

<400> 341 Met Lys Tyr Ser Ser Ile Phe Ser Met Leu Ser Phe Phe Ile Leu Phe 10 Ala Cys Asn Glu Thr Ala Val Tyr Gly Ser Asp Glu Asn Ile Ile Phe 25 Met Arg Tyr Val Glu Lys Leu His Leu Asp Lys Tyr Ser Val Lys Asn 40 Thr Val Lys Thr Glu Thr Met Ala Ile Gln Leu Ala Glu Ile Tyr Val 55 Arg Tyr Arg Tyr Gly Glu Arg Ile Ala Glu Glu Glu Lys Pro Tyr Leu 70 75 Ile Thr Glu Leu Pro Asp Ser Trp Val Val Glu Gly Ala Lys Leu Pro 85... 90 Tyr Glu Val Ala Gly Gly Val Phe Ile Ile Glu Ile Asn Lys Lys Asn 100 105 Gly Cys Val Leu Asn Phe Leu His Ser Lys

<210> 342 <211> 236 <212> PRT <213> E. Coli

<400> 342

Met Leu Ala Leu Met Asp Ala Asp Gly Asn Ile Ala Trp Ser Gly Glu 10 Tyr Asp Glu Trp Gly Asn Gln Leu Asn Glu Glu Asn Pro His His Leu 20 25 His Gln Pro Tyr Arg Leu Pro Gly Gln Gln Tyr Asp Lys Glu Ser Gly. 40 Leu Tyr Tyr Asn Arg Asn Arg Tyr Tyr Asp Pro Leu Gln Gly Arg Tyr 55 Ile Thr Gln Asp Pro Ile Gly Leu Glu Gly Gly Trp Ser Leu Tyr Ala 70 75 Tyr Pro Leu Asn Pro Val Asn Gly Ile Asp Pro Leu Gly Leu Ser Pro 85 90 Ala Asp Val Ala Leu Ile Arg Arg Lys Asp Gln Leu Asn His Gln Arg 100 105 . 110 Ala Trp Asp Ile Leu Ser Asp Thr Tyr Glu Asp Met Lys Arg Leu Asn 120 Leu Gly Gly Thr Asp Gln Phe Phe His Cys Met Ala Phe Cys Arg Val

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130
                        135
Ser Lys Leu Asn Asp Ala Gly Val Ser Arg Ser Ala Lys Gly Leu Gly
                    150
Tyr Glu Lys Glu Ile Arg Asp Tyr Gly Leu Asn Leu Phe Gly Met Tyr
                165
                                    170
Gly Arg Lys Val Lys Leu Ser His Ser Glu Met Ile Glu Asp Asn Lys
            180
                                185
Lys Asp Leu Ala Val Asn Asp His Gly Leu Thr Cys Pro Ser Thr Thr
                            200
Asp Cys Ser Asp Arg Cys Ser Asp Tyr Ile Asn Pro Glu His Lys Lys
    210
                        215
Thr Ile Lys Ala Leu Gln Asp Ala Gly Tyr Leu Lys
                    230
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<210> 343 <21i> 86 <212> PRT <213> E. Coli

<400> 343

 Met
 Leu
 Ala
 Ile
 Ser
 Ser
 Asn
 Leu
 Ser
 Lys
 Met
 Ile
 Ile
 Phe
 Ile
 Phe
 Ile
 Phe
 Ile
 Phe
 Ile
 Phe
 Ile
 Ile
 Ile
 Ile
 Ile
 Val
 Val
 Leu
 Cys
 Val
 Ile
 Thr
 Tyr
 Leu
 Tyr
 Leu
 Tyr
 Leu
 Tyr
 Leu
 Tyr
 Met
 Ala
 Ala</th

<210> 344 <211> 63 <212> PRT <213> E. Coli

<400> 344

 Met Arg Ala Arg Glu Gln Val Ala Lys Ile Val Ser Lys Asn Asp Pro 1
 5
 10
 15

 Asp Thr Lys Lys Val Trp Cys Lys Tyr Gly Lys Ile Pro Gly Gln Gly 20
 25
 30

 Asp Gly Val Asn Leu Phe Phe Val Gly Glu Ile Asn Val Thr His Tyr 35
 40
 45

 Phe Ile Thr Asn Ile Gly Ala Gly Leu Pro Asp Ala Cys Ala Glu 50
 55

<210> 345 <211> 167 <212> PRT <213> E. Coli

<400> 345

Met Pro Gly Asn Ser Pro His Tyr Gly Arg Trp Pro Gln His Asp Phe 1 5 10 15

Thr Ser Leu Lys Lys Leu Arg Pro Gln Ser Val Thr Ser Arg Ile Gln 25 Pro Gly Ser Asp Val Ile Val Cys Ala Glu Met Asp Glu Gln Trp Gly Tyr Val Gly Ala Lys Ser Arg Gln Arg Trp Leu Phe Tyr Ala Tyr Asp Ser Leu Arg Lys Thr Val Val Ala His Val Phe Gly Glu Arg Thr Met 75 Ala Thr Leu Gly Arg Leu Met Ser Leu Leu Ser Pro Phe Asp Val Val 90 Ile Trp Met Thr Asp Gly Trp Pro Leu Tyr Glu Ser Arg Leu Lys Gly 105 Lys Leu His Val Ile Ser Lys Arg Tyr Thr Gln Arg Ile Glu Arg His 120 Asn Leu Asn Leu Arg Gln His Leu Ala Arg Leu Gly Arg Lys Ser Leu 135 140 Ser Phe Ser Lys Ser Val Glu Leu His Asp Lys Val Ile Gly His Tyr 150 155 Leu Asn Ile Lys His Tyr Gln 165

<210> 346 <211> 91 <212> PRT <213> E. Coli

<210> 347 <211> 138 <212> PRT <213> E. Coli

85

 400>
 347

 Met Met Thr Lys Thr Gln Ile Asn Lys Leu Ile Lys Met Met Asn Asp 1
 5
 10
 15

 Leu Asp Tyr Pro Phe Glu Ala Pro Leu Lys Glu Ser Phe Ile Glu Ser 20
 25
 30

 Ile Ile Gln Ile Glu Phe Asn Ser Asn Ser Thr Asn Cys Leu Glu Lys 35
 40
 45

 Leu Cys Asn Glu Val Ser Ile Leu Phe Lys Asn Gln Pro Asp Tyr Leu 50
 55
 60

 Thr Phe Leu Arg Ala Met Asp Gly Phe Glu Val Asn Gly Leu Arg Leu 65
 70
 75
 80

 Phe Ser Leu Ser Ile Pro Glu Pro Ser Val Lys Asn Leu Phe Ala Val 85
 90
 95

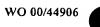
Asn Glu Phe Tyr Arg Asn Asn Asp Asp Phe Ile Asn Pro Asp Leu Gln
100 105 110

Glu Arg Leu Val Ile Gly Asp Tyr Ser Ile Ser Ile Phe Thr Tyr Asp
115 120 125

Ile Lys Gly Asp Ala Ala Asn Leu Leu Ile

<210> 348 <211> 392 <212> PRT <213> E. Coli

<400> 348 Met Ser Asn Ile Val Tyr Leu Thr Val Thr Gly Glu Gln Gln Gly Ser 10 Ile Ser Ala Gly Cys Gly Thr Ser Glu Ser Thr Gly Asn Arg Trp Gln 25 Ser Gly His Glu Asp Glu Ile Phe Thr Phe Ser Leu Leu Asn Asn Ile 40 Asn Asn Thr Gly Leu Gly Ser Gln Phe His Gly Ile Thr Phe Cys Lys 55 Leu Ile Asp Lys Ser Thr Pro Leu Phe Ile Asn Ser Ile Asn Asn Asn 70 Glu Gln Leu Phe Met Gly Phe Asp Phe Tyr Arg Ile Asn Arg Phe Gly 85 Arg Leu Glu Lys Tyr Tyr Ile Gln Leu Arg Gly Ala Phe Leu Ser 105 Ala Ile His His Gln Ile Ile Glu Asn Gln Leu Asp Thr Glu Thr Ile 120 -125 Thr Ile Ser Tyr Glu Phe Ile Leu Cys Gln His Leu Ile Ala Asn Thr ·..' 135 Glu Phe Ser Tyr Leu Ala Leu Pro Glu Asn Tyr Asn Arg Leu Phe Leu 150 155 Pro Asn Ser Lys Asn Gln Thr Asn Asn Arg Phe Lys Thr Leu Asn Ser 165 170 Lys Ala Ile Gly Arg Leu Leu Ala Ala Gly Gly Val Tyr Asn Gly Asn 185 Ile Glu Gly Phe Arg Asp Thr Ala Glu Lys Leu Gly Gly Asp Ala Ile 200 Lys Gly Tyr Asp Gln Ile Leu Asn Glu Lys Thr Ala Gly Ile Ala Ile 215 Ala Thr Ala Ser Ile Leu Leu Thr Lys Arg Ser Asn Val Asp Thr Tyr 230 235 Thr Glu Ile Asn Ser Tyr Leu Gly Lys Leu Arg Gly Gln Gln Lys Leu 245 250 Leu Asp Gly Ile Asp Ile Ile Glu Ile Ile Tyr Ile Lys Arg Pro Ser 260 265 Lys Asp Leu Ala Asn Leu Arg Lys Glu Phe Asn Lys Thr Val Arg Lys 280 Asn Phe Leu Ile Lys Leu Ala Lys Thr Ser Glu Ala Ser Gly Arg Phe 295 Asn Ala Glu Asp Leu Leu Arg Met Arg Lys Gly Asn Val Pro Leu Asn 310 315 Tyr Asn Val His His Lys Leu Ser Leu Asp Asp Gly Gly Thr Asn Asp 325 330 Phe Glu Asn Leu Val Leu Ile Glu Asn Glu Pro Tyr His Lys Val Phe 340 345 Thr Asn Met Gln Ser Arg Ile Ala Lys Gly Ile Leu Val Gly Glu Ser 360 Lys Ile Thr Pro Trp Ala Ile Pro Ser Gly Ser Ile Tyr Pro Pro Met



370 375 Lys Asn Ile Met Asp His Thr Lys 385 390 380

<210> 349 <211> 221 <212> PRT <213> E. Coli

<400> 349 Met Val Leu Ala Leu Asn Tyr Asn Met His Gly Val Asn Ile Arg Ser Glu Asn Ala Ala Lys Pro His Thr Met Pro Ser Arg Tyr Leu Cys Glu 20 Tyr Ile Arg Ser Ile Glu Lys Asn Gly His Ala Leu Asp Phe Gly Cys 40 Gly Lys Leu Arg Tyr Ser Asp Glu Leu Ile Ser Lys Phe Asp Glu Val 50 55 Thr Phe Leu Asp Ser Lys Arg Gln Leu Glu Arg Glu Gln Ile Ile Arg 70 Gly Ile Lys Thr Lys Ile Ile Asp Tyr Val Pro Arg Tyr Tyr Lys Asn 85 90 Ala Asn Thr Val Ala Phe Glu Asp Val Asp Lys Ile Ile Gly Gly Tyr . 100 105 Asp Phe Ile Leu Cys Ser Asn Val Leu Ser Ala Val Pro Cys Arg Asp 115 120 Thr Ile Asp Lys Ile Val Leu Ser Ile Lys Arg Leu Leu Lys Ser Gly 135 140 Gly Glu Thr Leu Ile Val Asn Gln Tyr Lys Ser Ser Tyr Phe Lys Lys 150 155 Tyr Glu Thr Gly Arg Lys His Leu Tyr Gly Tyr Ile Tyr Lys Asn Ser 170 Lys Ser Val Ser Tyr Tyr Gly Leu Leu Asp Glu Leu Ala Val Gln Glu 185 Ile Cys Ser Ser His Gly Leu Glu Ile Leu Lys Ser Trp Ser Lys Ala 200 Gly Ser Ser Tyr Val Thr Val Gly Ser Cys Asn Ala Ile 210 215

<210> 350 <211> 234 <212> PRT <213> E. Coli

100 105 Gly Val Asn Arg Lys Leu Asn Lys Asp Glu Ala His Asn Val Met Ser 1.20 Asn Leu Tyr Tyr Pro Glu Val Arg Lys Ile Glu Asp Lys His Tyr Ile 135 -Glu Leu Phe Trp Asn Lys Val Tyr Tyr Phe Trp Ile Phe Phe Glu Phe 150 155 Ser Ile Ile Ala Leu Ile Ser Phe Leu Ile Ile Phe Phe Cys Lys Gln 170 Met Asp Ile Phe His Val Glu Gly Ser Leu Leu Ser Leu Phe Phe 180 185 · Val Ile Leu Ser Phe Ser Val Ser Gly Ile Ile Phe Ala Leu Thr Val 200 Lys Pro Arg Thr Glu Ser Gln Val Gly Lys Ile Pro Asp Asp Lys Ile 215 Lys Glu Phe Phe Thr Lys Asn Asn Ile Asn 230

<210> 351 <211> 94 <212> PRT <213> E. Coli

<400> 351 Met Phe Thr Ile Asn Ala Glu Val Arg Lys Glu Gln Gly Lys Gly Ala Ser Arg Arg Leu Arg Ala Ala Asn Lys Phe Pro Ala Ile Ile Tyr Gly Gly Lys Glu Ala Pro Leu Ala Ile Glu Leu Asp His Asp Lys Val Met 40 Asn Met Gln Ala Lys Ala Glu Phe Tyr Ser Glu Val Leu Thr Ile Val Val Asp Gly Lys Glu Ile Lys Val Lys Ala Gln Asp Val Gln Arg His 70 Pro Tyr Lys Pro Lys Leu Gln His Ile Asp Phe Val Arg Ala 85

90

<210> 352 <211> 658 <212> PRT <213> E. Coli

<400> 352 Met Val Leu Phe Tyr Arg Ala His Trp Arg Asp Tyr Lys Asn Asp Gln 10 Val Arg Ile Met Met Asn Leu Thr Thr Leu Thr His Arg Asp Ala Leu 25 Cys Leu Asn Ala Arg Phe Thr Ser Arg Glu Glu Ala Ile His Ala Leu 40 Thr Gln Arg Leu Ala Ala Leu Gly Lys Ile Ser Ser Thr Glu Gln Phe 55 Leu Glu Glu Val Tyr Arg Arg Glu Ser Leu Gly Pro Thr Ala Leu Gly Glu Gly Leu Ala Val Pro His Gly Lys Thr Ala Ala Val Lys Glu Ala 85 90 Ala Phe Ala Val Ala Thr Leu Ser Glu Pro Leu Gln Trp Glu Gly Val 105 Asp Gly Pro Glu Ala Val Asp Leu Val Val Leu Leu Ala Ile Pro Pro

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	130	J				135	Ö				140	Ala	Leu		Thr
145	)				150	)				155	Gln	Ser			Thr 160
				165	>				170	)				175	Pro
Ser	Ala	a Sei	Phe 180	e Ser	Asn	Ala	Pro	Thr 185	Ile	val	Cys	Val	Thr 190	Ala	Cys
		195	)				200	)				-205	Leu	Glu	Lys
	210	) .				215	)				220	•			Ala
225		·			230					235					Thr 240
				245	•				250					255	Phe
			260	)				265					270		His
		2/5					280					285			Glu
	290					295				Val	300				
305					310					Ser 315					320
				325					330	Ala				335	
			340					345		Glu			350		
		355					360			Gly		365			
	3/0					375				Ala	380				
385					390					Asn 395					400
				405					410	Ala	•			415	
			420					425		Lys			430		
		435					440			Leu		445			
	450					455				Trp	460				
465					4/0					Asn 475 Leu					480
				485		•			490	Met				495	
			500					505		Met			510		
		212					520			Leu		525			
	530					535				Gly	540				
545			•		550					555 Pro					5.60
				565			•		570	Ala			•	575	
			280					585		Ile			590		
		595					600		y			605	u	File	nen

 Leu His Asp Asn Gly Ala Gly Gly Val Met Ala Ala Ile Gly Trp Phe

 610
 615
 620

 Gly Ala Ala Leu Val Gly Ala Ala Ile Ser Thr Ala Ile Leu Leu Met
 625
 630
 635
 640

 Trp Arg Arg His Ala Val Lys His Gly Asn Tyr Leu Thr Asp Gly Val
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 Met Pro
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 655
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<210> 353 <211> 877 <212> PRT <213> E. Coli

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			· ·												
		555	5				360	)				365	5		
Arç	Phe 370	Tyr	Met	Arc	J Lys	375	Ala		Asr	n Met	Prc 380	Glr	Ser	Asp	Ala
Asp 385	Lys	Leu	ı Val	Leu	Phe 390	Asn	Leu	Met	Pṛc	Trp 395	Pro	. Arç	Glu	Glu	Val 400
Ile	. Asi	Thr	Thr	Val 405	Arg	Leu	Arg	Ala	Ser 410	Gln		Asn	Leu	Arg 415	Asp
Asp	Arg	. Gl	Gln 420	Pro	Val	Pro	Tyr	Phe 425	Il€	Arg	His	Ala	Arg	Glu	Ile
Asp	Pro	Gly 435	Leu	Ile	Asp	Arg	Gln 440	Ile		His	Tyr	Gly 445	Asn	Tyr	Asp
	450	,				455				Gln	460	Val	Pro		
465			· .		470					475	Pro	Gly	•		Ile 480
•				485					490	Glu				495	Gln
			500					505	•	Leu			510	Asp	
		212					520			Glu		525			
	530					535				Glu	540				
545					550					Ile 555					560
				565					.570					575	
			580					585		Val			590		
		595					600			Asp		605			
	610					615				Val	620				
625					630					Gln 635					640
	•			645					650	Trp				655	
		•	660					665		Leu Ser			670		
		6/5					680			Phe		685			
	690					695				Leu	700				
703					110					715 Asp					720
				725					730	Tyr				735	
			740					745		Trp			750		
		155					760			Lys		765			
	770					775					780				
185					790					795 Asp					800
Leu				805					810					815	
Ala			820					825					830		
		835	,		-	-	840	- , _				845		.σp	o + u

His Ile Thr Thr Glu Glu Asn Gln Gly Ser Asn Leu Ser Gly Pro Phe 850 855 860 Leu Pro Gly Gln Ser Arg Thr Phe Ser Tyr Arg Leu Ala 865 870 875

<210> 354 <211> 523 <212> PRT <213> E. Coli

<400> 354 . Met Met Leu Asp Ile Val Glu Leu Ser Arg Leu Gln Phe Ala Leu Thr 10 Ala Met Tyr His Phe Leu Phe Val Pro Leu Thr Leu Gly Met Ala Phe Leu Leu Ala Ile Met Glu Thr Val Tyr Val Leu Ser Gly Lys Gln Ile Tyr Lys Asp Met Thr Lys Phe Trp Gly Lys Leu Phe Gly Ile Asn Phe Ala Leu Gly Val Ala Thr Gly Leu Thr Met Glu Phe Gln Phe Gly Thr 70 75 Asn Trp Ser Tyr Tyr Ser His Tyr Val Gly Asp Ile Phe Gly Ala Pro Leu Ala Ile Glu Gly Leu Met Ala Phe Phe Leu Glu Ser Thr Phe Val 105 Gly Leu Phe Phe Gly Trp Asp Arg Leu Gly Lys Val Gln His Met 120 125 Cys Val Thr Trp Leu Val Ala Leu Gly Ser Asn Leu Ser Ala Leu Trp 135 140 Ile Leu Val Ala Asn Gly Trp Met Gln Asn Pro Ile Ala Ser Asp Phe .<sub>...</sub>150 . 155 Asn Phe Glu Thr Met Arg Met Glu Met Val Ser Phe Ser Glu Leu Val 165 170 Leu Asn Pro Val Ala Gln Val Lys Phe Val His Thr Val Ala Ser Gly 185 Tyr Val Thr Gly Ala Met Phe Ile Leu Gly Ile Ser Ala Trp Tyr Met 200 205 Leu Lys Gly Arg Asp Phe Ala Phe Ala Lys Arg Ser Phe Ala Ile Ala 215 220 Ala Ser Phe Gly Met Ala Ala Val Leu Ser Val Ile Val Leu Gly Asp 230 235 Glu Ser Gly Tyr Glu Met Gly Asp Val Gln Lys Thr Lys Leu Ala Ala 245 250 Ile Glu Ala Glu Trp Glu Thr Gln Pro Ala Pro Ala Ala Phe Thr Leu 260 265 Phe Gly Ile Pro Asp Gln Glu Glu Glu Thr Asn Lys Phe Ala Ile Gln 280 285 Ile Pro Tyr Ala Leu Gly Ile Ile Ala Thr Arg Ser Val Asp Thr Pro 295 300 Val Ile Gly Leu Lys Glu Leu Met Val Gln His Glu Glu Arg Ile Arg 310 315 Asn Gly Met Lys Ala Tyr Ser Leu Leu Glu Gln Leu Arg Ser Gly Ser 325 330 Thr Asp Gln Ala Val Arg Asp Gln Phe Asn Ser Met Lys Lys Asp Leu 345 Gly Tyr Gly Leu Leu Leu Lys Arg Tyr Thr Pro Asn Val Ala Asp Ala 360 Thr Glu Ala Gln Ile Gln Gln Ala Thr Lys Asp Ser Ile Pro Arg Val 375

Ala Pro Leu Tyr Phe Ala Phe Arg Ile Met Val Ala Cys Gly Phe Leu

390 395 Leu Leu Ala Ile Ile Ala Leu Ser Phe Trp Ser Val Ile Arg Asn Arg 405 410 Ile Gly Glu Lys Lys Trp Leu Leu Arg Ala Ala Leu Tyr Gly Ile Pro 425 Leu Pro Trp Ile Ala Val Glu Ala Gly Trp Phe Val Ala Glu Tyr Gly . 435 440 Arg Gln Pro Trp Ala Ile Gly Glu Val Leu Pro Thr Ala Val Ala Asn 455 460 Ser Ser Leu Thr Ala Gly Asp Leu Ile Phe Ser Met Val Leu Ile Cys 470 475 Gly Leu Tyr Thr Leu Phe Leu Val Ala Glu Leu Phe Leu Met Phe Lys 485 490 Phe Ala Arg Leu Gly Pro Ser Ser Leu Lys Thr Gly Arg Tyr His Phe 500 505 Glu Gln Ser Ser Thr Thr Thr Gln Pro Ala Arg 520

<210> 355 <211> 379 <212> PRT <213> E. Coli

<400> 355 Met Ile Asp Tyr Glu Val Leu Arg Phe Ile Trp Trp Leu Leu Val Gly 10 Val Leu Leu Ile Gly Phe Ala Val Thr Asp Gly Phe Asp Met Gly Val 25 Gly Met Leu Thr Arg Phe Leu Gly Arg Asn Asp Thr Glu Arg Arg Ile 40 Met Ile Asn Ser Ile Ala Pro His Trp Asp Gly Asn Gln Val Trp Leu 55 . Ile Thr Ala Gly Gly Ala Leu Phe Ala Ala Trp Pro Met Val Tyr Ala 70 75 Ala Ala Phe Ser Gly Phe Tyr Val Ala Met Ile Leu Val Leu Ala Ser 85 90 Leu Phe Phe Arg Pro Val Gly Phe Asp Tyr Arg Ser Lys Ile Glu Glu 105 Thr Arg Trp Arg Asn Met Trp Asp Trp Gly Ile Phe Ile Gly Ser Phe 115 120 Val Pro Pro Leu Val Ile Gly Val Ala Phe Gly Asn Leu Leu Gln Gly 135 Val Pro Phe Asn Val Asp Glu Tyr Leu Arg Leu Tyr Tyr Thr Gly Asn 150 155 Phe Phe Gln Leu Leu Asn Pro Phe Gly Leu Leu Ala Gly Val Val Ser . 165 170 Val Gly Met Ile Ile Thr Gln Gly Ala Thr Tyr Leu Gln Met Arg Thr 185 Val Gly Glu Leu His Leu Arg Thr Arg Ala Thr Ala Gln Val Ala Ala 200 205 Leu Val Thr Leu Val Cys Phe Ala Leu Ala Gly Val Trp Val Met Tyr 215 220 Gly Ile Asp Gly Tyr Val Val Lys Ser Thr Met Asp His Tyr Ala Ala 230 235 Ser Asn Pro Leu Asn Lys Glu Val Val Arg Glu Ala Gly Ala Trp Leu 250 Val Asn Phe Asn Asn Thr Pro Ile Leu Trp Ala Ile Pro Ala Leu Gly 265 Val Val Leu Pro Leu Leu Thr Ile Leu Thr Ala Arg Met Asp Lys Ala 280

<210> 356 <211> 456 <212> PRT <213> E. Coli

<400> 356

Met Glu Leu Ser Ser Leu Thr Ala Val Ser Pro Val Asp Gly Arg Tyr 10 Gly Asp Lys Val Ser Ala Leu Arg Gly Ile Phe Ser Glu Tyr Gly Leu 25 Leu Lys Phe Arg Val Gln Val Glu Val Arg Trp Leu Gln Lys Leu Ala 40 Ala His Ala Ala Ile Lys Glu Val Pro Ala Phe Ala Ala Asp Ala Ile 55 Gly Tyr Leu Asp Ala Ile Val Ala Ser Phe Ser Glu Glu Asp Ala Ala 70 75 Arg Ile Lys Thr Ile Glu Arg Thr Thr Asn His Asp Val Lys Ala Val 85... 90 Glu Tyr Phe Leu Lys Glu Lys Val Ala Glu Ile Pro Glu Leu His Ala 105 Val Ser Glu Phe Ile His Phe Ala Cys Thr Ser Glu Asp Ile Asn Asn 120 Leu Ser His Ala Leu Met Leu Lys Thr Ala Arg Asp Glu Val Ile Leu 135 140 Pro Tyr Trp Arg Gln Leu Ile Asp Gly Ile Lys Asp Leu Ala Val Gln 150 155 Tyr Arg Asp Ile Pro Leu Leu Ser Arg Thr His Gly Gln Pro Ala Thr 165 170 Pro Ser Thr Ile Gly Lys Glu Met Ala Asn Val Ala Tyr Arg Met Glu 185 Arg Gln Tyr Arg Gln Leu Asn Gln Val Glu Ile Leu Gly Lys Ile Asn 200 Gly Ala Val Gly Asn Tyr Asn Ala His Ile Ala Ala Tyr Pro Glu Val 215 220 Asp Trp His Gln Phe Ser Glu Glu Phe Val Thr Ser Leu Gly Ile Gln 230 235 Trp Asn Pro Tyr Thr Thr Gln Ile Glu Pro His Asp Tyr Ile Ala Glu 245 250 Leu Phe Asp Cys Val Ala Arg Phe Asn Thr Ile Leu Ile Asp Phe Asp 260 265 Arg Asp Val Trp Gly Tyr Ile Ala Leu Asn His Phe Lys Gln Lys Thr - 280 Ile Ala Gly Glu Ile Gly Ser Ser Thr Met Pro His Lys Val Asn Pro 295 300 Ile Asp Phe Glu Asn Ser Glu Gly Asn Leu Gly Leu Ser Asn Ala Val 310 315 Leu Gln His Leu Ala Ser Lys Leu Pro Val Ser Arg Trp Gln Arg Asp

```
325
Leu Thr Asp Ser Thr Val Leu Arg Asn Leu Gly Val Gly Ile Gly Tyr
            340
                                345
Ala Leu Ile Ala Tyr Gln Ser Thr Leu Lys Gly Val Ser Lys Leu Glu
                            360
Val Asn Arg Asp His Leu Leu Asp Glu Leu Asp His Asn Trp Glu Val
                        375
                                            380
Leu Ala Glu Pro Ile Gln Thr Val Met Arg Arg Tyr Gly Ile Glu Lys
                    390
                                        395
Pro Tyr Glu Lys Leu Lys Glu Leu Thr Arg Gly Lys Arg Val Asp Ala
                                    410
Glu Gly Met Lys Gln Phe Ile Asp Gly Leu Ala Leu Pro Glu Glu Glu
            420
                                425
Lys Ala Arg Leu Lys Ala Met Thr Pro Ala Asn Tyr Ile Gly Arg Ala
                            440
Ile Thr Met Val Asp Glu Leu Lys
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<210> 357 <211> 61 <212> PRT <213> E. Coli

<400> 357

 Met Leu Ile Leu Thr Arg Arg Val Gly Glu Thr Leu Met Ile Gly Asp

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 5
 10
 15

 Glu Val Thr Val Thr Val Leu Gly Val Lys Gly Asn Gln Val Arg Ile
 20
 25
 30

 Gly Val Asn Ala Pro Lys Glu Val Ser Val His Arg Glu Glu Ile Tyr
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 40
 45

 Gln Arg Ile Gln Ala Glu Lys Ser Gln Gln Ser Ser Tyr
 50
 55
 60

<210> 358 <211> 83 <212> RNA <213> E. Coli

<400> 358

gcauccgggg uucgaaucce cgccucaccg cca ggaguaugcg gucaaaagcu

60 83

<210> 359 <211> 200 <212> PRT <213> E. Coli

<400> 359

 Meu
 Lys
 Asn
 Lys
 Ala
 Asp
 Asn
 Lys
 Lys
 Arg
 Asn
 Phe
 Leu
 Thr
 His
 Ser

 Glu
 Ile
 Glu
 Ser
 Leu
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 Ala
 Asn
 Thr
 Gly
 Pro
 His
 Ala
 Ala

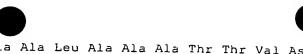
 Arg
 Asn
 Tyr
 Cys
 Leu
 Thr
 Leu
 Leu
 Cys
 Phe
 Ile
 His
 Gly
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 Arg
 Ala
 Lys
 Asp
 Ile
 Asp
 Leu
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 Ala
 Lys
 Ala
 Ala
 Ala
 Asp
 Ile
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 Ala
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 Ala

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Leu Leu Asn Lys Glu Val Gln Ala Leu Lys Asn Trp Leu Ser Ile Arg
                                   90
Thr Ser Tyr Pro His Ala Glu Ser Glu Trp Val Phe Leu Ser Arg Lys
                               105
Gly Asn Pro Leu Ser Arg Gln Gln Phe Tyr His Ile Ile Ser Thr Ser
                           120
                                               125
Gly Gly Asn Ala Gly Leu Ser Leu Glu Ile His Pro His Met Leu Arg
                       135
                                           140
His Ser Cys Gly Phe Ala Leu Ala Asn Met Gly Ile Asp Thr Arg Leu
                   150
                                      155 .
Ile Gln Asp Tyr Leu Gly His Arg Asn Ile Arg His Thr Val Trp Tyr
               165 .
                                  170
Thr Ala Ser Asn Ala Gly Arg Phe Tyr Gly Ile Trp Asp Arg Ala Arg
                           185
           180 .
Gly Arg Gln Arg His Ala Val Leu
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<210> 360 <211> 198 <212> PRT <213> E. Coli

<400> 360 Met Ser Lys Arg Arg Tyr Leu Thr Gly Lys Glu Val Gln Ala Met Met 10 Gln Ala Val Cys Tyr Gly Ala Thr Gly Ala Arg Asp Tyr Cys Leu Ile 20 Leu Leu Ala Tyr Arg His Gly Met Arg Ile Ser Glu Leu Leu Asp Leu 40 His Tyr Gln Asp Leu Asp Leu Asn Glu Gly Arg Ile Asn Ile Arg Arg ... · 55 Leu Lys Asn Gly Phe Ser Thr Val His Pro Leu Arg Phe Asp Glu Arg 70 75 Glu Ala Val Glu Arg Trp Thr Gln Glu Arg Ala Asn Trp Lys Gly Ala 8.5 90 Asp Arg Thr Asp Ala Ile Phe Ile Ser Arg Arg Gly Ser Arg Leu Ser 100 105 Arg Gln Gln Ala Tyr Arg Ile Ile Arg Asp Ala Gly Ile Glu Ala Gly 115 120 Thr Val Thr Gln Thr His Pro His Met Leu Arg His Ala Cys Gly Tyr 135 Glu Leu Ala Glu Arg Gly Ala Asp Thr Arg Leu Ile Gln Asp Tyr Leu. 150 155 Gly His Arg Asn Ile Arg His Thr Val Arg Tyr Thr Ala Ser Asn Ala 165 , .170 Ala Arg Phe Ala Gly Leu Trp Glu Arg Asn Asn Leu Ile Asn Glu Lys 180 185 Leu Lys Arg Glu Glu Val 195

<210> 361 <211> 182 <212> PRT <213> E. Coli



Ser Ser Thr Ala Ala Leu Ala Ala Ala Thr Thr Val Asn Gly Gly Thr Val His Phe Lys Gly Glu Val Val Asn Ala Ala Cys Ala Val Asp Ala 40 Gly Ser Val Asp Gln Thr Val Gln Leu Gly Gln Val Arg Thr Ala Ser Leu Ala Gln Glu Gly Ala Thr Ser Ser Ala Val Gly Phe Asn Ile Gln 75 Leu Asn Asp Cys Asp Thr Asn Val Ala Ser Lys Ala Ala Val Ala Phe 90 Leu Gly Thr Ala Ile Asp Ala Gly His Thr Asn Val Leu Ala Leu Gln . 105 Ser Ser Ala Ala Gly Ser Ala Thr Asn Val Gly Val Gln Ile Leu Asp 120 Arg Thr Gly Ala Ala Leu Thr Leu Asp Gly Ala Thr Phe Ser Ser Glu 135 Thr Thr Leu Asn Asn Gly Thr Asn Thr Ile Pro Phe Gln Ala Arg Tyr 150 155 Phe Ala Thr Gly Ala Ala Thr Pro Gly Ala Ala Asn Ala Asp Ala Thr 165 170 Phe Lys Val Gln Tyr Gln . 180

<210> 362 <211> 215 <212> PRT <213> E. Coli

<400> 362

Met Leu Leu Met Arg Met Arg Pro Ser Arg Phe Ser Ile Asn Asn Leu 5. 10 Pro Arg Phe Arg Asp Val Ile Thr Gly Arg Asp Ala His Pro Cys Ala 20 25 Ile Lys Ile Thr Met Lys Arg Lys Arg Leu Phe Leu Leu Ala Ser Leu 40 Leu Pro Met Phe Ala Leu Ala Gly Asn Lys Trp Asn Thr Thr Leu Pro 55 Gly Gly Asn Met Gln Phe Gln Gly Val Ile Ile Ala Glu Thr Cys Arg 70 75 Ile Glu Ala Gly Asp Lys Gln Met Thr Val Asn Met Gly Gln Ile Ser 90 85 Ser Asn Arg Phe His Ala Val Gly Glu Asp Ser Ala Pro Val Pro Phe 100 105 Val Ile His Leu Arg Glu Cys Ser Thr Val Val Ser Glu Arg Val Gly 120 Val Ala Phe His Gly Val Ala Asp Gly Lys Asn Pro Asp Val Leu Ser 135 140 Val Gly Glu Gly Pro Gly Ile Ala Thr Asn Ile Gly Val Ala Leu Phe 150 155 Asp Asp Glu Gly Asn Leu Val Pro Ile Asn Arg Pro Pro Ala Asn Trp 165 170 Lys Arg Leu Tyr Ser Gly Ser Thr Ser Leu His Phe Ile Ala Lys Tyr 185 Arg Ala Thr Gly Arg Arg Val Thr Gly Gly Ile Ala Asn Ala Gln Ala - 200 Trp Phe Ser Leu Thr Tyr Gln

<210> 363 <211> 241 <212> PRT <213> E. Coli

<400> 363 Met Ser Asn Lys Asn Val Asn Val Arg Lys Ser Gln Glu Ile Thr Phe 10 . Cys Leu Leu Ala Gly Ile Leu Met Phe Met Ala Met Met Val Ala Gly 25 Arg Ala Glu Ala Gly Val Ala Leu Gly Ala Thr Arg Val Ile Tyr Pro 40 Ala Gly Gln Lys Gln Glu Gln Leu Ala Val Thr Asn Asn Asp Glu Asn 55 Ser Thr Tyr Leu Ile Gln Ser Trp Val Glu Asn Ala Asp Gly Val Lys 70 Asp Gly Arg Phe Ile Val Thr Pro Pro Leu Phe Ala Met Lys Gly Lys 85 90 Lys Glu Asn Thr Leu Arg Ile Leu Asp Ala Thr Asn Asn Gln Leu Pro 100 105 Gln Asp Arg Glu Ser Leu Phe Trp Met Asn Val Lys Ala Ile Pro Ser 115 120 Met Asp Lys Ser Lys Leu Thr Glu Asn Thr Leu Gln Leu Ala Ile Ile 135 Ser Arg Ile Lys Leu Tyr Tyr Arg Pro Ala Lys Leu Ala Leu Pro Pro 150 155 Asp Gln Ala Ala Glu Lys Leu Arg Phe Arg Arg Ser Ala Asn Ser Leu 165 170 Thr Leu Ile Asn Pro Thr Pro Tyr Tyr Leu Thr Val Thr Glu Leu Asn 180 185 Ala Gly Thr Arg Val Leu Glu Asn Ala Leu Val Pro Pro Met Gly Glu 200 Ser Thr Val Lys Leu Pro Ser Asp Ala Gly Ser Asn Ile Thr Tyr Arg 215 220 Thr Ile Asn Asp Tyr Gly Ala Leu Thr Pro Lys Met Thr Gly Val Met 225 230 Glu

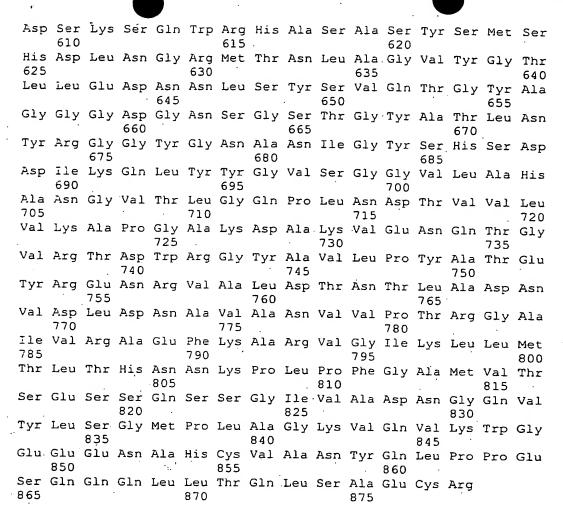
<210> 364 <211> 878 <212> PRT <213> E. Coli

<400> 364

 Met
 Ser
 Tyr
 Leu
 Asn
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 Arg
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 Tyr
 Gln
 Arg
 Asn
 Thr
 Gln
 Cys
 Leu
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 Thr
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			115	5		•		120	)				125	,		
		130	)				135	Val	Pro			140	Met	. Val		Asp
	Ala 145	Thr	Ala	a His	Let	Asp 150		. Gly	Gln	Gln	Arg 155		Asn	Leu	Thr	Ile 160
					165	5				170					175	Glu
	Leu	Trp	Asp	Pro 180	Gly	, Ile	. Asn	Ala	Gly 185		Leu	Asn	Tyr	Asn 190		Ser
			195	<b>.</b>				200					205	Tyr	Ala	Tyr
		210	)				215					220				Asp
	225					230					235			-		Lys 240
	••				245					250					255	Pro
				260		Thr			265					270	_	
			275		•	Phe		280					285			
		290				Gln	295					300			_	
	305			•		Gln 310					315					320
					325	Pro				330					335	_
	•			340		Gly			345					350		_
			355			Phe		360		_			365			
		370				Thr	375					380				_
	385		•			Gln 390					395					400
					405	Ala				410					415	
				420		Ala			425					430		_
			435			Ser		440					445		•	
		450				His	455					460				
4	465					Ser 470					475					480
					485	Tyr				490					495	_
				500		Ile			505					510		_
			515			Tyr		520					525			_
		530				Thr	535					540				_
5	45					Gln 550					555					560
					565	Leu Thr				570					575	
				580		Val			585					590		
-		,	595		- 1.011	- 41	- 111	600		- 11C	SET		605 <sub>.</sub>	Leu	игд	ser



<210> 365 <211> 176 <212> PRT <213> E. Coli

<400> 365

Met Arg Asn Lys Pro Phe Tyr Leu Leu Cys Ala Phe Leu Trp Leu Ala 10 Val Ser His Ala Leu Ala Ala Asp Ser Thr Ile Thr Ile Arg Gly Tyr 25 Val Arg Asp Asn Gly Cys Ser Val Ala Ala Glu Ser Thr Asn Phe Thr 40 Val Asp Leu Met Glu Asn Ala Ala Lys Gln Phe Asn Asn Ile Gly Ala 55 Thr Thr Pro Val Val Pro Phe Arg Ile Leu Leu Ser Pro Cys Gly Asn 70 75 Ala Val Ser Ala Val Lys Val Gly Phe Thr Gly Val Ala Asp Ser His 85 90 Asn Ala Asn Leu Leu Ala Leu Glu Asn Thr Val Ser Ala Ala Ser Gly 105 Leu Gly Ile Gln Leu Leu Asn Glu Gln Gln Asn Gln Ile Pro Leu Asn 115 120 Ala Pro Ser Ser Ala Leu Ser Trp Thr Thr Leu Thr Pro Gly Lys Pro 135 140 Asn Thr Leu Asn Phe Tyr Ala Arg Leu Met Ala Thr Gln Val Pro Val

र करते अवस्थाना हुने हैं। द

145 150 155 160 Thr Ala Gly His Ile Asn Ala Thr Ala Thr Phe Thr Leu Glu Tyr Gln 165 170 175

<210> 366 <211> 167 <212> PRT <213> E. Coli

<400> 366 Met Lys Trp Cys Lys Arg Gly Tyr Val Leu Ala Ala Ile Leu Ala Leu 10 Ala Ser Ala Thr Ile Gln Ala Ala Asp Val Thr Ile Thr Val Asn Gly 25 Lys Val Val Ala Lys Pro Cys Thr Val Ser Thr Thr Asn Ala Thr Val 40 Asp Leu Gly Asp Leu Tyr Ser Phe Ser Leu Met Ser Ala Gly Ala Ala 55 Ser Ala Trp His Asp Val Ala Leu Glu Leu Thr Asn Cys Pro Val Gly 70 75 Thr Ser Arg Val Thr Ala Ser Phe Ser Gly Ala Ala Asp Ser Thr Gly 85 90 Tyr Tyr Lys Asn Gln Gly Thr Ala Gln Asn Ile Gln Leu Glu Leu Gln 105 Asp Asp Ser Gly Asn Thr Leu Asn Thr Gly Ala Thr Lys Thr Val Gln 120 Val Asp Asp Ser Ser Gln Ser Ala His Phe Pro Leu Gln Val Arg Ala 135 140 Leu Thr Val Asn Gly Gly Ala Thr Gln Gly Thr Ile Gln Ala Val Ile 150 Ser Ile Thr Tyr Thr Tyr Ser 165

<210> 367 <211> 300 <212> PRT <213> E. Coli

<400> 367

Met Lys Arg Val Ile Thr Leu Phe Ala Val Leu Leu Met Gly Trp Ser Val Asn Ala Trp Ser Phe Ala Cys Lys Thr Ala Asn Gly Thr Ala Ile Pro Ile Gly Gly Ser Ala Asn Val Tyr Val Asn Leu Ala Pro Val 45 Val Asn Val Gly Gln Asn Leu Val Val Asp Leu Ser Thr Gln Ile Phe 5.5 Cys His Asn Asp Tyr Pro Glu Thr Ile Thr Asp Tyr Val Thr Leu Gln 70 Arg Gly Ser Ala Tyr Gly Gly Val Leu Ser Asn Phe Ser Gly Thr Val 85 90 Lys Tyr Ser Gly Ser Ser Tyr Pro Phe Pro Thr Thr Ser Glu Thr Pro 100 105 Arg Val Val Tyr Asn Ser Arg Thr Asp Lys Pro Trp Pro Val Ala Leu 120 Tyr Leu Thr Pro Val Ser Ser Ala Gly Gly Val Ala Ile Lys Ala Gly

Ser Leu Ile Ala Val Leu Ile Leu Arg Gln Thr Asn Asn Tyr Asn Ser 150 155 Asp Asp Phe Gln Phe Val Trp Asn Ile Tyr Ala Asn Asn Asp Val Val 165 170 Val Pro Thr Gly Gly Cys Asp Val Ser Ala Arg Asp Val Thr Val Thr 185 Leu Pro Asp Tyr Pro Gly Ser Val Pro Ile Pro Leu Thr Val Tyr Cys 200 Ala Lys Ser Gln Asn Leu Gly Tyr Tyr Leu Ser Gly Thr Thr Ala Asp 215 220 Ala Gly Asn Ser Ile Phe Thr Asn Thr Ala Ser Phe Ser Pro Ala Gln 230 235 Gly Val Gly Val Gln Leu Thr Arg Asn Gly Thr Ile Ile Pro Ala Asn 24.5 250 Asn Thr Val Ser Leu Gly Ala Val Gly Thr Ser Ala Val Ser Leu Gly 260 265 Leu Thr Ala Asn Tyr Ala Arg Thr Gly Gly Gln Val Thr Ala Gly Asn 280 Val Gln Ser Ile Ile Gly Val Thr Phe Val Tyr Gln 295

<210> 368 <211> 521 <212> PRT <213> E. Coli

<400> 368 Met Leu Ser Lys Leu Pro Arg Arg Leu Arg Ser Phe Gln Thr Tyr Cys 10 Thr Ile Arg Val His Arg Gly Glu Asp Met Lys Ser Met Asp Lys Leu Thr Thr Gly Val Ala Tyr Gly Thr Ser Ala Gly Asn Ala Gly Phe Trp 40 Ala Leu Gln Leu Leu Asp Lys Val Thr Pro Ser Gln Trp Ala Ala Ile Gly Val Leu Gly Ser Leu Val Phe Gly Leu Leu Thr Tyr Leu Thr Asn 70 Leu Tyr Phe Lys Ile Lys Glu Asp Arg Arg Lys Ala Ala Arg Gly Glu 85 Ser Asn Asp Ser Arg Leu Thr Gly Cys Glu Arg Ser Pro Phe Glu Ser 100 105 Tyr Gly Asn Cys Ser Leu Thr Gly Gln Arg Thr Leu Arg Asn Phe Pro 120 Gly Cys Arg His Gly Pro Cys Arg Ser Cys Ala Gly Val Leu Gly Ser 135 Ser Gln Lys Glu Arg Pro Ala Ser Leu Pro Gly Ser Ser Arg Lys Ile 150 155 Val Arg Lys Ser Val Leu Ser Ala Ala Ser Val Leu Leu Asp Lys Ser 165 170 Cys Gln Ala Arg Ala Ser Ser Ser Ile Ser Met Asn Thr Lys Ile Arg 180 185 Tyr Gly Leu Ser Ala Ala Val Leu Ala Leu Ile Gly Ala Gly Ala Ser 195 200 Ala Pro Gln Ile Leu Asp Gln Phe Leu Asp Glu Lys Glu Gly Asn His 215 Thr Met Ala Tyr Arg Asp Gly Ser Gly Ile Trp Thr Ile Cys Arg Gly 230 235 Ala Thr Val Val Asp Gly Lys Thr Val Phe Pro Asn Met Lys Leu Ser 245 . 250



Lys Glu Lys Cys Asp Gln Val Asn Ala Ile Glu Arg Asp Lys Ala Leu 260 265 Ala Trp Val Glu Arg Asn Ile Lys Val Pro Leu Thr Glu Pro Gln Lys 280 Ala Gly Ile Ala Ser Phe Cys Pro Tyr Asn Ile Gly Pro Gly Lys Cys 295 Phe Pro Ser Thr Phe Tyr Lys Arg Leu Asn Ala Gly Asp Arg Lys Gly 310 315 Ala Cys Glu Ala Ile Arg Trp Trp Ile Lys Asp Gly Gly Arg Asp Cys 330 Arg Ile Arg Ser Asn Asn Cys Tyr Gly Gln Val Ile Arg Arg Asp Gln 345 Glu Ser Ala Leu Thr Cys Trp Gly Ile Glu Gln Ile Arg Tyr Ser Trp 360 Phe Phe Ser Cys Cys Gln Asp Leu Ser Ser Glu Met Ser Gly Ala Thr 375 380 Glu Asp Gly Lys Lys Asn Gly Arg Asn Val Met Leu Pro His Tyr His 390 395 Lys Arg Met Leu Asn Leu Leu Clu Leu Asn Arg Gly Glu Leu Pro 405 410 Val Met Arg Leu Leu Lys Met Arg Asn Arg Asn Leu Leu Lys Phe Leu 425 Pro Gly Leu Leu Ile Cys Leu Ile Val Leu Thr Ser Cys Val Pro Lys 440 445 Gln Lys Asn Met Pro Tyr Ala Leu Thr Gln Arg Ser Ile Pro Gln Ile 455 460 Leu Pro Leu Pro Ser Glu Ala Lys Gln Pro Lys Pro Pro Lys Glu Cys 470 475 Ser Pro Thr Cys Ser Glu Ile Leu Gln Gln Lys Leu Ser Phe Met Leu 485 490 495 Lys Leu Leu Thr Asn Ala Thr Ser Gln Glu Leu Val Asn Arg Ser Met 500 505 Asn Leu Glu Ile Lys Ser Ile Lys Cys 515

<210> 369 <211> 177 <212> PRT <213> E. Coli

<400> 369

 Met
 Asn
 Thr
 Lys
 Ile
 Arg
 Tyr
 Gly
 Leu
 Ser
 Ala
 Ala
 Ala
 Leu
 Ala
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 Ala
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 Ala
 Leu
 Ala
 Leu
 Ala
 Leu
 Ala
 Ala</th

Asp Gly Gly Arg Asp Cys Arg Ile Arg Ser Asn Asn Cys Tyr Gly Gln 145 150 155 160

Val Ile Arg Arg Asp Gln Glu Ser Ala Leu Thr Cys Trp Gly Ile Glu 165 170 175

Gln

<210> 370 <211> 103 <212> PRT <213> E. Coli

 <400>
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 Met Thr Gln Asp Tyr Glu Leu Val Val Lys Gly Val Arg Asn Phe Glu 1
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 Asn Lys Val Thr Val Thr Val Thr Val Ala Leu Gln Asp Lys Glu Arg Phe Asp 20
 25
 30
 61u Arg Phe Asp Asp Arg Val Glu Gly Ala Ala Met Asp Arg Val Glu Gly Ala 35
 40
 45

 Ala Leu Glu Phe Tyr Glu Ala Ala Ala Ala Ala Arg Arg Ser Val Arg Gln Val 50
 55
 60
 60

 Phe Leu Glu Val Ala Glu Lys Leu Ser Glu Lys Val Glu Ser Tyr Leu 65
 70
 75
 80

 Gln His Gln Tyr Ser Phe Lys Ile Glu Asn Pro Ala Asn Lys His Glu 85
 90
 95

 Arg Pro His His Lys Tyr Leu 100

<210> 371 <211> 96 <212> PRT <213> E. Coli

<400> 371

 Met
 Leu
 Ser
 Lys
 Leu
 Pro
 Arg
 Arg
 Leu
 Arg
 Ser
 Phe
 Gln
 Thr
 Tyr
 Cys

 Thr
 Ile
 Arg
 Val
 His
 Arg
 Gly
 Glu
 Asp
 Met
 Lys
 Ser
 Met
 Asp
 Lys
 Leu

 Thr
 Ile
 Arg
 Val
 Thr
 Ser
 Ala
 Gly
 Phe
 Trp

 Ala
 Leu
 Gly
 Asp
 Lys
 Val
 Thr
 Pro
 Ser
 Gln
 Trp
 Ala
 Ala
 Ile
 Ile
 Ile
 Ala
 Ile
 Ile

<210> 372 <211> 71 <212> PRT <213> E. Coli

<400> 372

Met Ser Asn Lys Met Thr Gly Leu Val Lys Trp Phe Asn Ala Asp Lys  $1 \\ 5 \\ 5 \\ 10 \\ 15 \\ 6$  Gly Phe Gly Phe Ile Ser Pro Val Asp Gly Ser Lys Asp Val Phe Val



20 25 30

His Phe Ser Ala Ile Gln Asn Asp Asn Tyr Arg Thr Leu Phe Glu Gly
35 40 45

Gln Lys Val Thr Phe Ser Ile Glu Ser Gly Ala Lys Gly Pro Ala Ala
50 55 60

Ala Asn Val Ile Ile Thr Asp
65 70

<210> 373 <211> 338 <212> PRT <213> E. Coli

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<210> 374 <211> 157 <212> PRT <213> E. Coli

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<210> 375 <211> 372 <212> PRT <213> E. Coli

<400> 375

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195
                            200
Val Phe Lys Asn Tyr Glu Leu Ile Ile Ser Ala Ala Arg Lys Leu Lys
                       215
Glu Gln Ser Asn Ile Lys Phe Leu Leu Thr Ile Ser Gly Thr Glu Asn
                    230 -
Ala Tyr Ala Lys Tyr Ile Ile Ser Leu Ala Glu Gly Leu Asp Asn Val
                245
His Phe Leu Gly Tyr Leu Asp Lys Glu Lys Ile Asp His Cys Tyr Asn
                               265
Ile Ser Asp Ile Val Cys Phe Pro Ser Arg Leu Glu Thr Trp Gly Leu
        275
                           280
Pro Leu Ser Glu Ala Lys Glu Arg Gly Lys Trp Val Leu Ala Ser Asp
                       295
                                            300
Phe Pro Phe Thr Arg Glu Thr Leu Gly Ser Tyr Glu Lys Lys Ala Phe
                   310
                                        315
Phe Asp Ser Asn Asn Asp Asp Met Leu Val Lys Leu Ile Ile Asp Phe
               325
                                   330
Lys Lys Gly Asn Leu Lys Lys Asp Ile Ser Asp Ala Asn Phe Ile Tyr
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Arg Asn Glu Asn Val Leu Val Gly Phe Asp Glu Leu Val Asn Phe Ile
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Thr Glu Glu His
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<210> 376 <211> 196 <212> PRT <213> E. Coli

<400> 376

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<210> 377 <211> 330 <212> PRT

195



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<210> 378 <211> 388 <212> PRT <213> E. Coli

<400> 378

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 Ile
 Tyr
 Leu
 Val
 Ile
 Ser
 Val
 Phe
 Leu
 Ile
 Thr
 Ala
 Phe
 Ile
 Cys
 Ile
 Cys
 Ile
 I



Leu Thr Phe Thr Leu Ser Cys Leu Leu Thr Glu Ser Val Leu Asp Leu 7.0 75 Asn Ile Arg Lys Val Asn Asn Ala Ile Tyr Ser Ile Pro Ser Lys Lys 85 90 Val His Asn Val Gly Leu Leu Val Ile Ser Phe Ser Met Ile Tyr Ile 100 105 Cys Met Arg Leu Ser Asn Tyr Gln Phe Gly Thr Ser Leu Leu Ser Tyr 115 120 Met Asn Leu Ile Arg Asp Ala Asp Val Glu Asp Thr Ser Arg Asn Phe 135 140 Ser Ala Tyr Met Gln Pro Ile Ile Leu Thr Thr Phe Ala Leu Phe Ile 150 155 Trp Ser Lys Lys Phe Thr Asn Thr Lys Val Ser Lys Thr Phe Thr Leu 165 170 Leu Val Phe Ile Val Phe Ile Phe Ala Ile Ile Leu Asn Thr Gly Lys 180 185 Gln Ile Val Phe Met Val Ile Ile Ser Tyr Ala Phe Ile Val Gly Val 195 200 Asn Arg Val Lys His Tyr Val Tyr Leu Ile Thr Ala Val Gly Val Leu 215 220 Phe Ser Leu Tyr Met Leu Phe Leu Arg Gly Leu Pro Gly Gly Met Ala 230 235 Tyr Tyr Leu Ser Met Tyr Leu Val Ser Pro Ile Ile Ala Phe Gln Glu 245 . 250 Phe Tyr Phe Gln Gln Val Ser Asn Ser Ala Ser Ser His Val Phe Trp 260 265 Phe Phe Glu Arg Leu Met Gly Leu Leu Thr Gly Gly Val Ser Met Ser 275 280 Leu His Lys Glu Phe Val Trp Val Gly Leu Pro Thr Asn Val Tyr Thr 295 300. Ala Phe Ser Asp Tyr Val Tyr Ile Ser Ala Glu Leu Ser Tyr Leu Met . 310 315 Met Val Ile His Gly Cys Ile Ser Gly Val Leu Trp Arg Leu Ser Arg 325 330 Asn Tyr Ile Ser Val Lys Ile Phe Tyr Ser Tyr Phe Ile Tyr Thr Phe 345 Ser Phe Ile Phe Tyr His Glu Ser Phe Met Thr Asn Ile Ser Ser Trp 360 Ile Gln Ile Thr Leu Cys Ile Ile Val Phe Ser Gln Phe Leu Lys Ala 375 Gln Lys Ile Lys 385

<210> 379 <211> 367 <212> PRT <213> E. Coli

<400> 379

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 Tyr
 Asp
 Tyr
 Ile
 Ile
 Val
 Gly
 Ser
 Gly
 Leu
 Phe
 Gly
 Ala
 Val
 Cys

 Ala
 Asn
 Glu
 Leu
 Lys
 Lys
 Lys
 Lys
 Val
 Leu
 Val
 Ile
 Glu
 Lys

 Arg
 Asn
 His
 Ile
 Gly
 Gly
 Asn
 Ala
 Tyr
 Thr
 Glu
 Asp
 Cys
 Glu
 Gly
 Ile

 Gln
 Ile
 His
 Lys
 Tyr
 Gly
 Ala
 His
 Ile
 Phe
 His
 Thr
 Asn
 Asp
 Lys
 Tyr

 Gln
 Ile
 His
 Lys
 Tyr
 Gly
 Ala
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 Ile
 Phe
 His
 Thr
 Asn
 Asp
 Lys
 Tyr

 Gln
 Ile
 His
 Lys
 Tyr
 Gly
 Ala
 His
 Ile
 Phe
 His
 Thr
 Asn
 Asp

Ser Pro Leu Ala Ile Tyr Lys Asp Lys Leu Phe Asn Leu Pro Phe Asn 85 Met Asn Thr Phe His Gln Met Trp Gly Val Lys Asp Pro Gln Glu Ala 100 105 Gln Asn Ile Ile Asn Ala Gln Lys Lys Lys Tyr Gly Asp Lys Val Pro 115 120 Glu Asn Leu Glu Glu Gln Ala Ile Ser Leu Val Gly Glu Asp Leu Tyr 135 Gln Ala Leu Ile Lys Gly Tyr Thr Glu Lys Gln Trp Gly Arg Ser Ala 150 155 Lys Glu Leu Pro Ala Phe Ile Ile Lys Arg Ile Pro Val Arg Phe Thr 165 170 Phe Asp Asn Asn Tyr Phe Ser Asp Arg Tyr Gln Gly Ile Pro Val Gly 185 Gly Tyr Thr Lys Leu Ile Glu Lys Met Leu Glu Gly Val Asp Val Lys 200 Leu Gly Ile Asp Phe Leu Lys Asp Lys Asp Ser Leu Ala Ser Lys Ala 215 220 His Arg Ile Ile Tyr Thr Gly Pro Ile Asp Gln Tyr Phe Asp Tyr Arg 230 235 Phe Gly Ala Leu Glu Tyr Arg Ser Leu Lys Phe Glu Thr Glu Arg His 245 250 Glu Phe Pro Asn Phe Gln Gly Asn Ala Val Ile Asn Phe Thr Asp Ala 265 Asn Val Pro Tyr Thr Arg Ile Ile Glu His Lys His Phe Asp Tyr Val 280 Glu Thr Lys His Thr Val Val Thr Lys Glu Tyr Pro Leu Glu Trp Lys 295 300 Val Gly Asp Glu Pro Tyr Tyr Pro Val Asn Asp Asn Lys Asn Met Glu 310 315 Leu Phe Lys Lys Tyr Arg Glu Leu Ala Ser Arg Glu Asp Lys Val Ile 325 330 Phe Gly Gly Arg Leu Ala Glu Tyr Lys Tyr Tyr Asp Met His Gln Val 345 Ile Ser Ala Ala Leu Tyr Gln Val Lys Asn Ile Met Ser Thr Asp 360

<210> 380 <211> 371 <212> PRT <213> E. Coli

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	130	l .				135					140				
145	, .				150					155					Leu 160
				165					170				_	175	Leu
			180					185					190		Ser
		195					200					205			Val
	210					Met 215					220				,
225					230	Glu				235					240
				245		Ile			250	•				255	
			260			Val		265					270		
		275				Leu	280					285		_	
	290					Val 295					300				
305					310	Arg				315					320
				325		Val		-	330					335	
			340			Ser		345					350		
Asp	Leu	Ala 355	Leu	Tyr	Lys	Ala	Lys 360	Lys	Ala	Gly	Arg	Asn 365	Arg	Thr	Glu
Val	Ala 370	Ala								•	-				

<210> 381 <211> 467 <212> PRT

<213> E. Coli

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Gly Thr Arg Leu Thr Asn Ser Arg His Gly Leu Ala Asp Asn Gly Gly
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Ala Trp Val Ser Tyr Phe Gly Gly Asn Phe Asn Gly Asp Asn Gly Thr
                       215
Ile Asn Tyr Asp Gln Asp Val Asn Gly Ile Met Val Gly Val Asp Thr
                   230
                                        235 -
Lys Ile Asp Gly Asn Asn Ala Lys Trp Ile Val Gly Ala Ala Ala Gly
                                    250
Phe Ala Lys Gly Asp Met Asn Asp Arg Ser Gly Gln Val Asp Gln Asp
                               265
Ser Gln Thr Ala Tyr Ile Tyr Ser Ser Ala His Phe Ala Asn Asn Val
                            280
Phe Val Asp Gly Ser Leu Ser Tyr Ser His Phe Asn Asn Asp Leu Ser
                       .295
                                            300
Ala Thr Met Ser Asn Gly Thr Tyr Val Asp Gly Ser Thr Asn Ser Asp
                   310
                                        315
Ala Trp Gly Phe Gly Leu Lys Ala Gly Tyr Asp Phe Lys Leu Gly Asp
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Ala Gly Tyr Val Thr Pro Tyr Gly Ser Val Ser Gly Leu Phe Gln Ser
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Gly Asp Asp Tyr Gln Leu Ser Asn Asp Met Lys Val Asp Gly Gln Ser
                           360
Tyr Asp Ser Met Arg Tyr Glu Leu Gly Val Asp Ala Gly Tyr Thr Phe
                      . 375
                                            380
Thr Tyr Ser Glu Asp Gln Ala Leu Thr Pro Tyr Phe Lys Leu Ala Tyr
                   390
                                        395
Val Tyr Asp Asp Ser Asn Asn Asp Asn Asp Val Asn Gly Asp Ser Ile
               405
                                    410
Asp Asn Gly Thr Glu Gly Ser Ala Val Arg Val Gly Leu Gly Thr Gln
           420
                               425
Phe Ser Phe Thr Lys Asn Phe Ser Ala Tyr Thr Asp Ala Asn Tyr Leu
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Tyr Thr Trp
465
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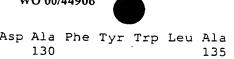
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<212> PRT

<213> E. Coli

<400> 382

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Asp Ala Phe Tyr Trp Leu Ala Trp Gln Asn Arg Ile Leu Glu Leu Arg 140 Asp Val Gln Leu Ile Gly His Asn Ser Tyr Glu Gln Ile Arg Ala Thr 150 155 Leu Leu Ser Met Ile Asp Trp Asn Glu Glu Leu Arg Ser Arg Ile Gly 165 170 Val Met Asn Tyr Ile His Gln Arg Thr Arg Ile Ser Arg Ser Val Val 180 185 190 Ala Glu Val Leu Ala Ala Leu Arg Lys Gly Gly Tyr Ile Glu Met Asn 200 Lys Gly Lys Leu Val Ala Ile Asn Arg Leu Pro Ser Glu Tyr 215

<210> 383 <211> 84 <212> PRT <213> E. Coli

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<210> 384 <211> 63 <212> PRT <213> E. Coli

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<210> 385 <211> 136 <212> PRT <213> E. Coli

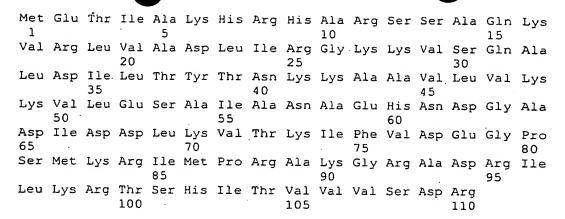
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35 Ala Arg Arg Ala Met Thr Arg Ala Val Lys Arg Gln Gly Lys Ile Trp 55 60 Ile Arg Val Phe Pro Asp Lys Pro Ile Thr Glu Lys Pro Leu Ala Val Arg Met Gly Lys Gly Lys Gly Asn Val Glu Tyr Trp Val Ala Leu Ile 90 Gln Pro Gly Lys Val Leu Tyr Glu Met Asp Gly Val Pro Glu Glu Leu 100 105 Ala Arg Glu Ala Phe Lys Leu Ala Ala Ala Lys Leu Pro Ile Lys Thr 115 120 Thr Phe Val Thr Lys Thr Val Met 130 135

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<210> 388 <211> 92 <212> PRT <213> E. Coli

<210> 389 <211> 273 <212> PRT <213> E. Coli

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 Pro
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 Pro
 Gly
 Arg
 Arg
 His
 Val

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 Lys
 Cys
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 Pro
 Thr
 Ser
 Pro
 Gly
 Lys
 Arg
 Arg
 His
 Val

 Val
 Lys
 Asn
 Pro
 Glu
 Leu
 His
 Lys
 Gly
 Lys
 Asn
 Asn
 Arg

 Leu
 Glu
 Lys
 Asn
 Ser
 Lys
 Ser
 Gly
 His
 Lys
 Asn
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 Gly
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 Arg
 <

130 Glu Met Lys Pro Gly Lys Gly Gly Gln Leu Ala Arg Ser Ala Gly Thr 150 155 Tyr Val Gln Ile Val Ala Arg Asp Gly Ala Tyr Val Thr Leu Arg Leu 165 170 Arg Ser Gly Glu Met Arg Lys Val Glu Ala Asp Cys Arg Ala Thr Leu 185 Gly Glu Val Gly Asn Ala Glu His Met Leu Arg Val Leu Gly Lys Ala 200 205 Gly Ala Ala Arg Trp Arg Gly Val Arg Pro Thr Val Arg Gly Thr Ala 215 220 Met Asn Pro Val Asp His Pro His Gly Gly Glu Gly Arg Asn Phe 230 235 Gly Lys His Pro Val Thr Pro Trp Gly Val Gln Thr Lys Gly Lys Lys 245 250 Thr Arg Ser Asn Lys Arg Thr Asp Lys Phe Ile Val Arg Arg Arg Ser 260 265 Lys

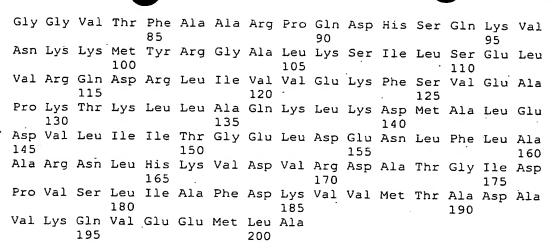
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<400> 390

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<210> 391 <211> 201 <212> PRT <213> E. Coli

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<210> 392 <211> 209 <212> PRT <213> E. Coli

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<210> 394 <211> 118 <212> PRT <213> E. Coli

<400> 394

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 Ala
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 His
 Lys
 Lys
 Ile
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 Arg
 Gly
 Ala
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<210> 395 <211> 65 <212> PRT <213> E. Coli

<400> 395

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 Arg
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 Leu
 Arg
 His
 Ile
 Arg
 His
 Leu
 Arg
 Pro
 Lys
 Ala

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 Asp
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 Gly
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 Val
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 Ala
 Cys
 Leu
 Pro
 Tyr

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 55
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<210> 396 <211> 180 <212> PRT <213> E. Coli

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PCT/US00/02200

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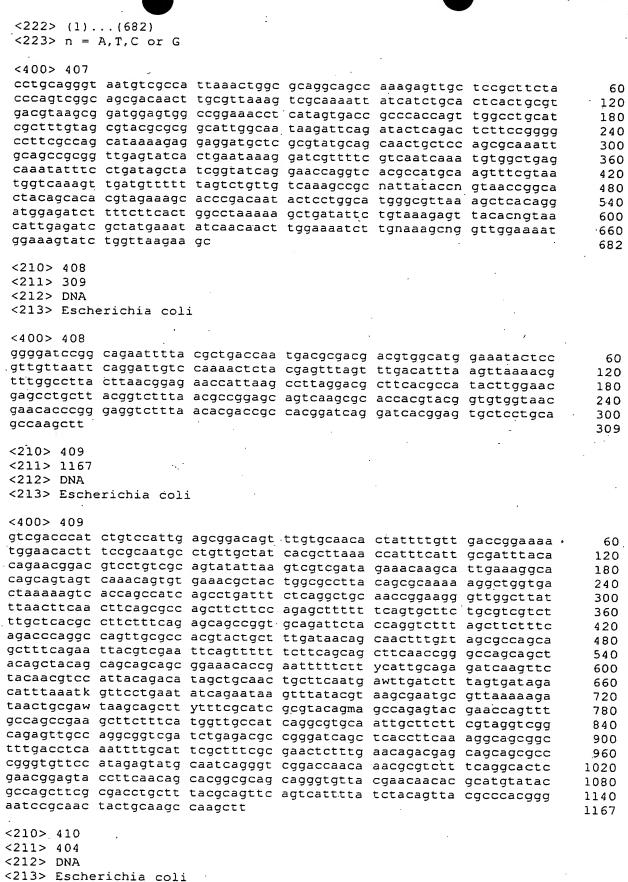
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aaattcattg	ttgaggacgc	gataatgaaa	acgttattac aacccagtat	caaacgttaa	tacqtctqaa	240 300

ttacgtctg tttttcttg agtgtatga	a tgccaaaag t ttccggaga c tgcagcaac	g atgtgcaca; g ccagttgat; c gaacagaaa;	tgaattcago ttgcagtcag ttcccggtaa	atttgtgct: g tgttcacago	t gctggctcgt t gttctgacag g acaatgcagg g gtcgataaag g taataaataa	420 480 540
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J	J	addycacac	grgcagettt	cacgg		465
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ctgccagcgg	acctgettte	agctgttcct	ggataccttt	atcaacggcc	gggatgtatt	180
cgccagggat	tacaccacct	ttaatgtcgt	tgatgaactc	gtagcctttc	gggtttgaac	240
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cadacacaca	tttaataact	atazatzaet	acglacttaa	ggtgcgtccg	gtgaaccagt	180
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	•	:				
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gcacaaattt	atcccacaac	tgttcttctg	tctcgacatg	cgccggatct	ttcacaatag	420
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.010	**					
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acaaggagg	caccadaacc	atcattgctg tgcgtccggt	gaaggaagga	gateettetg	ttcttataac	120
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cagagtagaa	tattaaattt	tatccgcgtg	atacatcaac	acaattaat	accelacete	300
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 agtcactcta agaggaggag aaattaggtt ggtattatag cttgtgcgcg ccatgattgg
                                                                      180
 cgcgcaattt aaacttagtg ctttacatcg ctattgtctt gatttctttg aattatttta
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 taaattaaaa aaacgactgt tatgtataag caaaggtccg aacgaaaaat acattccaaa
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 taaatgettg ettaaatete tatateette eeegaaaaat gacacataaa attgagatat
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 tccaaaaaga gatactacaa ataaagatgc ctttatttta ttatttctaa taaaaataga
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 agcaataaaa aataataaca atgatataaa totaatgttt ttaaatatat tgtottttat
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cgatggcgca gaaattgcgc catcaacgat cagtgataat taccaaccac aaacatcatg
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240
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cacategace tgateateaa aetgaatage ggeetgeteg taagttteet gggeggacae
                                                                     240
eggegeggea teggetttea teateegeae eattgggetg ggetgatagt tggaaacatg
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                                                                         180
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                                                                         240
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 ttagcgatag actgcattca gggcgaaagg aggtaagccg atgatttcag cgggacgctg
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                                                                        600
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catcgtggtg ctcttagtca taagcttccc cgcttactaa gactaccagg gcgggggaaa
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ttaaaaataa gatgttgctg ggtgcgcttt tgctggttac cagtgccgcc tgggccgcac
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cagccaccgc gggttcgacc aatacctcgg gaatttctaa gtatgagtta agtagtttca
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                                                                        480
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                                                                        540 /
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cagccaccgc gggttcgacc aatacctcgg gaatttctaa gtatgagtta agtagtttca
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60

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<210> 445 <211> 341 <212> DNA <213> Escherichia co	li				
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cgggcgctga ttcttcccad	: gcggttattt	tggcacacac	cagatccagc	aaggggtttt	180
caggatcgtt gagcagcaga	tgatctacca	gttccagcgc	ctgggtgtat	tgttcctcgt	240
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tttatgcgtc ataacttcac	gtatgtagca	cttttqcqat	tcaaaaaaga	ccattgctac	360 420
aacacgtaat tcattgcccc	caacattgaa	aacataatgc	ttatccagat	atttgaagtt	480
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cgtgktcccc agagccaccaccacaatgctttt tgagttatca	actccgtttt	atgttgcggg	tatttttccg	cagcatcttt	600
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                                                                          180
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 attgaatgtt gacgctatgt gtttatgagg gagaggtatt ttcagttgat ctggattgtt
                                                                          300
 aaattcatat aatgegeett tgeteatgaa tggatgeeag tatgtagtgg gaaattataa
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                                                                         120
 atgatatett eegatttate ttaategttt atggataaeg geaaaggget tegttttte
                                                                         180
 ctatacttat tcagcactca caaataaagg aacgccaatg aaaattatac tctgggctgt
                                                                         240
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                                                                         360
 gegeteetgt catgtgeatt getteatata ateaetggeg caaggagege egeaggegna
                                                                         420
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- (81) Designated States (national): AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, UZ, VN, YU, ZA, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW). Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE). OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

#### Published:

- With international search report.
- (88) Date of publication of the international search report: 1 February 2001

For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

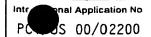
**A**3

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(54) Title: GENES IDENTIFIED AS REQUIRED FOR PROLIFERATION IN ESCHERICHIA COLI

(57) Abstract: The sequences of nucleic acids encoding proteins required for *E. coli* proliferation are disclosed. The nucleic acids can be used to express proteins or portions thereof, to obtain antibodies capable of specifically binding to the expressed proteins, and to use those expressed proteins as a screen to isolate candidate molecules for rational drug discovery programs. The nucleic acids can also be used to screen for homologous genes that are required for proliferation in microorganisms other than *E. coli*. The nucleic acids can also be used to design expression vectors and secretion vectors. The nucleic acids of the present invention can also be used in various assay systems to screen for proliferation required genes in other organisms as well as to screen for antimicrobial agents.





A. CLASSIFICATION OF SUBJECT MATTER IPC 7 C12N15/31 C12N C12N15/11

C DOCUMENTS CONSIDERED TO BE DELEVANT

C12N15/10

C07K14/245

According to International Patent Classification (IPC) or to both national classification and IPC

#### **B. FIELDS SEARCHED**

 $\begin{array}{ll} \text{Minimum documentation searched (classification system followed by classification symbols)} \\ IPC & 7 & C12N & C07K \end{array}$ 

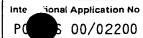
Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ, STRAND, BIOSIS, BIOTECHNOLOGY ABS, CHEM ABS Data

Category °	Citation of document, with indication, where appropriate, of the re	elevant passages	Relevant to claim No.
<b>A</b>	POST L E ET AL: "NUCLEOTIDE SEQ THE RIBOSOMAL PROTEIN GENE CLUST ADJACENT TO THE GENE FOR RNA POL SUBUNIT BETA IN ESCHERICHIA COLI PROCEEDINGS OF THE NATIONAL ACAD SCIENCES OF THE USA,US,NEW YORK, vol. 76, no. 4, 1 April 1979 (19 pages 1697-1701, XP000574791	ER YMERASE " EMY OF NY,	1
A	abstract WO 99 02673 A (DUGOURD DOMINIQUE 21 January 1999 (1999-01-21)		1
	page 7, line 25 -page 9, line 30 examples 2-6	-/	
X Furt	her documents are listed in the continuation of box C.	χ Patent family members are listed	in annex.
"A" docume	ent defining the general state of the art which is not defend to be of particular relevance	T later document published after the inte or priority date and not in conflict with cited to understand the principle or the invention	the application but
"L" docume which citation	ent which may throw doubts on priority claim(s) or is cited to establish the publication date of another n or other special reason (as specified)	"X" document of particular relevance; the cannot be considered novel or cannot involve an inventive step when the do "Y" document of particular relevance; the cannot be considered to involve an in-	be considered to current is taken alone lairned invention ventive step when the
other of the other	ent referring to an oral disclosure, use, exhibition or means ent published prior to the international filing date but nan the priority date claimed	document is combined with one or moments, such combination being obvior in the art.  *&* document member of the same patent	us to a person skilled
	actual completion of the international search  1 October 2000	Date of mailing of the international sea	00
Name and i	mailing address of the ISA  European Patent Office, P.B. 5818 Patentlaan 2  NL – 2280 HV Rijswijk  Tel. (+31–70) 340–2040, Tx. 31 651 epo nl,  Fax: (+31–70) 340–3016	Authorized officer  De Kok, A	

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		PU 5 00/02200
	ation) DOCUMENTS CONSIDERED TO BE RELEVANT	
Category °	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 98 21366 A (QBI ENTERPRISES LTD) 22 May 1998 (1998-05-22) page 8, line 9 - line 13 page 21, line 30 -page 25, line 2 page 26, line 11 -page 27, line 35	
X	BLATTNER F R ET AL: "THE COMPLETE GENOME SEQUENCE OF ESCHERICHIA COLI K-12" SCIENCE., vol. 277, 5 September 1997 (1997-09-05), pages 1453-1462, XP002923023 LANCASTER, PA., US ISSN: 0036-8075 the whole document, especially figure 3	8,9
<b>X</b>	VAN HEESWIJK W.C. ET AL.: "The genes of the glutamine synthetase adenylylation cascade are not regulated by nitrogen in Escherichia coli" MOLECULAR MICROBIOLOGY, vol. 9, 1993, pages 443-457, XP000926027 OXFORD GB nt4271-4371 of glnE sequence 100% identical with nt1-100 of seq.id.165 abstract	9
A	LEE N.G. ET AL.: "Molecular cloning and characterization of the nontypable Haemophilus influenzae-2019 rfaE gene required for lipopolysaccharide biosynthesis" INFECTION AND IMMUNITY., vol. 63, no. 3, 1995, pages 818-824, XP000953326 WASHINGTON., US ISSN: 0019-9567	8
A	the whole document  AUSTIN A.E. ET AL.: "Genetic analysis of lipopolysaccharide core biosynthesis by Escherichia coli k12 insertion mutagenesis of the RFA locus"  JOURNAL OF BACTERIOLOGY, vol. 172, 1990, pages 5312-5325, XP000926028  WASHINGTON US the whole document	8
	<del></del>	

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C 10:		/02200 .
C.(Continu Category °	tion) DOCUMENTS CONSIDERED TO BE RELEVANT  Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Т	VALVANO M.A. ET AL.: "The rfaE gene from Escherichia coli encodes a bifunctional protein involved in biosynthesis of the lipopolysaccharide core precursor ADP-L-glycero-D-manno-heptose." JOURNAL OF BACTERIOLOGY, vol. 182, January 2000 (2000-01), pages 488-497, XP000926030 WASHINGTON US the whole document	8
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Form PCT/ISA/210 (continuation of second sheet) (July 1992)



Box I	Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)
This Inte	ernational Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:
1.	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:
2. X	Claims Nos.: 11 13 34-45 47 48 50 51 53 55 57-63 65 67-93 95-105 107-110 because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:
	see FURTHER INFORMATION sheet PCT/ISA/210
3.	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).
Box II	Observations where unity of invention is lacking (Continuation of item 2 of first sheet)
This Int	ernational Searching Authority found multiple inventions in this international application, as follows:
	see additional sheet
1.	As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.
	As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment
-	of any additional fee.
<i>.</i>	
3. 1	As only some of the required additional search fees were timely paid by the applicant, this International Search Report
* LX_	covers only those claims for which fees were paid, specifically claims Nos.:
	1-10, 12, 14-33, 46, 49, 52, 54, 56, 64, 66, 94 and 106, all partially
4	No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:
Remari	k on Protest  The additional search fees were accompanied by the applicant's protest.
	χ No protest accompanied the payment of additional search fees.
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## FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

Continuation of Box I.2

Claims Nos.: 11 13 34-45 47 48 50 51 53 55 57-63 65 67-93 95-105 107-110

In view of the large number and also the wording of the claims presently on file, which render it difficult, if not impossible, to determine the matter for which protection is sought, the present application fails to comply with the clarity and conciseness requirements of Article 6 PCT (see also Rule 6.1(a) PCT) to such an extent that a meaningful search is impossible. Consequently, the search has been carried out for those parts of the application which do appear to be clear (and concise), namely the nucleic acid sequences as identified in claims 1 and 8 respectively, sequences related to said sequences as well as their use. This corresponds to the subject-matter of claims 1-10,12,14-33,46,49,52,54,56,64,66,94 and 106.

It should be noted that since claim 46 has been searched, the subject-matter of claims 35-45 has been searched restricted to the gene products of claim 46, i.e. for those gene products for which (additional) search fees have been paid

The applicant's attention is drawn to the fact that claims, or parts of claims, relating to inventions in respect of which no international search report has been established need not be the subject of an international preliminary examination (Rule 66.1(e) PCT). The applicant is advised that the EPO policy when acting as an International Preliminary Examining Authority is normally not to carry out a preliminary examination on matter which has not been searched. This is the case irrespective of whether or not the claims are amended following receipt of the search report or during any Chapter II procedure.

## FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-7, 12, 49, 52, 56, 66, all partially

#### Invention 1:

A purified or isolated nucleic acid sequence consisting of SEQ.ID. No.: 405, a vector comprising said sequence, a host comprising said vector, the use of said sequence for inhibiting cellular proliferation, a composition comprising said sequence, the use of said sequence for inhibiting the expression of a gene and the use of said nucleic acid sequence for identifying bacterial strains.

2. Claims: 1-7, 12, 49, 52, 56, 66, all partially

Inventions 2 to 81:

Idem as invention 1, but for SEQ.ID.NO's 406-485 respectively

3. Claims: Claims 8-10,12,14-33,46,54,64,66,94 and 106, all partially:

### Invention 82:

A purified or isolated nucleic acid consisting of SEQ.ID.No.: 82, a vector comprising said nucleic acid sequence, a host comprising said vector, a polypeptide encoded by said nucleic acid sequence and having the sequence of SEQ.ID.No.: 243, an antibody binding said polypeptide, a method for producing said polypeptide, a method for identifying compounds influencing the activity of said polypeptide, a method for identifying compounds influencing the level of said polypeptide, a method for inhibiting the expresion of said nucleic acid, the use of said nucleic acid sequence for identifying bacterial strains and the use of said nucleic acid sequence for identifying proliferation inhibitors.

4. Claims: Claims 8-10,12,14-33,46,54,64,66,94 and 106, all partially:

Inventions 83 to 242:

Idem as invention 82, but for SEQ.ID.No's 83-88, 90-242 (and their corresponding polypeptide sequences, see Table II) respectively.

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nation on patent family members

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WO 9821366	Ą	22-05-1998	AU EP	5442198 A 0960212 A	03-06-1998 01-12-1999	

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BNSDOCID: <WO\_\_\_\_\_0044906A3\_I\_>

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